SMART 2

Colorimeter

OPERATOR'S MANUAL

LaMotte

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GENERAL INFORMATION

PACKAGING & DELIVERY

Experienced packaging personnel at LaMotte Company assure adequate protection against normal hazards encountered in transportation of shipments. After the product leaves the manufacturer, all responsibility for its safe delivery is assured by the transportation company. Damage claims must be filed immediately with the transportation company to receive compensation for damaged goods.

Should it be necessary to return the instrument for repair or servicing, pack instrument carefully in suitable container with adequate packing material. A return authorization number must be obtained from LaMotte Company by calling 1-800-344-3100. Attach a letter with the authorization number to the shipping carton which describes the kind of trouble experienced. This valuable information will enable the service department to make the required repairs more efficiently.

GENERAL PRECAUTIONS

Before attempting to set up or operate this instrument it is important to read the instruction manual. Failure to do so could result in personal injury or damage to the equipment.

The SMART2 Colorimeter should not be stored or used in a wet or corrosive environment. Care should be taken to prevent water or reagent chemicals from wet colorimeter tubes from entering the colorimeter chamber.

NEVER PUT WET TUBES IN COLORIMETER.

SAFETY PRECAUTIONS

Read the labels on all LaMotte reagent containers prior to use. Some containers include precautionary notices and first aid information. Certain reagents are considered hazardous substances and are designated with a * in the instruction manual. Material Safety Data Sheets (MSDS) are supplied for these reagents. Read the accompanying MSDS before using these reagents. Additional emergency information for all LaMotte reagents is available 24 hours a day from the Poison Control Center listed in the front of the phone book. Be prepared to supply the name and four-digit LaMotte code number found on the container label or at the top of the MSDS. LaMotte reagents are registered with a computerized poison control information system available to all local poison control centers.

Keep equipment and reagent chemicals out of the reach of young children.

Protect Yourself and Equipment: Use Proper Analytical Techniques

LIMITS OF LIABILITY

Under no circumstances shall LaMotte Company be liable for loss of life, property, profits, or other damages incurred through the use or misuse of its products.

LIMITED WARRANTY

This instrument is guaranteed to be free from defects in material and workmanship for a period of two (2) years from original purchase date. In the event that a defect is found during the warranty time frame, LaMotte Company agrees that it will be repaired or replaced without charge except for the transporation costs. This guarantee does not cover batteries.

This product can not be returned without a return authorization number from Lamotte Company. For warranty support or a Return Authorization Number, contact LaMotte Company at 1-800-344-3100 or tech @ lamotte.com.

LIMITATIONS

This guarantee is void under the following circumstances:

- Damage due to operator negligence, misuse, accident or improper application.
- Damage or alterations from attempted repairs by an unauthorized (non-LaMotte) service.
- Damage due to improper power source, AC adapter or battery.
- Damage caused by acts of God or natural disaster.
- Damage occurred while in transit with a shipping carrier.

LaMotte Company will service and repair out-of warranty products at a nominal charge.

SPECIFICATIONS

INSTRUMENT TYPE: Colorimeter

Readout	Graphical 4 line, 16 character per line LCD
Wavelengths	430nm, 520 nm, 570 nm, 620 nm
Wavelength Accuracy	± 2
Readable Resolution	Determined by reagent system
Wavelength Bandwidth	10 typical
Photometric Range	–2 to + 2AU
Photometric Precision	± 0.001AU at 1.0AU
Photometric Accuracy	± 0.005AU at 1.0AU
Sample Chamber	Accepts 25 mm diameter flat-bottomed test tubes, 10 mm square cuvettes, 16 mm COD test tubes
Light Sources	4 LEDs
Detectors	4 silicon photodiodes with integrated interference filters
Modes	Absorbance, pre-programmed tests
Pre-Programmed Tests	YES, with automatic wavelength selection
User Defined Tests	Up to 10 user tests can be input
RS232 Port	8 pin mini-DIN, 9600b, 8, 1, n
Power Requirements	*Battery Operation:* 9 volt alkaline, Line Operation: 110/ AC; 50/60 Hz with adapter, 6V 500 mA DC
Dimensions (LxWxH)	8.5 x 16.2 x 6.7 cm, 3.4 x 6 .4 x 2.6 inches
Weight	312 g, 11 oz (meter only)
Data Logger	350 test results stored for download to a PC

STATISTICAL AND TECHNICAL DEFINITIONS RELATED TO PRODUCT SPECIFICATIONS

Method Detection Limit (MDL): "The method detection limit (MDL) is defined as the minimum concentration of a substance that can be measured and reported with 99% confidence that the analyte concentration is greater than zero and is determined from analysis of a sample in a given matrix containing the analyte."[1] Note that, "As Dr. William Horwitz once stated, 'In almost all cases when dealing with a limit of detection or limit of determination, the primary purpose of determining that limit is to stay away from it.'"[2]

1. CFR 40, part 136, appendix B
2. Statistics in Analytical Chemistry: Part 7 – A Review, D. Coleman and L Vanatta, American Laboratory, Sept 2003, P. 31.

Precision: Precision is the numerical agreement between two or more measurements.[3] The precision can be reported as a range for a measurement (difference between the min and max). It can also be reported as the standard deviation or the relative standard deviation. It is a measure of how close together the measurements are, not how close they are to the correct or true value. *The precision can be very good and the accuracy very bad. This is a useful measure of the performance of a test method.*

3. Skoog, D.A., West, D. M., *Fundamental of Analytical Chemistry,* 2nd ed., Holt Rinehart and Winston, Inc, 1969, p. 26.

Accuracy: Accuracy is the nearness of a measurement to the accepted or true value.[4] The accuracy can be expressed as a range, about the true value, in which a measurement occurs (i.e. ±0.5 ppm). It can also be expressed as the % recovery of a know amount of analyte in a determination of the analyte (i.e. 103.5 %). *This is a useful measure and what most customers are interested in when they want to know about the performance of a test method.*

4. Skoog D.A., West D. M., *Fundamental of Analytical Chemistry,* 2nd ed., Holt Rinehart and Winston, Inc, 1969, p. 26.

Resolution: Resolution is the smallest discernible difference between any two measurements that can be made.[5] For meters this is usually how many decimal places are displayed. (i.e. 0.01). For titrations and various comparators it is the smallest interval the device is calibrated or marked to (i.e. 1 drop = 10 ppm, 0.2 ppm for a DRT, or ±half a unit difference for an octaslide or color chart). Note that the resolution many change with concentration or range. In some cases the resolution may be less than the smallest interval, if it is possible to make a reading that falls between calibration marks. This is often done with various comparators. *One caveat is, that resolution has very little relationship to accuracy or precision. The resolution will always be less than the accuracy or precision but it is not a statistical measure of how well a method of analysis works. The resolution can be very very good and the accuracy and precision can be very, very bad! This is not a useful measure of the performance of a test method.*

5. Statistics in Analytical Chemistry: Part 7 – A Review, D. Coleman and L Vanatta, American Laboratory, Sept 2003, P. 34.

Sensitivity: Sensitivity is the resolution based on how this term is used in LaMotte catalogs. This term is not listed in any of the references. Sometimes it is used for detection limit. It is a confusing term and should be avoided.

Repeatability: Repeatability is the within-run precision.[6] A run is a single data set, from set up to clean up. Generally, one run occurs on one day. However, for meter calibrations, a single calibration is considered a single run or data set, even though it may take 2 or 3 days.

6. Jeffery G. H., Basset J., Mendham J., Denney R. C., *Vogel's Textbook of Quantitative Chemical Analysis*, 5th ed., Longman Scientific & Technical, 1989, p. 130.

Reproducibility: Reproducibility is the between-run precision.[7]

7. Jeffery G. H., Basset J., Mendham J., Denney R. C., *Vogel's Textbook of Quantitative Chemical Analysis*, 5th ed., Longman Scientific & Technical, 1989, p. 130.

CONTENTS AND ACCESSORIES

CONTENTS

SMART2 Colorimeter

Test Tubes, with Caps

Power Supply, 110/220V

SMART2 Colorimeter Quick Start Guide

SMART2 Colorimeter Manual

ACCESSORIES

COD Adapter	Code 5-0087
UDV Adapter	Code 5-0086
Small Field Carrying Case	Code 1919-GCS150
Large Field Carrying Case	Code 1919-BCS440
SMARTLink2 Program & Interface Cable (3.5 disk)	Code 1912-3
SMARTLink2 Program & Interface Cable (CD)	Code 1912-CD

EPA COMPLIANCE

The SMART2 Colorimeter is an EPA-Accepted instrument. EPA-Accepted means that the instrument meets the requirements for instrumentation as found in test procedures that are approved for the National Primary Drinking Water Regulations (NPDWR) or National Pollutant Discharge Elimination System (NPDES) compliance monitoring programs. EPA-Accepted instruments may be used with approved test procedures without additional approval.

CE COMPLIANCE

The SMART2 Colorimeter has earned the European CE Mark of Compliance for electromagnetic compatibility and safety.

DECLARATION OF CONFORMITY

Standards to which Conformity Declared:	EN61326:1998, IEC61326:1997, IEC61000-4-2:1995, IEC61000-4-3:1995 IEC61000-4-4:1995, IEC61000-4-5:1995 IEC61000-4-6:1996, IEC61000-4-11:1994, EN61000-3-2:1995, EN61000-3-3:1994-12, EN55011/CISPR11, FCCCFR47 Part 15, EN61558
Manufacturer's Name:	LaMotte Company
Manufacturer's Address:	802 Washington Avenue PO Box 329 Chestertown, MD 21620
Type of Equipment:	Colorimeter
Model Name:	SMART 2
Year of Manufacture:	2001
Testing Performed By:	Windermere 2000 Windermere Court Annapolis, MD 21401

I, the undersigned, hereby declare that the equipment specified above conforms to the above Directive and Standards

Chestertown, Maryland
Place

1/15/02
Date

Scott H. Steffen
Signature

Scott H. Steffen
Name

VP New Products & Quality
Position

CHEMICAL TESTING

WATER SAMPLING FOR CHEMICAL ANALYSIS

Taking Representative Samples

The underlying factor to be considered for any type of water sampling is whether or not the sample is truly representative of the source. To properly collect a representative sample:

- Sample as frequently as possible.
- Collect a large sample or at least enough to conduct whatever tests are necessary.
- Make a composite sample for the same sampling area.
- Handle the sample in such a way as to prevent deterioration or contamination before the analysis is performed.
- Perform analysis for dissolved gases such as dissolved oxygen, carbon dioxide, and hydrogen sulfide immediately at the site of sampling. These factors, as well as samples for pH, cannot be stored for later examination.
- Make a list of conditions or observations which may affect the sample. Other considerations for taking representative samples are dependent upon the source of the sample. Taking samples from surface waters involves different considerations than taking samples from impounded and sub-surface waters.

Sampling of Open Water Systems

Surface waters, such as those found in streams and rivers, are usually well mixed. The sample should be taken downstream from any tributary, industrial or sewage pollution source. For comparison purposes samples may be taken upstream and at the source of the pollution before mixing.

In ponds, lakes, and reservoirs with restricted flow, it is necessary to collect a number of samples in a cross section of the body of water, and where possible composite samples should be made to ensure representative samples.

To collect samples from surface waters, select a suitable plastic container with a tight fitting screw cap. Rinse the container several times with the sample to be tested, then immerse the container below the surface until it is filled to overflowing and replace the cap. If the sample is not to be tested immediately, pour a small part of the sample out and reseal. This will allow for any expansion. Any condition which might affect the sample should be listed.

Sub-surface sampling is required to obtain a vertical profile of streams, lakes, ponds, and reservoirs at specific depths. This type of sampling requires more sophisticated sampling equipment.

For dissolved oxygen studies, or for tests requiring small sample sizes, a Water Sampler (LaMotte Code 1060) will serve as a subsurface or in-depth sampler.

This weighted device is lowered to the sampling depth and allowed to rest at this depth for a few minutes. The water percolates into the sample chamber displacing the air which bubbles to the surface. When the bubbles cease to rise, the device has flushed itself approximately five times and it may be raised to the surface for examination. The inner chamber of the sampling device is lifted out and portions of the water sample are carefully dispensed for subsequent chemical analysis.

A Snap-Plunger Water Sampler (LaMotte Code 1077) is another "in-depth" sampling device which is designed to collect large samples which can be used for a multitude of tests. Basically, this collection apparatus is a hollow cylinder with a spring loaded plunger attached to each end. The device is cocked above the surface of the water and lowered to the desired depth. A weighted messenger is sent down the calibrated line to trip the closing mechanism and the plungers seal the sample from mixing with intermediate layers as it is brought to the surface. A special drain outlet is provided to draw off samples for chemical analysis.

▪ Sampling of Closed System

To obtain representative samples from confined water systems, such as pipe lines, tanks, vats, filters, water softeners, evaporators and condensers, different considerations are required because of chemical changes which occur between the inlet and outlet water. One must have a basic understanding of the type of chemical changes which occur for the type of equipment used. Also, consideration should be given to the rate of passage and retaining time for the process water.

Temperature changes play an important part in deciding exactly what test should be performed. Process water should be allowed to come to room temperature, 20–25°C, before conducting any tests.

When drawing off samples from an outlet pipe such as a tap, allow sample to run for several minutes, rinsing the container several times before taking the final sample. Avoid splashing and introduction of any contaminating material.

FILTRATION

When testing natural waters that contain significant turbidity due to suspended solids and algae, filtration is an option. Reagent systems, whether EPA, Standard Methods, LaMotte or any others, will generally only determine dissolved constituents. Both EPA and Standard Methods suggest filtration through a 0.45 micron filter membrane, to remove turbidity, for the determination of dissolved constituents.** To test for total constituents, organically bound and suspended or colloidal materials, a rigorous high temperature acid digestion is necessary.

**LaMotte offers a filtering apparatus: syringe assembly (Code 1050) and membrane filters, 0.45 micron, (Code 1103).

AN INTRODUCTION TO COLORIMETRIC ANALYSIS

Most test substances in water are colorless and undetectable to the human eye. To test for their presence we must find a way to "see" them. The SMART2 Colorimeter can be used to measure any test substance that is itself colored or can be reacted to produce a color. In fact a simple definition of colorimetry is "the measurement of color" and a colorimetric method is "any technique used to evaluate an unknown color in reference to known colors". In a colorimetric chemical test the intensity of the color from the reaction must be proportional to the concentration of the substance being tested. Some reactions have limitations or variances inherent to them that may give misleading results. Many such interferences are discussed with each particular test instruction. In the most basic colorimetric method the reacted test sample is visually compared to a known color standard. However, accurate and reproducible results are limited by the eyesight of the analyst, inconsistencies in the light sources, and the fading of color standards.

To avoid these sources of error, a colorimeter can be used to photoelectrically measure the amount of colored light absorbed by a colored sample in reference to a colorless sample (blank).

White light is made up of many different colors or wavelengths of light. A colored sample typically absorbs only one color or one band of wavelengths from the white light. Only a small difference would be measured between white light before it passes through a colored sample versus after it passes through a colored sample. The reason for this is that the one color absorbed by the sample is only a small portion of the total amount of light passing through the sample. However, if we could select only that one color or band of wavelengths of light to which the test sample is most sensitive, we would see a large difference between the light before it passes through the sample and after it passes through the sample.

The SMART2 Colorimeter passes one of four colored light beams through one of four optical filters which transmits only one particular color or band of wavelengths of light to the photodectector where it is measured. The difference in the amount of colored light transmitted by a colored sample is a measurement of the amount of colored light absorbed by the sample. In most colorimetric tests the amount of colored light absorbed is directly proportional to the concentration of the test factor producing the color and the path length through the sample. However, for some tests the amount of colored light absorbed is inversely proportional to the concentration.

The choice of the correct wavelength for testing is important. It is interesting to note that the wavelength that gives the most sensitivity (lower detection limit) for a test factor is the complementary color of the test sample. For example the Nitrate-Nitrogen test produces a pink color proportional to the nitrate concentration in the sample (the greater the nitrate concentration, the darker the pink color). A wavelength in the green region should be selected to analyze this sample since a pinkish-red solution absorbs mostly green light.

REAGENT BLANK

Some tests will provide greater accuracy if a reagent blank is determined to compensate for any color or turbidity resulting from the reagents themselves. A reagent blank is performed by running the test procedure on 10 mL of demineralized water. Use sample water to SCAN BLANK. Insert the reagent blank in the colorimeter chamber and select SCAN SAMPLE. Note result of reagent blank. Perform the tests on the sample water as described. Subtract results of reagent blank from all subsequent test results. NOTE: Some tests require a reagent blank to be used to SCAN BLANK.

COLORIMETER TUBES

Colorimeter tubes which have been scratched through excessive use should be discarded and replaced with new ones. Dirty tubes should be cleaned on both the inside and outside. Fingerprints on the exterior of the tubes can cause excessive light scattering and result in errors. Handle the tubes carefully, making sure the bottom half of the tube is not handled.

LaMotte Company makes every effort to provide high quality colorimeter tubes. However, wall thicknesses and diameter of tubes may still vary slightly. This may lead to slight variations in results (e.g. if a tube is turned while in the sample chamber, the reading will likely change slightly). To eliminate this error put the tubes into the sample chamber with the same orientation every time.

The tubes that are included with the colorimeter have an index mark to facilitate this. If possible, use the same tube to SCAN BLANK and SCAN SAMPLE.

METER CARE

The optical system of the SMART2 must be kept clean and dry for optimal performance. Dry the colorimeter tubes before placing them in the chamber to avoid introducing moisture. For best results store the instrument in a area that is dry and free from aggressive chemical vapors.

SELECTING AN APPROPRIATE WAVELENGTH

The most appropriate wavelength to use when creating a calibration curve is usually the one which gives the greatest change from the lowest reacted standard concentration to the highest reacted standard concentration. However, the absorbance of the highest reacted standard concentration should never be greater than 2.0 absorbance units. Scan the lowest and highest reacted standards at different wavelengths using the absorbance mode to find the wavelength which gives the greatest change in absorbance without exceeding 2.0 absorbance units. Use this wavelength to create a calibration curve.

Below is a list of suggested wavelengths for the color of the reacted samples. Use these as a starting point.

Sample Color	Wavelength Range
Yellow	430
Pink	520
Red	570
Green and Blue	620

CALIBRATION

As with all pre-calibrated meters, it is highly recommended, even if not required by regulations, that the user periodically verify the performance of the meter by running standards with a predetermined concentration. Results outside of specification are an indication that the meter needs to be adjusted. This can be done following the user calibration described on page 31. If the user calibration fails to properly adjust the meter then the meter should be returned to LaMotte Company for recalibration. (See page 5).

CALIBRATION CURVES

The Smart2 Colorimeter contains tests for the LaMotte reagent systems (see Page 49). The first step in using a non-LaMotte reagent system with your Smart2 Colorimeter is to create a calibration curve for the reagent system. To create a calibration curve, prepare standard solutions of the test factor and use the reagent system to test the standard solutions with the Smart2 Colorimeter. Select a wavelength for the test as described above.

Plot the results (in ABS or %Transmittance) versus concentration to create a calibration curve. The calibration curve may then be used to identify the concentration of an unknown sample by testing the unknown, reading Absorbance or %T, and finding the corresponding concentration from the curve. The linear range of the reagent system can be determined and this information can be used to input a User Test into the Smart2 Colorimeter (see EDIT USER TESTS, page 36).

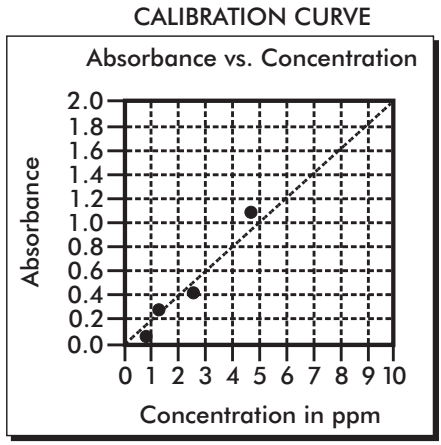
PROCEDURE

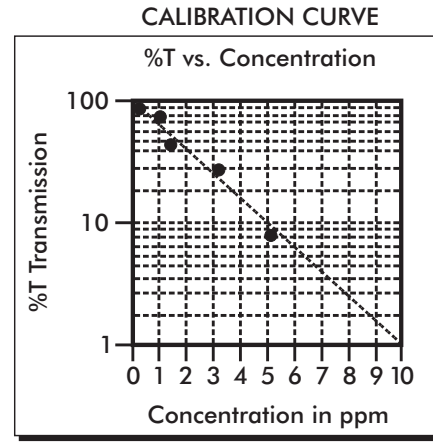
1. Prepare 5 or 6 standard solutions of the factor being tested. The concentration of these standards should be evenly distributed throughout the range of the reagent system, and should include a 0 ppm standard (distilled water). For instance, the solutions could measure 0, 10%, 30%, 50%, 70%, and 90% of the system's maximum range.
2. Turn on the Smart2 Colorimeter. Select the appropriate wavelength from the absorbance mode. Be sure to select the appropriate wavelength for the color produced by the reagent system.

3. Use the unreacted 0 ppm standard to standardize the colorimeter by using it to scan blank.
4. Following the individual reagent system instructions, react each standard solution beginning with 0 ppm. Continue with standards in increasing concentration. Record the reading and the standard solution concentration on a chart. Readings can be recorded as percent transmittance (%T) or absorbance (A).
5. Plot results on graph paper or computer using any available plotting program. If results are as %T versus concentration, semilog graph paper must be used. Plot the standard solution concentrations on the horizontal, linear axis, and the %T on the vertical, logarithmic axis. If results are as absorbance versus standard solution concentration, simple linear graph paper can be used. Plot the standard solution concentration on the horizontal axis, and the absorbance on the vertical axis.
6. After plotting the results, draw a line, or curve, of best fit through the plotted points. The best fit may not connect the points. There should be approximately an equal number of points above the curve as below the curve. Some reagent systems will produce a straight line, while others produce a curve. Many computer spreadsheet programs can produce the curve of best fit by regression analysis of the standard solution data.

NOTE: Only reagent systems which produce a straight line can be used for a User Test.

A sample of each type of graph appears below:





▪ PREPARING DILUTE STANDARD SOLUTIONS

Standard solutions should be prepared to create a calibration curve. Standard solutions can be prepared by diluting a known concentrated standard by specified amounts. A chart or computer spreadsheet can be created to determine the proper dilutions. Use volumetric flasks and volumetric pipets for all dilutions.

1. In Column A – Record the maximum concentration of test as determined by the range and path length.
2. In Column B – Record the percent of the maximum concentration the standard solution will be.
3. In Column C – Calculate the final concentration of the diluted standard solutions by multiplying the maximum concentration (In Column A) by the % of maximum concentration divided by 100. ($C = A \times {}^{B}/_{100}$).
4. In Column D – Record the final volume of the diluted sample (i.e. volume of volumetric flask).
5. In Column E – Record the concentration of the original standard.
6. In Column F – Calculate the milliliters of original standard required ($C \times {}^{D}/_{E} = F$).

A sample chart appears below:

A	B	$C = A \times {}^{B}/_{100}$	D	E	$F = C \times {}^{D}/_{E}$
Maximum concentration of test	% of Maximum concentration	Final concentration of Diluted Standard	Volume of Standard	Concentration of Original Standard	mL of Original Standard Required
10.0 ppm	90	9.0 ppm	100 mL	1000 ppm	0.90 mL
10.0 ppm	70	7.0 ppm	100 mL	1000 ppm	0.70 mL
10.0 ppm	50	5.0 ppm	100 mL	1000 ppm	0.50 mL
10.0 ppm	30	3.0 ppm	100 mL	1000 ppm	0.30 mL
10.0 ppm	10	1.0 ppm	100 mL	1000 ppm	0.10 mL
10.0 ppm	0	0 ppm	100 mL	1000 ppm	0 mL

■ STANDARD ADDITIONS

A common method to check the accuracy and precision of a test is by standard additions. In this method a sample is tested to determine the concentration of the test substance. A second sample is then "spiked" by the addition of a known quantity of the test substance. The second sample is then tested. The determined concentration of the spiked sample should equal the concentration of the first plus the amount added with the spike. The procedure can be repeated with larger and larger "spikes." If the determined concentrations do not equal the concentration of the sample plus that added with the "spike", then an interference may exist.

For example, a 10.0 mL water sample was determined to contain 0.3 ppm iron. To a second 10.0 mL sample, 0.1 mL of 50 ppm iron standard was added. The

concentration of iron due to the "spike" was (0.10 mL x 50 ppm)/10.0 mL = 0.50 ppm. The concentration of iron determined in the spiked sample should be 0.3 + 0.5 = 0.8 ppm iron. (Note: any error due to the increased volume from the "spike" is negligible).

LaMotte offers a line of calibration standards which can be used to generate calibration curves and perform standard additions.

SAMPLE DILUTION TECHNIQUES & VOLUMETRIC MEASUREMENTS

If a test result using the Smart2 Colorimeter gives an over range message then the the sample must be diluted. The test should be repeated on the diluted sample to obtain a reading which is in the concentration range for the test. (Note: This is not true for colorimetric determination of pH.)

Example:

Measure 5 mL of the water sample into a graduated cylinder. Add demineralized water until the cylinder is filled to the 10 mL line. The sample has been diluted by one-half, and the dilution factor is therefore 2. Perform the test procedure, then multiply the resulting concentration by 2 to obtain the test result.

The following table gives quick reference guidelines on dilutions of various proportions. All dilutions are based on a 10 mL volume, so several dilutions will require small volumes of the water sample. Graduated pipets should be used for all dilutions.

Size of Sample	Deionized Water to Bring Volume to 10 mL	Multiplication Factor
10 mL	0 mL	1
5 mL	5 mL	2
2.5 mL	7.5 mL	4
1 mL	9 mL	10
0.5 mL	9.5 mL	20

If the above glassware is not available, dilutions can be made with the colorimeter tube. Fill the tube to the 10 mL line with the sample then transfer it to another container. Add 10 mL volumes of demineralized water to the container and mix. Transfer back 10 mL of the diluted sample to the tube and follow the test procedure. Continue diluting and testing until a reading, which is in the concentration range for the test, is obtained. Be sure to multiply the concentration found by the dilution factor (the number of total 10 mL volumes used).

Example:

10 mL of sample is diluted with three 10 mL volumes of demineralized water; the dilution factor is four.

INTERFERENCES

LaMotte reagent systems are designed to minimize most common interferences. Each individual test instruction discusses interferences unique to that test. Be aware of possible interferences in the water being tested.

The reagent systems also contain buffers to adjust the water sample to the ideal pH for the reaction. It is possible that the buffer capacity of the water sample may exceed the buffer capacity of the reagent system and the ideal pH will not be obtained. If this is suspected, measure the pH of a reacted distilled water reagent blank using a pH meter. This is the ideal pH for the test. Measure the pH of a reacted water sample using the pH meter. If the pH is significantly different from the ideal value, the pH of the sample should be adjusted before testing.

Interferences due to high concentration of the substance being tested, can be overcome by sample dilution (see page 16)

STRAY LIGHT INTERFERENCE

When scanning samples in 16 mm tubes, such as COD, the sample chamber lid can not be closed. The COD adapter minimizes stray light. To further reduce stray light interference, do not scan sample in direct sunlight.

OPERATION OF THE SMART2 COLORIMETER

OVERVIEW

The SMART2 Colorimeter is a portable, microprocessor controlled, direct reading colorimeter. It uses a graphical 4 line, 16 character liquid crystal display for graphical and numerical messages. The operation is controlled with the driven software in response to selections shown on the d

The test aMotte tests (not all ilable at present) e LaMotte tests are pr LaMotte reagent s displays the resu in units of User Tests" may b al calibrati y be arranged in any of 3 sequences. These sequence tless number of t esting needs.

The opti ored LEDs. Each g silicon p rated interferenc filters sel nt from the corresponding for the colorime microprocessor autom the correct L ation for a test.

A RS-23 of the colorimeter, a tware, allows the rfaced with an IBM compatible personal computer for real time data acquisition and data storage. This port also allows an interface with a RS-232 serial printer.

Due to its portability, alternate power sources, uction, the SMART2 Colorimeter is ideal for lab and fie

POWER SOURCE

The SMART2 Colorimeter uses a 500 mA to the Parts List for the code number for the corr

U E OF ANY AC ADAPTER OTHER ECTED FO USE W COLORI AGE THE ME R A RRANTY. Do not use the adapter sold wi

To use the AC adapter into the small hole on th nto an appropriate wall socket or

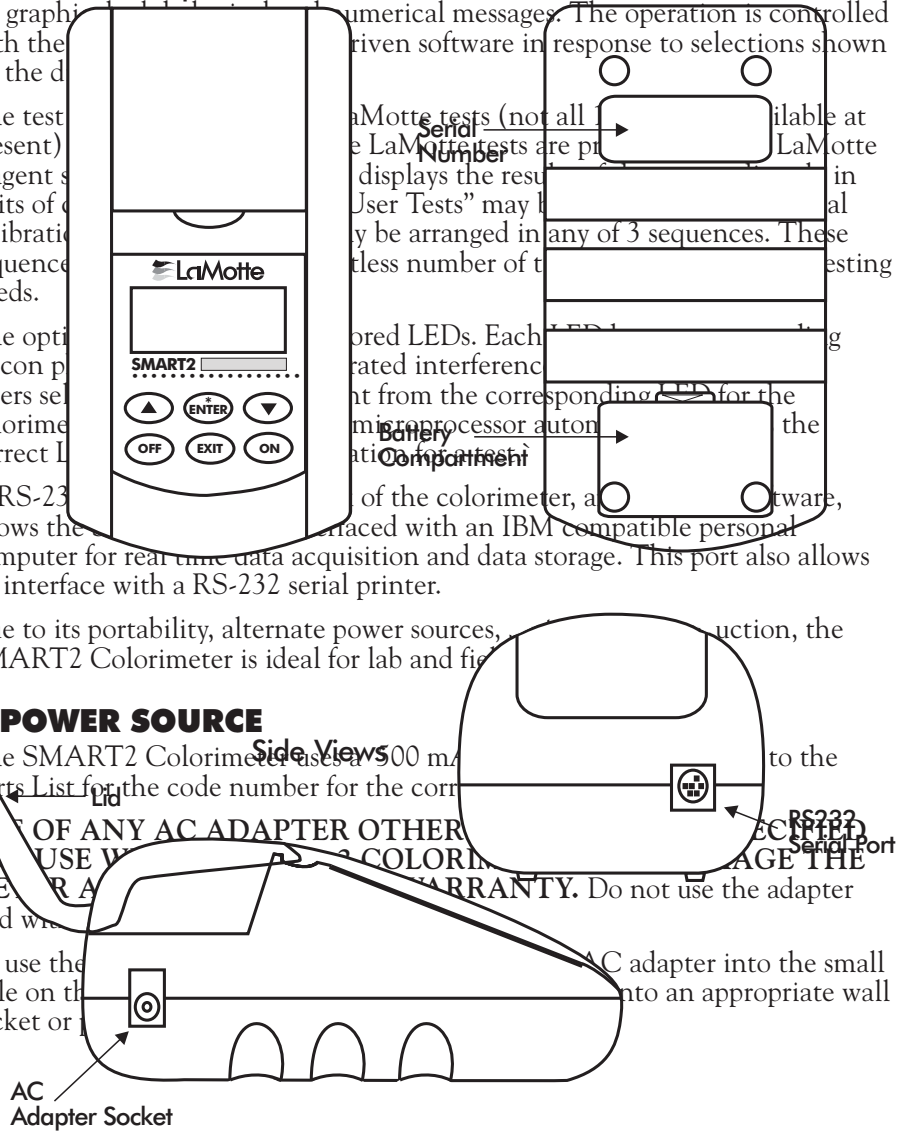


Figure 1

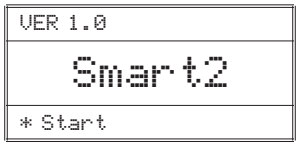
COMPONENTS

Figure 1 shows a diagram of the Smart2 Colorimeter and its components.

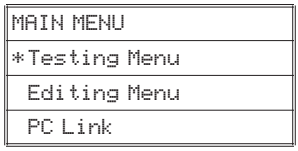
QUICK START

Some quick instructions to get into testing.

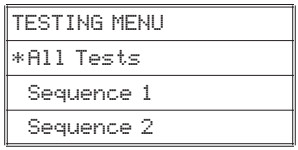
1. Press **ON** to turn on the SMART2. The LaMotte logo screen will appear for about 2 seconds and then the Start screen appears. Press ✱/**ENTER** to start testing.



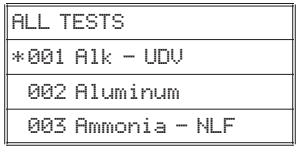
2. The Main Menu will appear. Press ✱/**ENTER** to select TESTING MENU.



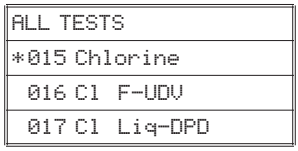
3. Press ✱/**ENTER** to select All Tests.



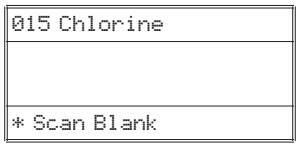
4. Press ▼ or ▲ to move the * to the desired test.



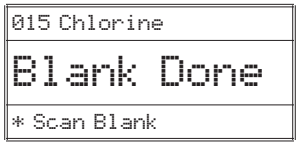
5. Press ✱/**ENTER** to select test.



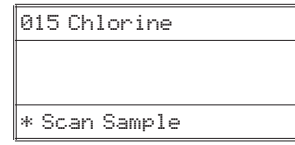
6. Insert blank, press ✱/**ENTER** to scan blank.



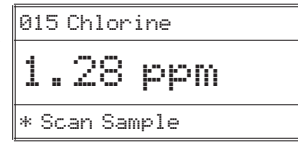
7. The screen will display Blank Done for about 1 second.



8. Insert the reacted sample. Press ✱/**ENTER** to scan sample. The SMART2 will scan the sample and display the concentration.



9. After recording test result, scroll with ▼ or ▲ and make another selection with ✱/**ENTER**. Press **EXIT** to escape to previous menus.



GENERAL OPERATING PROCEDURES

The operation of the SMART2 Colorimeter is controlled by a microprocessor. The microprocessor is programmed with menu driven software. A menu is a list of choices. This allows a selection of various tasks for the colorimeter to perform, such as, scan blank, scan sample, and edit test sequences. The keypad is used to make menu selections which are viewed in the display. There are three selections accessible from the `MAIN MENU`: `Testing Menu`, `Editing Menu` and `PC Link`.

THE KEYPAD

The keypad has 6 buttons which are used to perform specific tasks.

ON	This button is used to turn the colorimeter on.
▼	This button will cause the display to scroll down through a list of menu choices. It will move through a list viewed in the display. It will auto scroll when held down.
▲	This button will cause the display to scroll up in a list of menu choices. It will move through a list viewed in the display. It will auto scroll when held down.
ENTER ✱	This button is used to select the menu choice adjacent to the "*" in a menu viewed in the display.
EXIT	This button is an exit or escape button. When pressed, the display will exit from the current menu and go to the previous menu.
OFF	This button turns the colorimeter off.

SAMPLE HOLDERS

The sample chamber is designed for 25 mm round tubes. Additional sample holders for 16 mm COD tubes and for 1 cm square UDV cuvettes are available for the SMART2 Colorimeter.

Position the COD adapter in the SMART2 chamber so that the grooves in the adapter are aligned with the ridges located at the rear of the chamber. The adapter should be inserted with the small hole, containing the ball plunger, at the top. The ball plunger can be adjusted with a small screwdriver to control the tightness of the fit of the tube in the adapter.

THE DISPLAY & THE MENUS

The display allows menu selections to be viewed and chosen. These choices instruct the colorimeter to perform specific tasks. The menus are viewed in the display using two general formats which are followed from one menu to the next. Each menu is a list of choices or selections.

There are four lines in the display. The top line in each menu is a title or pertinent instruction. The top line does not change unless a new menu is selected. The second and third lines are used in two ways. One way is to display menu choices. The second way takes advantage of the graphical capabilities of the display. Both lines are used to display important messages, such as test results, in a large, easy to read format. The fourth line is used for menu choices.

DISPLAY

TESTING MENU
*FIRST CHOICE
SECOND CHOICE
ANOTHER
AND ANOTHER
AND SO ON

TITLE or INSTRUCTION

MENU CHOICE WINDOW

Think of the menu choices as a vertical list in the display which moves up or down each time an arrow button is pressed. This list or menu is viewed through a window, the menu choice window, in the display. The menu choice window is the lower 2 or 3 lines of the display. Pushing the arrow buttons brings another portion of the menu into menu choice window. This is referred to as scrolling through the menu.

TESTING MENU
*FIRST CHOICE
SECOND CHOICE
ANOTHER
AND ANOTHER
AND SO ON
LAST CHOICE

▼

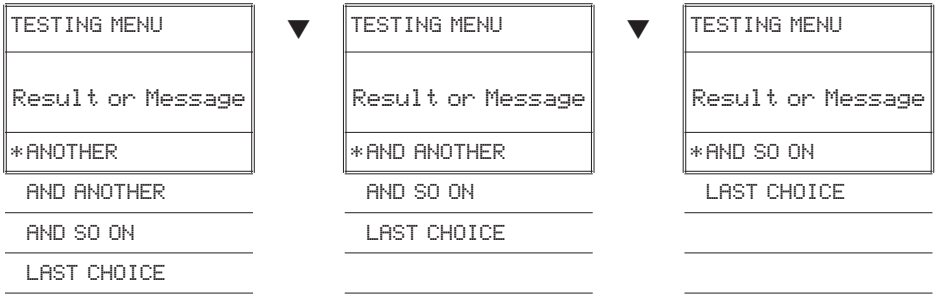
TESTING MENU
SECOND CHOICE
*ANOTHER
AND ANOTHER
AND SO ON
LAST CHOICE

▼

TESTING MENU
ANOTHER
AND ANOTHER
*AND SO ON
LAST CHOICE

An asterisk, "*", will start in the far left position of the top line in the menu choice window. As the menu is scrolled through, different choices appear next to the "*". The "*" in the display corresponds with the **✱/ENTER** button. Pushing the **✱/ENTER** button selects the menu choice which is adjacent to the "*" in the menu choice window.

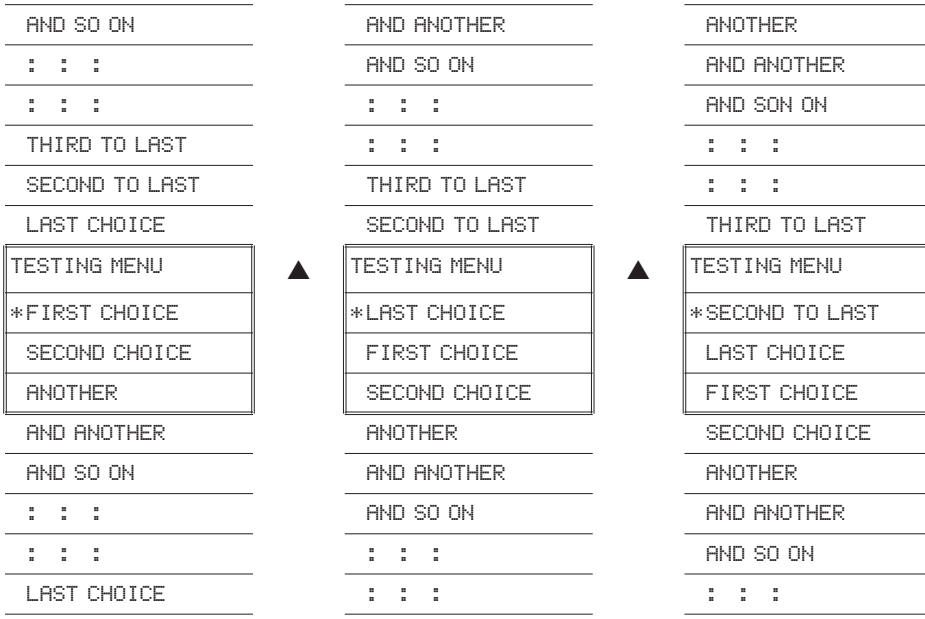
The second general format of the display takes advantage of the graphics capabilities of the display. The top line of the display is still a title line. The middle two lines of the display are used to display important messages, results or graphics in a large, easy to read format. The menus work in the same way as described previously but only one line of the menu is visible at the bottom of the display.



As described previously, the **EXIT** button allows an exit or escape from the current menu and a return to the previous menu. This allows a rapid exit from an inner menu to the main menu by repeatedly pushing the **EXIT** button. Pushing **OFF** at any time will turn the colorimeter off.

LOOPING MENUS

Long menus, such as `All Tests`, incorporate a looping feature which allow the user to quickly reach the last choice in the menu from the first choice. In a looping menu the last choices in the menu are above the first choice and scrolling upward moves through the menu in reverse order. Scrolling downward moves through the menu from first choice to last but the menu starts over following the last choice. So all menu choices can be reached by scrolling in either direction. The diagrams below demonstrate a looping menu.

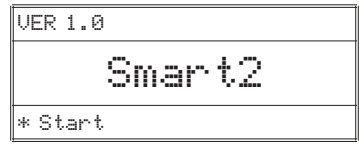


TESTING

TESTING MENU

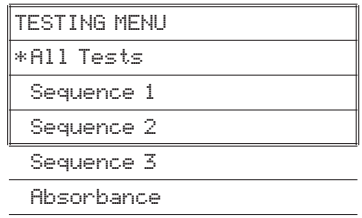
The Testing Menu is used to run all LaMotte pre-programmed tests, USER TESTS and Absorbance test at one of four wavelengths. Testing from any of three sequences can also be done.

1. Press the **ON** button to turn on the SMART2 Colorimeter. The LaMotte logo will appear for about 2 seconds and the the Start screen appears. Press the ✱/**ENTER** button to begin testing.



2. The MAIN MENU will appear. Press the ✱/**ENTER** button to select Testing Menu.

3. Scroll with the ▼ or ▲ buttons and make a selection with the ✱/**ENTER** button. All Tests has all the available tests. The three sequences have selected tests and Absorbance has %T/ABS tests.



SEQUENCES OF TESTS

SEQUENCE 1, SEQUENCE 2, and SEQUENCE 3 are alterable sequences. They may be edited using the Editing Menu. Any of the LaMotte pre-programmed tests or User Tests may be placed in these sequences in whatever testing order that is preferred. Some examples of typical sequences are given below.

SEQUENCE 1
*015 Chlorine
079 Phosphate H
009 Bromine-LR
076 pH TB
061 Moly-HR
086 Silica Hi
045 Hydrazine
032 Cu-DDC
051 Iron Bipyr

SEQUENCE 2
*002 Aluminum
035 Cyanide
041 Fluoride
053 Iron Phen
055 Manganese L
064 Nitrate-N L
067 Nitrite-N L
077 Phenol
078 Phosphate L
090 Sulfide-LR

SEQUENCE 3
*003 Ammonia-N LF
032 Cu-DDC
064 Nitrate-N L
067 Nitrite-N L
074 pH CPR
078 Phosphate L
085 Silica Lo

These alterable sequences allow a series of tests to be setup that are run frequently. The order of the individual tests in the sequence is determined by the user. After running a test, use the * button to select the next test in the sequence. Continue this pattern until the entire sequence has been completed.

All Tests is a fixed sequence containing the LaMotte pre-programmed tests, User Tests, and Absorbance tests.

Modification of the alterable sequences is accomplished through the Editing Menu. This menu is explained in greater detail in EDITING MENU (p. 32).

Pressing the **EXIT** button while in a sequence menu will escape back to the Testing Menu.

Pressing the **OFF** button at any time will turn the colorimeter off.

GENERAL TESTING PROCEDURES

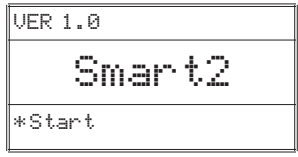
The following are some step by step examples of how to run tests from the Testing Menu. These test procedures are designed to be used with LaMotte SMART Reagent Systems.

LaMotte Company continuously updates the list of pre-programmed tests as the calibrations become available. Pre-programmed calibrations can be added to the SMART2 Colorimeter in the field. A Windows-based computer running a Windows Operating System and an 8 pin mini-DIN/9 pin F D-submin serial cable (order Code 1771) are required.

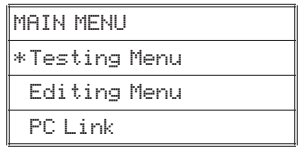
Call LaMotte Technical Services at 1-800-344-3100 (410-778-3100 outside the USA) or email at tech@lamotte.com for a current list of available calibrations and downloading instructions.

TESTING WITH THE LaMOTTE PRE-PROGRAMMED TESTS

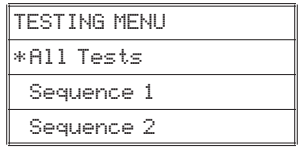
Press ON to turn on the Smart2 Colorimeter. The LaMotte logo will appear for about 2 seconds and then the Start screen appears. Press the **✱/ENTER** button to start testing.



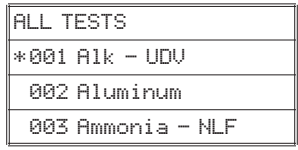
The MAIN MENU will appear. Press the **✱/ENTER** button to select Testing Menu.



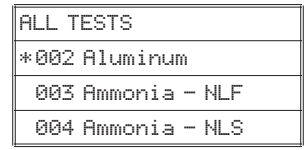
Press the **✱/ENTER** button to select All Tests.



Press the ▼ button to move to the 002 Aluminum to *.

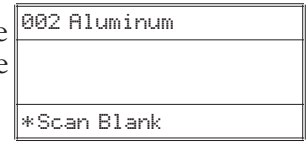


Press the ✱/**ENTER** button to select `002 Aluminum`.

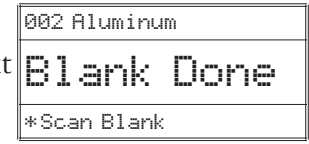


The SMART2 Colorimeter is ready to scan at the correct wavelength. Place the blank in the sample chamber, close the lid and press the ✱/**ENTER** button to scan blank.

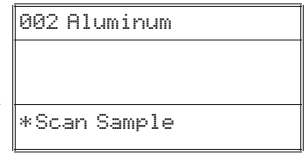
NOTE: Do not keep the button depressed.



The screen will display `Blank Done` for about 1 second. `Scan Sample` will be positioned next to *.

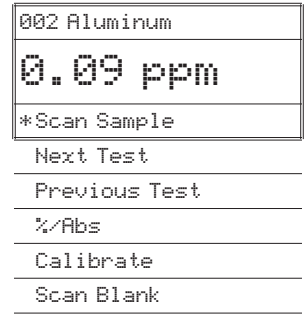


Place the reacted sample in the chamber, close the lid and press the ✱/**ENTER** button to scan sample. The colorimeter will scan the sample and the results screen will appear.



Record test result. To repeat the test, press the ✱/**ENTER** button to scan the sample again. The last blank scanned is used to zero the colorimeter for repeated scans. A different blank can be used by pressing the ▲ button to scroll back to `Scan Blank` and then scanning another blank. Scroll with the ▼ or ▲ buttons and make another selection with the ✱/**ENTER** button. The %T or Absorbance of the last test can be viewed by choosing `%T/Abs`. Press the **EXIT** button to escape to previous menus.

NOTE: The menus loop in this screen so either the ▲ or ▼ buttons will lead to the menu selection needed.



CALIBRATING LaMOTTE PRE-PROGRAMMED TESTS

The LaMotte Pre-Programmed Tests have been pre-calibrated. Recalibration of the pre-programmed tests by the user is not possible. However, a procedure to standardize the calibration can be performed to obtain the most accurate readings or to meet regulatory requirements.

The LaMotte Pre-Programmed tests are standardized with one standard solution. To standardize over the full range of the test, the concentration of the standard should be chosen from the high end of the range. Alternatively, if samples do not cover the full range of the test, a standard should be chosen that is close to the concentration of the samples.

The standardization procedure should be followed as often as required by regulations and laws for compliance monitoring.

In the example below the Aluminum calibration will be standardized.

Prepare a standard solution to be tested. Use 0.10 ppm aluminum.

Use the ▲ or ▼ button to scroll to `002 Aluminum`. Follow instructions in the SMART2 Manual for testing the aluminum standard. Scan the blank.

002 Aluminum
*Scan Blank

The screen will display `Blank Done` for about 1 second. `Scan Sample` will be positioned next to *.

002 Aluminum
Blank Done
*Scan Sample

Place the reacted sample in the chamber, close the lid and press **✱/ENTER** to scan sample. The result will be displayed.

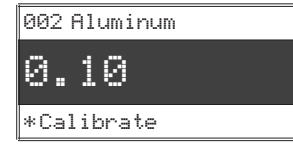
002 Aluminum
*Scan Sample

The displayed result can now be standardized. Use the ▲ or ▼ buttons to scroll to `Calibrate`. Press **✱/ENTER** to select.

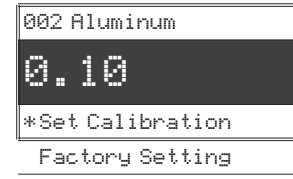
002 Aluminum
0.09 ppm
*Scan Sample
Next Test
Previous Test
%T/Abs
Calibrate
Scan Blank

A reverse font (dark background with light characters) will appear to indicate that the reading can be adjusted. Use ▲ or ▼ to scroll to the concentration of the sample, 0.10 ppm in this example.

Set the calibration by pressing ***/ENTER** to select `Calibrate`.

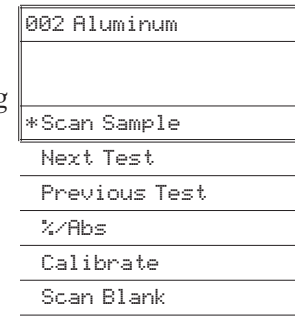


Two menu choices will be offered, `Set Calibration` and `Factory Setting`. Set the calibration by pressing ***/ENTER** to select `Set Calibration`; or use ▲ or ▼ to scroll to and select `Factory Setting` t o revert to the factory calibration.



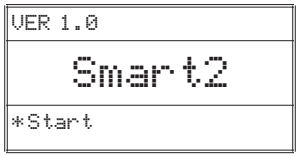
The meter will display the message "`Storing`" and return to `002 Aluminum` test.

The calibration for `002 Aluminum` has now been standardized and can be used for testing. The standardization can be removed by repeating the calibration and selecting `Factory Setting`.

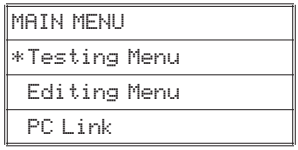


MEASURING IN THE ABSORBANCE MODE

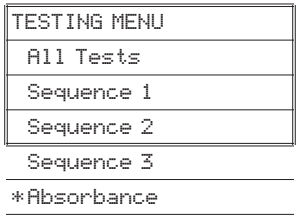
Press **ON** to turn on the SMART2 Colorimeter. The LaMotte logo will appear for about 2 seconds and then the Start screen appears. Press the **✱/ENTER** button to start testing.



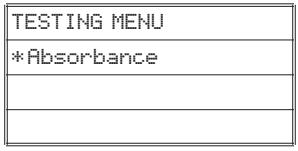
The MAIN MENU will appear. Press the **✱/ENTER** button to select Testing Menu.



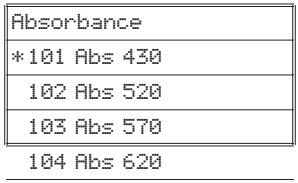
Press the ▼ button to scroll to Absorbance.



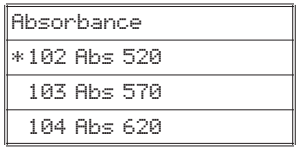
Press the **✱/ENTER** button to select Absorbance.



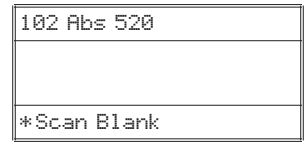
Press the ▼or ▲ buttons to move to the desired test.



Press the **✱/ENTER** button to select test.

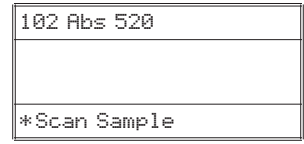


Insert blank, press the ✱/**ENTER** button to scan blank.



The screen will display `Blank Done` for about 1 second.

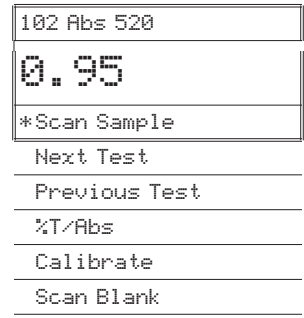
Insert the reacted sample. Press the ✱/**ENTER** button to scan the sample.



Record test result. To repeat the test, press the ✱/**ENTER** button to scan the sample again. The last blank scanned is used to zero the colorimeter for repeated scans. A different blank can be used by pressing the ▲ button to scroll back to `Scan Blank` and then scanning another blank. Scroll with ▼ or ▲ and make another selection with ✱/**ENTER**. The %T or Absorbance of the last test can be viewed by choosing `%T/Abs`. Press **EXIT** to escape to previous menus.

NOTE: The menus loop in this screen so either ▼ or ▲ will lead to the menu selection needed.

NOTE: The Calibrate function does not work in the Absorbance mode.



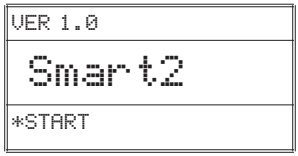
EDITING MENU

The EDITING MENU allows the user to edit sequences, edit user tests, set the clock, edit the logging function, and set the power saving function.

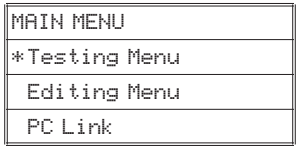
EDIT A SEQUENCE

The EDIT SEQUENCE menu allows three alterable test sequences (SEQUENCE 1, SEQUENCE 2, and SEQUENCE 3) to be edited.

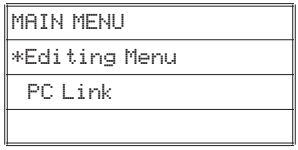
Press **ON** to turn on the SMART2 Colorimeter. The LaMotte logo will appear for about 2 seconds and then the Start screen appears. Press the ***/ENTER** button to start testing.



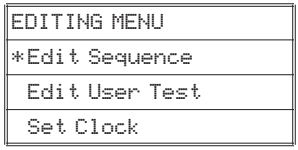
The Main Menu will appear. Press the ▼ button to scroll to Editing Menu.



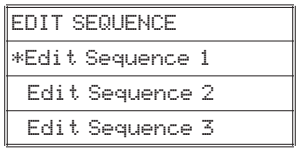
Press the ***/ENTER** button to select Editing Menu.



The Editing Menu appears. Press the ***/ENTER** button to select Editing Sequence.



The Edit Sequence menu appears. Press the ***/ENTER** button to scroll to select Edit Sequence 1.



Sequence 1 appears.

EDIT SEQUENCE 1
*015 Chlorine
079 Phosphate H
009 Bromine-LR

ADDING OR DELETING TESTS

There are three ways to alter a sequence: `Insert Before`, `Insert After`, and `Delete`. `Insert Before` adds a new test to the sequence before the selected test. `Insert After` adds a new test to the sequence after the selected test. `Delete` is used to remove an existing test from a sequence.

Below is a step by step example of how to add a test to `SEQUENCE 1` starting from the `EDIT SEQUENCE 1` menu.

Press the ▼ button to scroll to `009 Bromine-LR`.

EDIT SEQUENCE 1
015 Chlorine
079 Phosphate H
*009 Bromine-LR

Press the **✱/ENTER** button to select `009 Bromine-LR`.

EDIT SEQUENCE 1
*009 Bromine-LR
076 pH TB
060 Moly-LR

Press the **✱/ENTER** button to select `Insert Before`.

EDIT SEQUENCE 1
*Insert Before
Insert After
Delete

The `ALL TESTS` menu appears. Press the ▼ button to move the `002 Aluminum` to `*`.

ALL TESTS
*002 Aluminum
003 Ammonia-N LF
004 Ammonia-N LS

Continued...

Press the ✱/**ENTER** button to select 002 Aluminum.

ALL TESTS
*002 Aluminum
003 Ammonia-N LF
004 Ammonia-N LS

Sequence 1 appears in EDIT SEQUENCE 1 menu and 002 Aluminum is now before Bromine-LR in the sequence. All changes to Sequence 1 are automatically saved. Press the **EXIT** button to exit the EDIT SEQUENCE 1 menu and return to the EDIT SEQUENCE menu or continue editing.

EDIT SEQUENCE 1
*015 Chlorine
079 Phosphate H
002 Aluminum
009 Bromine-LR
076 pH TB
060 Moly-LR

The EDIT SEQUENCE menu appears. Select another sequence to edit or press the **EXIT** button to return to the EDITING MENU. Press the **EXIT** button again to return the the MAIN MENU.

EDIT SEQUENCE 1
*Edit Sequence 1
Edit Sequence 2
Edit Sequence 3

Below is a step by step example of how to delete a test from SEQUENCE 1 starting from the EDIT SEQUENCE 1 menu. The test 002 Aluminum, added in the previous example, will be deleted.

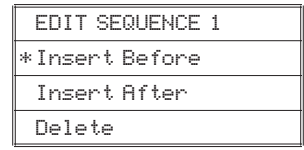
Press the ▼ button to scroll to 002 Aluminum.

EDIT SEQUENCE 1
*015 Chlorine
079 Phosphate H
002 Aluminum
009 Bromine-LR
076 pH TB
060 Moly-LR

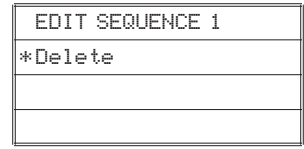
Press the ✱/**ENTER** button to select 002 Aluminum.

EDIT SEQUENCE 1
*002 Aluminum
009 Bromine-LR
076 pH TB

Press the ▼ button to scroll to Delete.

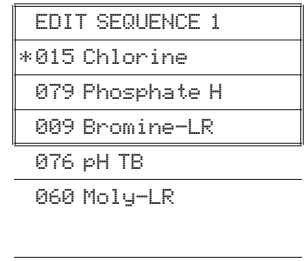


Press the ***/ENTER** button to select Delete.

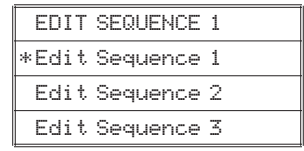


Sequence 1 appears in the EDIT SEQUENCE 1 menu and 002 Aluminum has been deleted. All changes to SEQUENCE 1 are automatically saved.

Press the **EXIT** button to exit the EDIT SEQUENCE 1 menu and return to the EDIT SEQUENCE menu or continue editing.



The EDIT SEQUENCE menu appears. Select another sequence to edit or press the **EXIT** button to return to the EDITING MENU. Press the **EXIT** button again to return the the MAIN MENU.



EDIT USER TESTS

If a test other than the LaMotte programmed tests is performed regularly, a calibration for it may be entered in one of the 10 User Tests. These tests are originally named "User Test 1 – 10". It will be possible to rename the test, select a wavelength, enter a new calibration, select the number of decimal places used to display the results, and select the units. A User Test may be added for a reagent system for which no precalibrated test exists. A calibration of a LaMotte reagent system may also be entered. The calibration of a User Test can be changed at any time.

The User Tests have the ability to handle 2 data points. The colorimeter will determine the absorbance of the standards and calculate a response that will be stored to determine the concentration of future samples of unknown concentration. These standards should cover all the concentrations for the range of the test being performed and be scanned beginning with the low concentration and finishing with the high concentration (for more information about this, see CALIBRATION CURVES, page 13). Prepare these solutions prior to entering a new calibration.

NOTE: A calibration procedure must be performed before using any of the User Tests.

The User Tests can be placed in any of the alterable sequences using EDIT SEQUENCES.

To edit a User Test, start at the EDITING MENU. Scroll down to Edit User Test.

EDITING MENU
*Edit Sequences
Edit User Test
Set Clock

Press the ***/ENTER** button to select the Edit User Test.

EDITING MENU
*Edit User Test
Set Clock
Edit Logging

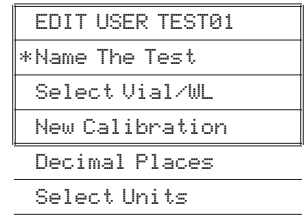
From the EDIT USER TEST menu, select the User Test to be entered or changed. In this example, choose 105 User Test 01. Use the ▼ and ▲ buttons to scroll to other User Tests if desired. Select the User Test by pressing the ***/ENTER** button.

EDIT USER TEST
*105 User Test01
106 User Test02
107 User Test03
108 User Test04
: : :
114 User Test10

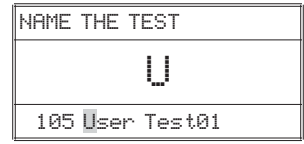
NAMING THE TEST

A User Test can be up to 11 characters long. The menu choices for each character are 26 upper case letters A to Z, 26 lower case letters a to z, ten numerals 0 to 9, a space (SP), a dash (–) and a decimal point (.). The existing name is displayed on the bottom line of the display. A cursor will be over the character which is to be edited and that character is also displayed in the center of the display. The character can be changed by using the ▼ and ▲ buttons to scroll to other characters. Use the ✱/**ENTER** button to select a character. The edited name is saved at any time by pressing **EXIT** or by pressing the ✱/**ENTER** button after selecting the eleventh character.

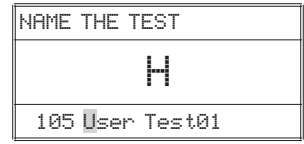
From the Edit User Test01 menu press the ✱/**ENTER** button to select Name The Test and change the name of User Test 01.



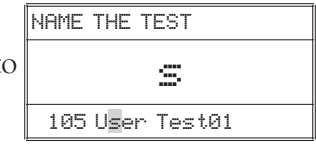
The cursor is over the letter "U" in 105 User Test01 and the letter "U" is displayed in the large font in the center of the display.



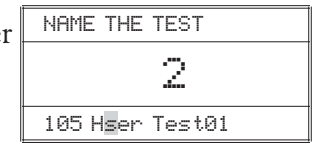
Change the name to H2O. Use the ▼ and ▲ buttons to scroll to the letter "H" into the center of the display. Press the ✱/**ENTER** button to select the letter "H".



The letter "H" has been entered in the first position of the name and the cursor has moved to the second letter "s".

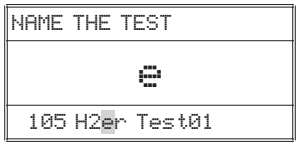


Use the ▼ and ▲ buttons to scroll to the number "2" into the center of the display. Press the ✱/**ENTER** button to select the number "2".

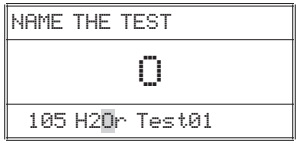


Continued...

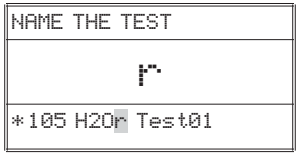
The number "2" has been entered in the second position of the name and the cursor has moved to the third letter "e".



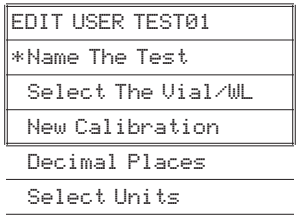
Use the ▼ and ▲ buttons to scroll to the letter "O" into the center of the display. Press the **✱/ENTER** button to select the letter "O".



The letter "O" has been entered in the third position of the name and the cursor has moved to the fourth letter "r". Press the **EXIT** button to save the name entered up to this point.



The meter will display the message "Storing" and return to the EDIT USER TEST01 menu.



SELECTING THE VIAL AND WAVELENGTH

The Smart2 Colorimeter has three different vials (the 25 mm 0290 tube, UDVs and COD tubes) and 4 different wavelengths (430, 520, 570, and 620 nm). The colorimeter uses different settings for each of the twelve combinations of vial and wavelength. These twelve settings are called channels. Choose the channel with the correct wavelength and vial for the test.

Use the ▼ button to scroll to `Select Vial/WL` and press ***/ENTER** button to select.

EDIT USER TEST01
*Name The Test
Select Vial/WL
New Calibration
Decimal Places
Select Units

Use the ▼ and ▲ buttons to scroll to the appropriate channel and press ***/ENTER** button to select.

NOTE: This is a looping menu.

: : :
Ch11 620nm COD
Ch12 570nm COD
SELECT CHANNEL
*Ch1 520nm 25mm
Ch2 430nm 25mm
Ch3 620nm 25mm
Ch4 570nm 25mm
Ch5 520nm UDV
Ch6 430nm UDV
: : :

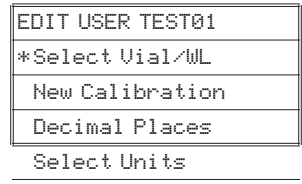
The meter will display the message "`Storing`" and return to the `EDIT USER TEST01` menu.

EDIT USER TEST01
*Select The Vial/WL
New Calibration
Decimal Places
Select Units

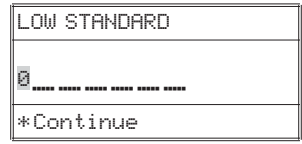
ENTERING A NEW CALIBRATION

To enter a new calibration two reacted standards solutions of known concentration are required: a "low standard" and a "high standard". These should be ready to use.

Use the ▼ button to scroll to `New Calibration` and press **✱/ENTER** button to select.

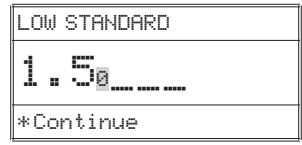


Input the concentration of the `LOW STANDARD` by using the ▼ and ▲ buttons to scroll the first digit of the concentration into the first position on the display. Press **✱/ENTER** button to select that digit (`1` for this example).



The number "`0`" is always the starting point for the next digit. Continue selecting digits or a decimal point to enter the concentration (up to seven characters).

"`1.5`" has been entered in this example. Press **✱/ENTER** button four times to input "`0`" as the last four digits. Pressing **✱/ENTER** after selecting the last digit saves the concentration.

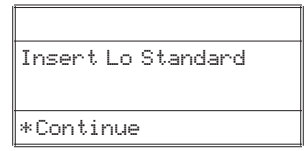


Input the concentration of the `HIGH STANDARD` by using the same method as for the low standard.

Place a clear blank in the sample chamber. Press the ✱/**ENTER** button to scan the blank.

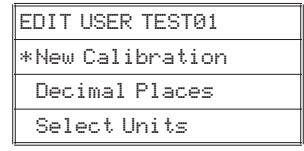
The screen will display `Blank Done` for about 1 second.

Place the reacted low standard in the sample chamber. Press ✱/**ENTER** to scan the low standard.



Place the reacted high standard in the sample chamber. Press ✱/**ENTER** to scan the high standard.

The meter will display the message "`Storing`" and return to the `EDIT USER TEST01` menu.



SELECTING THE NUMERICAL FORMAT OF THE RESULT

To input tests with very different ranges, the number of decimal places displayed for a result can be selected. A test which ranges from 20 to 1000 ppm should not be displayed with three decimal places. A test with a range from 0.010 to 0.500 needs three decimal places (the microprocessor will always calculate the concentration to many more significant figures than will be displayed). Menu choices of 0, 1, 2, or 3 decimal places will be given for the display.

Use the ▼ button to scroll to `Decimal Places` and press ✱/**ENTER** button to select.

EDIT USER TEST01
*New Calibration
Decimal Places
Select Units

Use the ▼ button to scroll to the number of decimal places to be shown and press ✱/**ENTER** to select.

DECIMAL PLACES?	
*None	0
One	0.0
Two	0.00
Three	0.000

The meter will display the message "`Storing`" and return to the `EDIT USER TEST01` menu.

Storing

EDIT USER TEST01
*Decimal Places
Select Units

SELECTING THE UNITS OF CONCENTRATION

The SMART2 Colorimeter has seven options for units of concentration. They are No Units, ppm, pH, FTU, ppb, ppt and mgL.

Use the ▼ button to scroll to `Select Units` and press **✱/ENTER** to select.

EDIT USER TEST01
*Decimal Places
Select Units

Use the ▼ button to scroll to the appropriate unit and press **✱/ENTER** to select.

SELECT UNITS
*No Units
ppm
pH
FTU
ppb
ppt
mgL

The meter will display the message "`Storing`" and return to the `EDIT USER TEST01` menu.

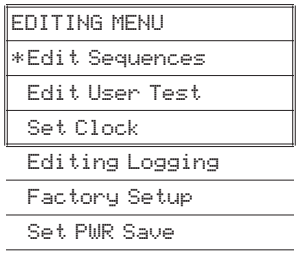
Storing

EDIT USER TEST01
*Select Units

SETTING THE CLOCK

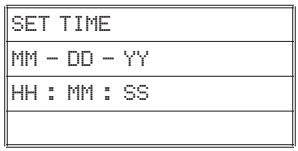
Setting the clock allows the correct time and date stamp to be stored with each reading in the data logger and with each reading sent out the serial port.

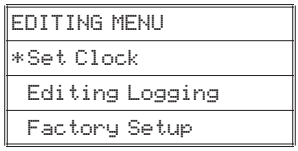
From the EDITING MENU use the ▼ button to scroll to Set Clock. Press ✱/**ENTER** to select.



The current date and time are displayed as month - day - year on the first line and as hours : minutes : seconds on the second line. A two-digit number is displayed for each setting. Use the ▼ and ▲ buttons to scroll to the appropriate number and press ✱/**ENTER** to select. The cursor will move to the next digit. Set all subsequent numbers in the same manner. Selecting the final digit in the seconds field stores the date and time and returns to the EDITING MENU.

NOTE: These are looping menus.

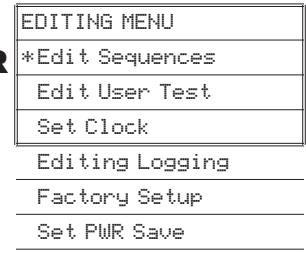




TURNING THE DATA LOGGER ON AND OFF

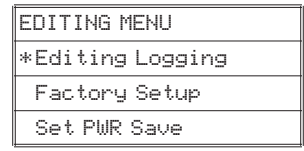
The default setting for the datalogger is "Enabled" or turned off. If there is no need for data logging, this setting is suggested. If data logging is needed, the data logger can be "Enabled" or turned on.

From the `EDITING MENU` use the ▼ button to scroll to `Edit Logging`. Press **∗/ENTER** to select.



The current setting is always displayed next to the ∗. To change the setting, use the ▼ or ▲ buttons to scroll to the other setting. Press **∗/ENTER** to select.

The meter will display the message "`Storing`" and return to the `EDITING MENU`.



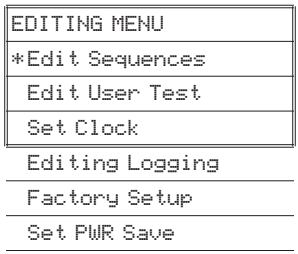
FACTORY SETUP

The Factory Setup menu is used in the manufacturing of the SMART2 Colorimeter. This menu is not for use by the operator in the field.

SETTING THE POWER SAVING FUNCTION

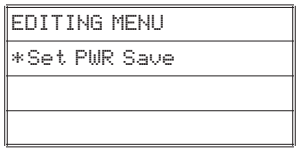
The SMART2 Colorimeter has a power saving function that turns the meter off after an interval of inactivity. If no buttons have been pressed during that interval the meter will turn itself off. This interval can be disabled or set for 5, 15, 30 or 60 minutes. The default setting is 5 minutes.

From the `EDITING MENU` use the ▼ button to scroll to `Set PWR Save`. Press ✱/**ENTER** to select.



The current setting is always displayed next to the *. To change the setting, use the ▼ or ▲ buttons to scroll to the appropriate setting. Press ✱/**ENTER** to select.

The meter will display the message "`Storing`" and return to the `EDITING MENU`.



PC LINK

The SMART2 Colorimeter may be interfaced with any Windows-based computer by using the LaMotte SMARTLink2 Program and Interface Cable (Order Code 1912-3 [3.5 disk] or 1912-CD [compact disk]). The program stores customer information and test data in a database. It can be used to download data stored in the Smart2 datalogger for each test site.

The colorimeter may also be interfaced with an RS-232 serial printer, using an interface cable (Order Code 1772) and setting the printer configuration to the Output as described below.

Choose PC Link from the Main Menu. The user can download the entire datalogging buffer. Downloading does not delete or empty the datalogger.

OUTPUT

RS-232 compatible, asynchronous serial, 9600 baud, no parity, 8 data bits, 1 stop bit.

COMPUTER CONNECTION

RS-232 interface connection, 8 pin mini-DIN/9 pin F D-submin. (Order Code 1772).

BATTERY OPERATION

The colorimeter may be run on battery power or AC using the AC adapter. If using the meter as a benchtop unit, keep it plugged in if possible. If used on only battery power, always have a spare battery on hand.

If the battery power is low, the SMART2 will display "LOW BATT" and turn off.

REPLACING THE BATTERY

The SMART2 Colorimeter uses a standard 9-volt alkaline battery that is available worldwide. The battery compartment is located on the bottom of the the case.

To replace the battery:

1. Open the battery compartment lid.
2. Remove the battery and disconnect the battery from the polarized plug.
3. Carefully connect the new battery to the polarized plug and insert it into the compartment.
4. Close the battery compartment lid.

MAINTENANCE

CLEANING

Clean with a damp, lint-free cloth.

DO NOT ALLOW WATER TO ENTER THE COLORIMETER CHAMBER OR ANY OTHER PARTS OF THE METER.

METER CARE

The optical system of the SMART2 must be kept clean and dry for optimal performance. Dry the colorimeter tubes before placing them in the chamber to avoid introducing moisture. For best results store the instrument in an area that is dry and free from aggressive chemical vapors.

METER DISPOSAL

Waste Electrical and Electronic Equipment (WEEE)

Natural resources were used in the production of this equipment. This equipment may contain materials that are hazardous to health and the environment. To avoid harm to the environment and natural resources, the use of appropriate take-back systems is recommended. The crossed out wheeled bin symbol on the meter encourages you to use these systems when disposing of this equipment.

Take-back systems will allow the materials to be reused or recycled in a way that will not harm the environment. For more information on approved collection, reuse, and recycling systems contact your local or regional waste administration or recycling service.

TROUBLESHOOTING GUIDE

ERROR MESSAGES

▪ OVER RANGE

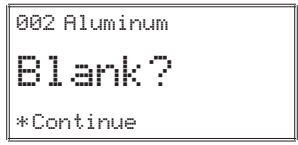
If the message OVERRANGE is displayed when scanning a sample, the sample may be over range or under range. If the sample is over range the sample should be diluted and tested again (see Sample Dilution Techniques and Volumetric Measurements, page 16).

If OVERRANGE is displayed, press the ✱/**ENTER** button to continue testing on diluted samples.

▪ BLANK

If the message Blank? is displayed when scanning a sample, the sample had a lower reading than the blank. Review test procedure to determine whether a reagent blank is required.Visually check for color development in reacted sample. Repeat test if necessary.

If Blank? is displayed, press the ✱/**ENTER** button to continue. Check to see if the meter was blanked properly.



CALIBRATION

As with all pre-calibrated meters, it is highly recommended, even if not required by regulations, that the user periodically verify the performance of the meter by running standards with a predetermined concentration. Results outside of specification are an indication that the meter needs to be adjusted. This can be done following the user calibration described on page 31. If the user calibration fails to properly adjust the meter then the meter should be returned to LaMotte Company for recalibration. (See page 5).

HELPFUL HINTS

STRAY LIGHT

The SMART2 Colorimeter should have no problems with stray light. Make sure that the sample compartment lid is always fully closed, except when testing COD with the adapter.

SMART 2

Colorimeter

REAGENT SYSTEMS

TEST INSTRUCTIONS

LaMotte

v.2.4 • Printed 12.08
1919-test

SMART2 COLORIMETER REAGENT SYSTEMS

SMART2 REAGENT SYSTEMS LIST

LaMotte Company continuously updates the list of pre-programmed tests as the calibrations become available. Pre-programmed calibrations can be added to the Smart2 Colorimeter in the field. A Windows-based computer running a Windows Operating System and an 8 pin mini-DIN/9 pin F D-submin serial cable (order Code 1771) are required.

Call LaMotte Technical Services at 1-800-344-3100 (410-778-3100 outside the USA) or email at tech@lamotte.com for a current list of available calibrations and downloading instructions.

Test Factor (Test #)	*Range (ppm)*	*Test Method (# of Reagents)*	*# of Tests*
Alkalinity-UDV (1)	0–200	Unit Dose Vials (1)	50
Aluminum (2)	0.00–0.30	Eriochrome Cyanine R (4)	50
Ammonia Nitrogen-Low Range, Fresh Water (3)	0.00–1.00	Salicylate (3)	25
Ammonia Nitrogen-Low Range, Salt Water (4)	0.00–1.00	Salicylate (3)	25
Ammonia Nitrogen-High Range (5)	0.00–4.00	Nesslerization (2)	50
Benzotrizole (10)	0.0–30.0	UV Photolysis (3)	50
Biguanide (7)	0–70	Colorimetric	50
Boron (8)	0.00–0.80	Azomethine-H (2)	25
Bromine-Low Range (9) *See Chlorine-Bromine-Iodine*	0.00–9.00	DPD (3)	100
Bromine-UDV (11)	0.0–22.0	DPD (1)	50
Cadmium (12)	0.00–1.00	PAN (4)	50
Ca & Mg Hardness-UDV (13)	0–400	Unit Dose Vials (1)	50
Carbohydrazide (14) *See Oxygen Scavengers*	0.000–0.900	Iron Reduction (3)	100
Chloride-TesTab (21)	0.0–30.0	Argentometric (1)	50
Chlorine (15)	0.00–4.00	DPD (3)	100
Chlorine-Free-UDV (16)	0.00–10.00	DPD (1)	50
Chlorine-Liquid DPD (17)	0.00–4.00	DPD (3)	144
Chlorine-Total-UDV (18)	0.00–10.00	DPD (1)	50
Chlorine Dioxide (20)	0.00–8.00	DPD (2)	100
Chromium (22)	0.00–1.00	Diphenylcarbohydrazide (1) or (5)	100
Chromium-TesTab (23)	0.00–1.00	Diphenylcarbohydrazide (1)	50

Test Factor (Test #)	*Range (ppm)*	*Test Method (# of Reagents)*	*# of Tests*
Cobalt (24)	0.00–2.00	PAN (3)	50
COD-Low Range (25)	5–150	Digestion (1)	25
COD-Standard Range (26)	0-1500	Digestion (1)	25
COD-High Range (27)	0–15000	Digestion (1)	25
Color (28)	0–1000	Platinum Cobalt (0)	∞
Copper-BCA-Low Range (29)	0.00–3.50	Bicinchoninic Acid (1)	50
Copper-Cuprizone (31)	0.00–2.00	Cuprizone (2)	50
Copper-DDC (32)	0.00–6.00	Diethyldithiocarbamate (1)	50
Copper-UDV (33)	0.0–4.0	Bicinchoninic Acid (1)	50
Cyanide (35)	0.00–0.50	Pyridine-Barbituric Acid (5)	50
Cyanuric Acid (36)	5–200	Melamine (1)	50
Cyanuric Acid-UDV (37)	5–150	Melamine (1)	50
DEHA (38) *See Oxygen Scavengers*	0.000–0.700	Iron Reduction (3)	100
Dissolved Oxygen (39)	0.0–11.0	Winkler Colorimetric (3)	100
Erythorbic Acid (40) *See Oxygen Scavengers*	0.00–3.00	Iron Reduction (3)	100
Fluoride (41)	0.00–2.00	SPADNS (2)	50
Hardness (Total) UDV (13)	15-400	U nit dose Vial (1)	50
Hydrazine (45)	0.00–1.00	P-dimethylaminobenzaldehyde (2)	50
Hydrogen Peroxide-Low Range (46)	0.00–1.50	DPD (2)	100
Hydrogen Peroxide-High Range (47)	0–60	DPD (2)	50
Hydrogen Peroxide-Shock (48)	0–225	DPD (2)	100
Hydroquinone (49) *See Oxygen Scavengers*	0.00–2.00	Iron Reduction (3)	100
Iodine (50) *See Chlorine-Bromine-Iodine*	0.00–14.00	DPD (2)	100
Iron-Bipyridyl (51)	0.00–6.00	Bipyridyl (2)	50
Iron-UDV (52)	0.00–10.00	Bipyridyl (1)	50
Iron-Phenanthroline (53)	0.00–5.00	1,10 Phenanthroline (2)	50
Lead (54)	0.00–5.00	PAR (5)	50
Manganese-Low Range (55)	0.00–0.70	PAN (3)	50
Manganese-High Range (56)	0.0–15.0	Periodate (2)	50
Mercury (57)	0.00–1.50	TMK (3)	50

Test Factor (Test #)	*Range (ppm)*	*Test Method (# of Reagents)*	*# of Tests*
Methylethylketoxime (58) *See Oxygen Scavengers*	0.00–3.00	Iron Reduction (3)	100
Molybdenum-High Range (61)	0.0–50.0	Thioglycolate (3)	50
Nickel (63)	0.00–8.00	Dimethylglyoxime (6)	50
Nitrate Nitrogen-Low Range (64)	0.00–3.00	Cadmium Reduction (2)	20
Nitrate-TesTab (66)	0.0–60.0	Zinc Reduction (1)	50
Nitrite Nitrogen-Low Range (67)	0.00–0.80	Diazotization (2)	20
Nitrite-TesTab (69)	0.00–1.25	Diazotization (1)	50
Nitrogen, Total (70)	0–25 mg/L	Chromotropic Acid/Digestion (6)	25
Oxygen Scanvengers	various	DEHA (3)	50
Ozone-DPD (73)	0.00–3.00	DPD (3)	144
Ozone-Low Range (71)	0.00–0.40	Indigo Trisulfonate (3)	100
Ozone-High Range (72)	0.00–2.50	Indigo Trisulfonate (3)	20
pH-Chlorophenol Red (74)	5.0–6.8	Chlorophenol Red (1)	100
pH-Phenol Red (75)	6.6–8.4	Phenol Red (1)	100
pH-Thymol Blue (76)	8.0–9.6	Thymol Blue (1)	100
Phenol (77)	0.00–6.00	Aminoantipyrine (3)	50
Phosphate-Low Range (78)	0.00–3.00	Ascorbic Acid Reduction (2)	50
Phosphate-High Range (79)	0.0–70.0	Vanodomolybdphosphoric Acid (1)	50
Phosphorus, Total Low-Range (82)	0.00–3.50 mg/L	Ascorbic Acid/Digestion (5)	25
Phosphorus, Total High-Range (83)	0.0–100.0 mg/L	Molybdovanadate/Digestion (5)	25
Potassium (81)	0.0–10.0	Tetraphenylboron (2)	100
Silica-Low Range (85)	0.0–4.0	Heteropoly Blue (4)	50
Silica-High Range (86)	0–75	Silicomolybdate (3)	50
Sulfate-High Range (89)	5–100	Barium Chloride (1)	50
Sulfide-Low Range (90)	0.00–1.50	Methylene Blue (3)	50
Surfactants (94)	0.0–8.0	Bromphenol Blue (3)	50
Tannin (96)	0.0–10.0	Tungsto-molybdophosphoric Acid (2)	50
Tolytriazole (97) *See Benzotriazole*	0–30	UV Photolysis (3)	50
Turbidity (98)	0–400 FTU	Absorption (0)	∞
Zinc-Low Range (99)	0.00–3.00	Zincon (6)	50

On the meter display, "NA" following the test number indicates that a calibration for that test number is not available.

ALKALINITY–UDV

UNIT DOSE VIALS • CODE 4318-H

QUANTITY	CONTENTS	CODE
1	Alkalinity Unit Dose Vials, 10 pouches	4318-H

Equipment needed but not supplied:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 6 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tip (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

APPLICATION: Drinking and surface waters; swimming pool water.

RANGE: 0–200 ppm as $CaCO_3$

METHOD: The sample is added to a buffered indicator reagent. The color that develops, ranging from yellow to blue, will indicate the amount of alkalinity in the sample.

SAMPLE HANDLING & PRESERVATION: Samples should be analyzed as soon as possible after collection. Sample may be refrigerated for 24 hours.

INTERFERENCES: Quats and poly quats at high concentrations will interfere.

PROCEDURE

Use 10 mm square cell adapter

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 1 ALKALINTIY-UDV) from TESTING MENU.
5. Scroll to and select 1 ALKALINITY-UDV from menu.
6. Rinse a clean vial (0156) with sample water.
7. Use the syringe (1184) to add 3 mL of sample to the vial.
8. Insert the vial into chamber, close lid and select SCAN BLANK.
9. Remove vial from the colorimeter.
10. Use the syringe (1184) to add 3 mL of sample to an Alkalinity-UDV vial (4318).
11. Wait 2 minutes.
12. Invert vial 3 times to mix.
    - ☑ NOTE: If powder residue remains in the bottom of the vial after inverting, invert once more and tap bottom of vial sharply once or twice to dislodge powder. Mix.
13. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
14. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack with indicator. When the indicator in the window turns from blue to pink, the bag should be replaced.

ALUMINUM

ERIOCHROME CYANINE R METHOD • CODE 364I-SC

QUANTITY	CONTENTS	CODE
5 g	*Aluminum Inhibitor Reagent	*7865-C
2 x 120 mL	*Aluminum Buffer Reagent	*7866-J
120 mL	Aluminum Indicator Reagent	7867-J
15 mL	Aluminum Complexing Reagent	7868-E
1	Spoon, 0.05 g, plastic	0696
2	Pipets, 1.0 mL, plastic	0354
1	Test Tube, glass, 5 mL w/cap	0230

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Aluminum is the third most common element in the earth's crust, which accounts for its wide appearance in many water supplies. Aluminum exists in water as soluble salts, colloidal compounds, and insoluble compounds. In wastewater that has been treated by alum coagulation it will appear in one or more of the above forms. Properly treated drinking water should have an aluminum concentration below 0.05 mg/L.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastewater.

RANGE: 0.00–0.30 ppm Aluminum

METHOD: Aluminum ions buffered to a pH of 6.0 react with Eriochrome Cyanine R dye to produce a pink to red complex in proportion to the concentration.

SAMPLE HANDLING & PRESERVATION: Collect sample in acid washed glass or plastic bottle. Analyze as soon as possible.

INTERFERENCES: Fluoride and polyphosphate will interfere. Interference from iron and manganese is eliminated by the addition of an inhibitor.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 2 Aluminum).
5. Scroll to and select 2 Aluminum from menu.
6. Rinse a clean colorimeter tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into colorimeter chamber and select SCAN BLANK.
8. Rinse a clean test tube (0230) with sample water. Fill to the 5 mL line with sample.
9. Remove tube from colorimeter. Empty sample from tube (0290).
10. Add 5 mL sample from test tube (0230) to empty tube (0290).
11. Use the 0.05 g spoon (0696) to add one measure of *Aluminum Inhibitor Reagent (7865). Cap and mix.
12. Use a 1.0 mL pipet (0354) to add 2 mL of *Aluminum Buffer Reagent (7866). Cap and mix.
13. Use a second 1.0 mL pipet (0354) to add 1 mL of Aluminum Indicator Reagent (7867). Cap and mix contents. Wait 5 minutes for maximum color development.
14. At end of 5 minute waiting period, mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For the best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Add 5 drops of Aluminum Complexing Reagent (7868). Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

AMMONIA-NITROGEN - LOW RANGE

SALICYLATE METHOD • CODE 3659-01-SC

QUANTITY	CONTENTS	CODE
60 mL	*Salicylate Ammonia #1	*3978-H
10 g	*Salicylate #2	*7457-D
2 x 5 g	*Salicylate #3	*7458-C
1	Spoon, 0.1 g, plastic	0699
1	Spoon, 0.15 g, plastic	0727
1	Pipet, 1.0 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Ammonia nitrogen is present in various concentrations in many surface and ground water supplies. Any sudden change in the concentration of ammonia nitrogen in a water supply is cause for suspicion. A product of microbiological activity, ammonia nitrogen is sometimes accepted as chemical evidence of pollution when encountered in natural waters.

Ammonia is rapidly oxidized in natural water systems by special bacterial groups that produce nitrite and nitrate. This oxidation requires that dissolved oxygen be available in the water. Ammonia is an additional source of nitrogen as a nutrient which may contribute to the expanded growth of undesirable algae and other forms of plant growth that overload the natural system and cause pollution.

APPLICATION: Low concentrations of ammonia in fresh, brackish and salt water; fresh and salt water aquariums.

RANGE: 0.00 - 1.00 ppm Ammonia-Nitrogen

METHOD: Salicylate and ammonia react at high pH in the presence of a chlorine donor and an iron catalyst to form a blue indophenol dye, the concentration of which is proportional to the ammonia concentration in the sample.

SAMPLE HANDLE & PRESERVATION: Ammonia solutions tend to be unstable and should be analyzed immediately. Samples may be stored for 24 hours at 4°C or 28 days at –20°C.

INTERFERENCES: There are few interferences in most natural waters. High concentrations of reducing agents, such as hydrazine, react with the chlorine donor and can result in negative interferences. Color and turbidity can also interfere.

PROCEDURE - FRESH WATER

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 3 Ammonia-NLF) from TESTING MENU.
5. Scroll to and select 3 Ammonia-NLF from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK. (See Note.)
8. Remove tube from colorimeter. Use the 1.0 mL plastic pipet (0354) to add 2.0 mL of *Salicylate Ammonia #1 (3978). Cap and mix.
9. Use the 0.15 g spoon (0727) to add two measures of *Salicylate #2 Reagent (7457). Cap and mix until dissolved. Wait 1 minute.
10. At end of 1 minute waiting period use 0.1 g spoon (0699) to add two measures of *Salicylate #3 Reagent Powder (7458). Cap and shake vigorously for at least 30 seconds and all solid has dissolved. Wait 12 minutes for maximum color development.
11. At the end of the 12 minute waiting period, immediately mix and insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

CALCULATIONS:

To express results as Unionized Ammonia (NH_3):

ppm Unionized Ammonia (NH_3) =
ppm Ammonia-Nitrogen (NH_3–N) x 1.2

To express results as Ionized Ammonia (NH_4):

ppm Ionized Ammonia (NH_4^+) =
ppm Ammonia-Nitrogen (NH_3–N) x 1.3

To determine the percentages of Unionized and Ionized Ammonia-Nitrogen, consult the Appendix.

NOTE:

For the best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

PROCEDURE - SALT WATER

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 4 Ammonia-NLS) from TESTING MENU.
5. Scroll to and select 4 Ammonia-NLS from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK. (See Note.)
8. Remove tube from colorimeter. Use the 1.0 mL plastic pipet (0354) to add 2.0 mL of *Salicylate Ammonia #1 (3978). Cap and mix.
9. Use the 0.15 g spoon (0727) to add two measures of *Salicylate #2 Reagent (7457). Cap and mix until dissolved. Wait 1 minute.
10. At end of 1 minute waiting period use 0.1 g spoon (0699) to add two measures of *Salicylate #3 Reagent Powder (7458). Cap and shake vigorously for at least 30 seconds and all solid has dissolved. Wait 20 minutes for maximum color development.
11. At the end of the 20 minute waiting period, immediately mix and insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

CALCULATIONS:

To express results as Unionized Ammonia (NH_3):

ppm Unionized Ammonia (NH_3) =
ppm Ammonia-Nitrogen (NH_3–N) x 1.2

To express results as Ionized Ammonia (NH_4):

ppm Ionized Ammonia (NH_4^+) =
ppm Ammonia-Nitrogen (NH_3–N) x 1.3

To determine the percentages of Unionized and Ionized Ammonia-Nitrogen, consult the Appendix.

NOTE:

For the best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

AMMONIA-NITROGEN - HIGH RANGE

NESSLERIZATION METHOD • CODE 3642-SC

QUANTITY	CONTENTS	CODE
30 mL	Ammonia Nitrogen Reagent #1	V-4797-G
2 x 30 mL	*Ammonia Nitrogen Reagent #2	*V-4798-G
1	Pipet, 1 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Ammonia nitrogen is present in various concentrations in many surface and ground water supplies. Any sudden change in the concentration of ammonia nitrogen in a water supply is cause for suspicion. A product of microbiological activity, ammonia nitrogen is sometimes accepted as chemical evidence of pollution when encountered in natural waters.

Ammonia is rapidly oxidized in natural water systems by special bacterial groups that produce nitrite and nitrate. This oxidation requires that dissolved oxygen be available in the water. Ammonia is an additional source of nitrogen as a nutrient which may contribute to the expanded growth of undesirable algae and other forms of plant growth that overload the natural system and cause pollution.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastes.

RANGE: 0.00–4.00 Ammonia Nitrogen

METHOD: Ammonia forms a colored complex with Nessler's Reagent in proportion to the amount of ammonia present in the sample. Rochelle salt is added to prevent precipitation of calcium or magnesium in undistilled samples.

SAMPLE HANDLING & PRESERVATION: Ammonia solutions tend to be unstable and should be analyzed immediately. Sample may be stored for 24 hours at 4°C or 28 days at –20°C.

INTERFERENCES: Sample turbidity and color may interfere. Turbidity may be removed by a filtration procedure. Color interference may be eliminated by blanking the instrument with a sample blank.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Scroll to and select ALL TESTS (or another sequence containing 5 Ammonia-N H) from TESTING MENU.
5. Scroll to and select 5 Ammonia-N H from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK. (See Note)
8. Remove tube from colorimeter. Add 8 drops of Ammonia Nitrogen Reagent #1 (V-4797). Cap and mix. Wait 1 minute.
9. Use the 1.0 mL pipet (0354) to add 1.0 mL of *Ammonia Nitrogen Reagent #2 (V-4798). Cap and mix. Allow 5 minutes for maximum color development.
10. At end of the 5 minute waiting period, immediately mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
11. Press **OFF** button to turn the colorimeter off or press the **EXIT** button exit to a previous menu or make another menu selection.

CALCULATIONS:

To express results as Unionized Ammonia (NH_3):

ppm Unionized Ammonia (NH_3) =
ppm Ammonia-Nitrogen (NH_3–N) x 1.2

To express results as Ionized Ammonia (NH_4):

ppm Ionized Ammonia (NH_4^+) =
ppm Ammonia-Nitrogen (NH_3–N) x 1.3

To determine the percentages of Unionized and Ionized Ammonia-Nitrogen, consult the Appendix.

☑ NOTE: For the best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

BENZOTRIAZOLE/TOLYLTRIAZOLE

UV Photolysis Method • CODE 4047

QUANTITY	CONTENTS	CODE
15 g	*Benzotriazole Reagent	*3818-E
25 mL	NaK Tartrate Solution	7841-G
25 mL	*Sulfuric Acid	*6139WT-G
1	pH Test Papers, 1–11	9259
1	Spoon, 0.25 g, plastic	0695
1	Erlenmeyer Flask, 25 mL, glass	2-2109
1	Graduated Cylinder, 25 mL, glass	0417

***WARNING:** Reagents marked with a * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact LaMotte by email, phone or fax.

Equipment needed but not supplied:

QUANTITY	CONTENTS	CODE
1	UV Safety Goggles	31041
1	Penray UV Lamp	31041-1
1	Penray Lamp Power Source	31041-2

Proper safety precautions must be followed when using the Penray UV lamp and power source (31041-1 and 31041-2) to prevent eye and skin damage. Always wear the UV Safety Goggles (31041) while the lamp is turned on. Never handle the lamp itself; always hold it by the socket. Wipe the lamp dry with a clean, soft tissue after each test. Do not operate the lamp outside the Erlenmeyer Flask filled with water.

Benzotriazole and tolyltriazole form strong complexes with metals. They are used in antifreeze for cars, lubricating oil, and photographic anti-fogging agents. In cooling water systems benzotriazole and tolyltriazole are used as corrosion and rust inhibitors together with many kinds of scale inhibitors, bactericides and algaecides.

APPLICATION: Corrosion and rust inhibitors in cooling water systems

RANGE: 0.0 – 30.0 ppm Benzotriazole
0.0 – 30.0 ppm Tolyltriazole

METHOD: Benzotriazole and tolyltriazole are UV-photolyzed in a buffered solution with a pH between 4 and 6. A yellow color develops in proportion to the concentration of triazole present.

SAMPLE HANDLING & PRESERVATION: Samples should be analyzed as soon as possible after collection.

INTERFERENCES: Tolyltriazole with interfere in the benzotriazole test. Benzotriazole will interfere in the tolyltriazole test. Strong reducing or oxidizing agents will interfere.

BENZOTRIAZOLE PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `Testing Menu`.
4. Select `ALL TESTS` (or another sequence containing `10 B triazole` from `TESTING MENU`.
5. Scroll to and select `10 B triazole` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter. Discard the sample.
9. Adjust the sample water temperature to between 20 and 25°C if necessary.
10. Fill the graduated cylinder (0417) to the 25 mL line with sample water. Transfer to the Erlenmeyer Flask (2-2109).
11. Use the pH Test Paper (9259) to check the pH of the sample. If the pH is not between 4 and 6, add one drop of *Sulfuric Acid, 1.0N (6139). Swirl to mix. Continue adding *Sulfuric Acid, 1.0N (6139) one drop at a time, swirling to mix and checking the pH after each drop, until the pH is between 4 and 6.
12. Add 10 drops of NaK Tartrate (7841).
13. Use the 0.25 g spoon (0695) to add one measure of *Benzotriazole Reagent (3818). Swirl to mix until the powder has dissolved.
14. Replace the flask in the slot in the case. Insert the Penray Lamp (31041-1) into the flask. Plug in the Penray Power Source (31041-2) and turn the lamp on for exactly 5 minutes. Remove the lamp from the flask. Rinse and wipe the lamp dry.
15. Fill a test tube (0290) to the 10 mL line with the digested sample. Cap tube.
16. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm Benzotriazole.
17. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

TOLYLTRIAZOLE PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `Testing Menu`.
4. Select `ALL TESTS` (or another sequence containing `97 T triazole` from `TESTING MENU`.
5. Scroll to and select `97 T triazole` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter. Discard the sample.
9. Adjust the sample water temperature to between 20 and 25°C if necessary.
10. Fill the graduated cylinder (0417) to the 25 mL line with sample water. Transfer to the Erlenmeyer Flask (2-2109).
11. Use the pH Test Paper (9259) to check the pH of the sample. If the pH is not between 4 and 6, add one drop of *Sulfuric Acid, 1.0N (6139). Swirl to mix. Continue adding *Sulfuric Acid, 1.0N (6139) one drop at a time, swirling to mix and checking the pH after each drop, until the pH is between 4 and 6.
12. Add 10 drops of NaK Tartrate (7841).
13. Use the 0.25 g spoon (0695) to add one measure of *Benzotriazole Reagent (3818). Swirl to mix until the powder has dissolved.
14. Replace the flask in the slot in the case. Insert the Penray Lamp (31041-1) into the flask. Plug in the Penray Power Source (31041-2) and turn the lamp on for exactly 5 minutes. Remove the lamp from the flask. Rinse and wipe the lamp dry.
15. Fill a test tube (0290) to the 10 mL line with the digested sample. Cap tube.
16. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm Tolyltriazole.
17. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

BIGUANIDE

COLORIMETRIC METHOD • CODE 4044

QUANTITY	CONTENTS	CODE
2 X 60 mL	Biguanide Indicator	3994-H
1	Pipet, plastic, 1.0 mL	0354

Biguanide is a non-chlorine, non-bromine chemical sanitizer. It is more stable than chlorine or bromine and has little chemical odor. Biquanide is an effective bacteriacide but, unlike chlorine and bromine, it does not destroy organic contaminants. Therefore, hydrogen peroxide is added to biguanide pools on a regular basis to eliminate organic contaminants. The optimum recommended level of biguanide is 30 to 50 ppm.

APPLICATION: Swimming pools

RANGE: 0–70 ppm

METHOD: Biguanide complexes with the proprietary indicator to produce a colored solution. The color ranges from yellow through green to blue depending on the biguanide concentration.

SAMPLE HANDLING & PRESERVATION: Samples should be analyzed as soon as possible.

INTERFERENCES: The only interfering substances that are likely to be encountered in pool water are oxidized manganese and oxidizing agents, such as chlorine, bromine and ozone.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select Testing Menu.
4. Select `ALL TESTS` (or another sequence containing `7 Biguanide` from `TESTING MENU`.
5. Scroll to and select `7 Biguanide` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Use the 1.0 mL pipet (0354) to add 2.0 mL of Biguanide Indicator (3994). Cap and invert three times to mix.
10. Wait 1 minute.
11. Insert the tube into chamber. Close lid.
12. Select `SCAN SAMPLE`. Record result in ppm Biguanide
13. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

BOREN

AZOMETHINE-H METHOD • CODE 4868

QUANTITY	CONTENTS	CODE
120 mL	*Boron Buffer	*4869-J
10 g	*Boron Indicator Powder	*4870-D
1	Pipet, plastic, 1.0 mL	0354
1	Spoon, 0.15 g	0727
1	Dark storage chamber, brown	0108

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Small amounts of boron are necessary for plant growth but large amounts can be toxic. In humans, boron aids in the uptake of calcium and the production of strong bones. An excess of boron can affect the central nervous system resulting in a syndrome known as borism. Some natural waters may contain small amounts of boron. Large concentrations may be due to industrial effluent entering waterways. Boron compounds are used in cleaning compounds, paper and paints, fertilizers, glass and ceramics, fire retardants and the production of alloys. In the atomic energy field, boron is a component of neutron shields and nuclear reactors. Some swimming pools use boron buffering systems.

APPLICATION: Surface and saline waters, hydroponic solutions, industrial waste, swimming pools.

RANGE: 0.00–0.80 ppm Boron

METHOD: Azomethine-H and borate form a yellow complex at pH 6 in proportion to the concentration of boron present.

SAMPLE HANDLING & PRESERVATION: Store samples in polyethylene bottles. Do not use borate detergents or glassware.

INTERFERENCES: Interferences in drinking water are unlikely. Manganese, zirconium, chromium, titanium, copper, vanadium, aluminum, beryllium and iron may cause high results.

PROCEDURE

1. This test requires a Reagent Blank. Rinse a tube (0290) with clear, colorless, boron free water. Fill to 10 mL line with clear, colorless, boron free water.
2. Use the 1.0 mL pipet (0354) to add 2 mL of *Boron Buffer (4869). Cap and mix.
3. Use the 0.15 g spoon (0727) to add one level measure of *Boron Indicator Powder (4870). Press full spoon against side of jar to compress powder. Scrape off excess powder on inside neck of bottle. Tap excess off spoon handle.
4. Cap and shake vigorously for 30 seconds.
5. Insert the tube into meter chamber. Close lid.
6. Start a timer set for 30 minutes. Do not open the lid during the waiting time. The reaction is photosensitive.
7. Rinse a clean tube (0290) with Sample Water. Fill to the 10 mL line with sample water. Repeat steps 2–4.
8. Insert the tube into the Dark Storage Chamber (0108). Close top.
9. Start a second timer set for 30 minutes. Do not open the chamber during the waiting time. The reaction is photosensitive.
10. When 2 minutes remain on the first timer (Reagent Blank), press and hold **ON** button until colorimeter turns on.
11. Press **ENTER** to start.
12. Press **ENTER** to select `Testing Menu`.
13. Select `ALL TESTS` (or another sequence containing `8 Boron`) from `TESTING MENU`.
14. Scroll to and select `8 Boron` from menu.
15. At the end of the Reagent Blank 30 minute waiting period, remove Reagent Blank tube from meter chamber. Invert several times to mix.
16. Insert the tube into meter chamber, close lid and select `SCAN BLANK`.
17. Remove the tube from colorimeter.
18. At the end of the Sample Water 30 minute waiting period, remove Sample Water tube from Dark Storage Chamber. Invert several times to mix.
19. Insert tube into meter chamber, close lid and select `SCAN SAMPLE`. Record result in ppm boron.
20. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

BROMINE - UDV

DPD METHOD–UNIT DOSE VIALS • CODE 4311-H

QUANTITY	CONTENTS	CODE
1	*Free Chlorine Unit Dose Vials, 10 pouches	*4311-H

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Equipment needed but not supplied:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 6 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tip (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

Like chlorine, bromine is an effective germicidal agent employed in drinking water treatment, pool and spa water sanitization, food service sanitation, and other public health applications.

APPLICATION: Drinking, surface, and saline waters; swimming pool water; domestic and industrial waters and wastes.

RANGE: 0.0–22.0 ppm Bromine

METHOD: In buffered sample bromine reacts with diethyl-p-phenylene diamine (DPD) to produce a pink-red color in proportion to the concentration of bromine present.

SAMPLE HANDLING & PRESERVATION: Bromine in aqueous solutions is not stable, and the bromine content of samples or solutions, particularly weak solutions, will rapidly decrease. Exposure to sunlight or agitation will accelerate the reduction of bromine present in such solutions. For best results start analysis immediately after sampling. Samples to be analyzed for bromine cannot be preserved or stored.

INTERFERENCE: The only interfering substance likely to be encountered in water is oxidized manganese. The extent of this interference can be determined by treating a sample with sodium arsenite to destroy the bromine present so that the degree of interference can be estimated.

Iodine and chlorine can also interfere, but these are not normally present unless they have been added as sanitizers.

PROCEDURE

Use 10 mm square cell adapter.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select Testing Menu.
4. Select `ALL TESTS` (or another sequence containing `11 Bromine-UDV`) from `TESTING MENU`.
5. Scroll to and select `11 Bromine-UDV` from menu.
6. Rinse a clean vial (0156) with sample water.
7. Use the syringe (1184) to add 3mL of sample to the vial.
8. Insert the vial into chamber, close the lid and select `SCAN BLANK`.
9. Remove the vial from the colorimeter.
10. Use the syringe (1184) to add 3mL of sample to a *Free Chlorine UDV vial (4311).
11. Shake vigorously until powder dissolves completely.
    - ☑ NOTE: If powder residue remains in the bottom of the vial after inverting or air bubbles form, invert once more and tap bottom of vial sharply once or twice to dislodge powder and bubbles. Mix.
12. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm bromine.
13. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack with indicator. When the indicator in the window turns from blue to pink, the bag should be replaced.

CADMIUM

PAN METHOD • CODE 4017

QUANTITY	CONTENTS	CODE
60 mL	*Buffered Ammonia Reagent	*4020-H
15 mL	Sodium Citrate, 10%	6253-E
30 mL	PAN Indicator	4021-G
30 mL	Stabilizing Reagent	4022-G
1	Pipet, 1.0 mL, plastic	0354
2	Pipet, 0.5 mL, plastic	0353

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Cadmium is used in batteries, paint pigments, electroplating processes, and with other metals in the preparation of alloys. The solubility of cadmium in natural water is proportional to the hardness or alkalinity of the water. Cadmium is not an essential nutrient for plants and animals. It is extremely toxic and can accumulate in the kidneys and liver.

APPLICATION: Drinking and surface waters; domestic and industrial wastewater.

RANGE: 0.00–1.00 Cadmium

METHOD: PAN (1-[2-Pyridylazo]-2-Naphthol) forms a red complex with Cadmium (Cd^{+2}) at a pH of 10.

SAMPLE HANDLING & PRESERVATION: Analyze sample as soon as possible. If sample must be stored, acidify with nitric acid to a pH below 2.

INTERFERENCES: Ag^{+2}, Co^{+2}, Cu^{+2}, Mn^{+2}, Ni^{+2}, Zn^{+2}, Y^{+3}, In^{+3}

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `12 Cadmium`) from `TESTING MENU`.
5. Scroll to and select `12 Cadmium` from menu.
6. Rinse a tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Use the 1.0 mL pipet (0354) to add 1.0 mL of *Buffered Ammonia Reagent (4020). Swirl to mix.
9. Add two drops of Sodium Citrate, 10% (6253). Swirl to mix.
10. Use a 0.5 mL pipet (0353) to add 0.5 mL of PAN Indicator (4021). Swirl to mix.
11. Use a 0.5 mL pipet (0353) to add 0.5 mL Stabilizing Reagent (4022). Cap and mix.
12. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
13. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

CALCIUM & MAGNESIUM (TOTAL) HARDNESS–UDV

UNIT DOSE VIALS • CODE 4309-H

QUANTITY	CONTENTS	CODE
1	Calcium Hardness Unit Dose Vials, 10 pouches	4309-H

Equipment needed but not supplied:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 3 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tips (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

APPLICATION: Drinking and surface waters; swimming pool water.

RANGE: 0–400 ppm as $CaCO_3$ Total Hardness

METHOD: Calcium and magnesium react in a strongly buffered medium with an indicator to develop a pale purple color in proportion to the concentration.

SAMPLE HANDLING & PRESERVATION: Samples should be analyzed as soon as possible after collection. If storage is necessary, add 0.5 mL of 20 % hydrochloric acid per 100 mL of sample. However, the added acid will have to be neutralized with NaOH before testing.

INTERFERENCES: Heavy metals will interfere.

PROCEDURE

Use 10 mm square cell adapter.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 13 Ca&Mg H–UDV) from TESTING MENU.
5. Scroll to and selec t 13 Ca&Mg Hard–UDV from menu.
6. Rinse a clean vial (0156) with sample water.
7. Use the syringe (1184) to add 3 mL of sample to the vial.
8. Insert the vial into chamber, close lid and select SCAN BLANK.
9. Remove vial from the colorimeter.
10. Use the syringe (1184) to add 3 mL of sample to a Calcium Hardness UDV vial (4309).
11. Shake vigorously for 10 seconds.
12. Wait one minute.
13. Invert vial 3 times to mix.

    ☑ NOTE: Firmly tap side of vial 5-10 times to remove all air bubbles.
14. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack with indicator. When the indicator in the window turns from blue to pink, the bag should be replaced.

CHLORIDE

ARGENTOMETRIC METHOD • CODE 3693-SC

QUANTITY	CONTENTS	CODE
50	*Chloride Spectrophotometric Grade Tablets	*3885A-H
1	Tablet Crusher	0175

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Chloride is one of the major anions found in water and sewage. The presence of chlorides in large amounts may be due to the natural process of water passing through salt formations in the earth, or it may be evidence of the intrusion of seawater or pollution from industrial processes or domestic wastes. The salt content of water affects the distribution of plant and animal life in an aquatic system, based on the amount of salt they can tolerate.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastewaters.

RANGE: 0.0–30.0 ppm Chloride

METHOD: Silver nitrate reacts with chloride to form turbid silver chloride in proportion to the amount of chloride in the sample.

SAMPLE HANDLING & PRESERVATION: Collect samples in clean, chemically resistant glass or plastic containers. No preservative is needed if sample is to be stored.

INTERFERENCES: Substances in amounts normally found in drinking water will not interfere. Bromide, iodide, cyanide, sulfide, thiosulfate, sulfide and orthophosphate will interfere.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `21 Chloride-TT`) from `TESTING MENU`.
5. Scroll to and select `21 Chloride-TT` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL line with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add one *Chloride Spectrophotometric Grade Tablet (3885A).
10. Use Tablet Crusher (0175) to crush tablet.
11. Cap tube.
12. Invert 2 times.
13. Wait 3 minutes. Do NOT mix.
14. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm chloride.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples.

The reagent system is temperature sensitive. The calibration is for 25°C If sample is at 30°C, multiply resulting ppm by 1.1. If the sample is at 20°, multiply ppm by 0.9.

CHLORINE

LIQUID DPD METHOD • CODE 4859

QUANTITY	CONTENTS	CODE
30 mL	DPD 1A Free Chlorine Reagent	P-6740-G
30 mL	*DPD 1B Free Chlorine Reagent	*P-6741-G
30 mL	*DPD 3 Total Chlorine Reagent	*P-6743-G

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

All water for cities and communities must be sanitized; even waters that come from clean sources, protected watersheds, reservoirs, and deep wells, are commonly sanitized to assure safety. Chlorine is the most commonly used sanitizer for several reasons: it is effective against a wide range of microorganisms, the cost is low, and the methods of applying it have been well developed. If an adequate concentration of chlorine is present in the water for a few minutes, disease producing bacteria will be destroyed. A number of conditions affect the sanitizing action of chlorine. In municipal systems these can be controlled so that if chlorine is detectable, it can be assumed that bacteria have been killed. The factors that influence the rate of sanitization are temperature, pH, presence of other materials that react with chlorine, time, and the concentrations of the various chlorine combinations that are formed in the water with ammonia and other substances that react with chlorine.

The fact that chlorine can be easily detected and measured makes chlorine a favorite water sanitizer of those concerned with the public safety of water supplies. Chlorine concentrations in the range of 0.1 to 0.4 parts per million are usually maintained in municipal supplies.

Chlorine can be added in the form of chlorine gas, liquid sodium hypochlorite (bleach), granular calcium hypochlorite or as organic chlorine compounds. Chlorine is not present in natural water supplies; if it is present it is the result of chlorination of a water supply or of chlorinated compounds being discharged as waste from industrial operations. The presence of chlorine in concentrations above 0.5 parts per million should be considered evidence of pollution from chlorine treated effluents or from a process in which high concentrations of chlorine are used.

APPLICATION: Drinking, surface, and saline waters; swimming pool water; domestic and industrial wastes.

RANGE: 0.00–4.00 ppm Chlorine

METHOD: In the absence of iodide, free available chlorine reacts instantly with DPD to produce a red color. Subsequent addition of potassium iodide evokes a rapid color response from the combined forms of chlorine (chloramines).

SAMPLE HANDLING & PRESERVATION: Chlorine in aqueous solutions is not stable, and the chlorine content of samples or solutions, particularly weak solutions, will rapidly decrease. Exposure to sunlight or agitation will accelerate the reduction of chlorine present in such solutions. For best results, start analysis immediately after sampling. Samples to be analyzed for chlorine cannot be preserved or stored.

INTERFERENCE: The only interfering substance likely to be encountered in water is oxidized manganese. The extent of this interference can be determined by treating a sample with sodium arsenite to destroy the chlorine present so that the degree of interference can be measured.

Iodine and bromine can give a positive interference, but these are not normally present unless they have been added as sanitizers.

PROCEDURE–FREE CHLORINE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `Testing Menu`.
4. Select `ALL TESTS` (or another sequence containing `17 Cl DPD-Liq`) from `TESTING MENU`.
5. Scroll to and select `17 Cl DPD-Liq` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add 5 drops of DPD 1A Free Chlorine Reagent (P-6740).
10. Add 5 drops of *DPD 1B Free Chlorine Reagent (P-6741). Cap and mix.
11. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result as ppm free chlorine.

PROCEDURE–TOTAL CHLORINE

12. Add 5 drops of *DPD 3 Total Chlorine Reagent (P-6743). Cap and mix.
    ☑ NOTE: For wastewater samples, *Standard Methods for the Examination of Water and Wastewater* recommends waiting 2 minutes for full color development.
13. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result as ppm total chlorine.
14. Subtract the Free Chlorine reading from the Total Chlorine reading to determine ppm combined chlorine.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

CHLORINE–BROMINE–IODINE

DPD METHOD • CODE 3643-SC

QUANTITY	CONTENTS	CODE
100	*Chlorine #1 Instrument Grade Tablets	*6903A-J
100	*Chlorine #3 Instrument Grade Tablets	*6197A-J
15 mL	Glycine Solution	6811A-E
1	Tablet Crusher	0175

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

All water for cities and communities must be sanitized; even waters that come from clean sources, protected watersheds, reservoirs, and deep wells, are commonly sanitized to assure safety. Chlorine is the most commonly used sanitizer for several reasons: it is effective against a wide range of microorganisms, the cost is low, and the methods of applying it have been well developed. If an adequate concentration of chlorine is present in the water for a few minutes, disease producing bacteria will be destroyed. A number of conditions affect the sanitizing action of chlorine. In municipal systems these can be controlled so that if chlorine is detectable, it can be assumed that bacteria have been killed. The factors that influence the rate of sanitization are temperature, pH, presence of other materials that react with chlorine, time, and the concentrations of the various chlorine combinations that are formed in the water with ammonia and other substances that react with chlorine.

The fact that chlorine can be easily detected and measured makes chlorine a favorite water sanitizer of those concerned with the public safety of water supplies. Chlorine concentrations in the range of 0.1 to 0.4 parts per million are usually maintained in municipal supplies.

Chlorine can be added in the form of chlorine gas, liquid sodium hypochlorite (bleach), granular calcium hypochlorite or as organic chlorine compounds. Chlorine is not present in natural water supplies; if it is present it is the result of chlorination of a water supply or of chlorinated compounds being discharged as waste from industrial operations. The presence of chlorine in concentrations above 0.5 parts per million should be considered evidence of pollution from chlorine treated effluents or from a process in which high concentrations of chlorine are used.

APPLICATION: Drinking, surface, and saline waters; swimming pool water; domestic and industrial wastes.

RANGE: 0.00–4.00 Chlorine

METHOD: In the absence of iodide, free available chlorine reacts instantly with DPD to produce a red color. Subsequent

addition of potassium iodide evokes a rapid color response from the combined forms of chlorine (chloramines).

SAMPLE HANDLING & PRESERVATION: Chlorine in aqueous solutions is not stable, and the chlorine content of samples or solutions, particularly weak solutions, will rapidly decrease. Exposure to sunlight or agitation will accelerate the reduction of chlorine present in such solutions. For best results, start analysis immediately after sampling. Samples to be analyzed for chlorine cannot be preserved or stored.

INTERFERENCE: The only interfering substance likely to be encountered in water is oxidized manganese. The extent of this interference can be determined by treating a sample with sodium arsenite to destroy the chlorine present so that the degree of interference can be measured.

Iodine and bromine can give a positive interference, but these are not normally present unless they have been added as sanitizers.

PROCEDURE–FREE CHLORINE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 15 Chlorine) from TESTING MENU.
5. Scroll to and select 15 Chlorine from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter and pour off all but a sufficient amount of sample water to cover a tablet. Add one *Chlorine DPD #1 Instrument Grade Tablet (6903A). Crush tablet with a tablet crusher (0175), then add sample water until tube is filled to 10 mL line. Cap tube and shake until tablet has dissolved. Solution will turn pink if free chlorine is present. Wait 15 seconds, but no longer than 30 seconds. Mix.
9. Insert tube into chamber, close lid and select SCAN SAMPLE.

PROCEDURE–COMBINED CHLORINE

10. Add one *Chlorine DPD #3 Instrument Grade Tablet (6197A) to sample from Step 8 above. Crush tablet with tablet crusher (0175). Cap tube and shake until tablet dissolves. An increase in color represents combined chlorine.
    - ☑ NOTE: For wastewater samples, *Standard Methods for the Examination of Water and Wastewater* recommends waiting 2 minutes for full color development.
11. Insert sample into chamber, close lid and select SCAN SAMPLE. Record result as Total Chlorine.
12. Subtract free chlorine reading from total chlorine reading to obtain concentration of combined chlorine.
13. Press the **OFF** button to turn off the colorimeter or press the **EXIT** button to exit to a previous menu or make another menu selection.

BROMINE

Like chlorine, bromine is an effective germicidal agent employed in drinking water treatment, pool and spa water sanitization, food service sanitation, and other public health applications.

APPLICATION: Drinking, surface, and saline waters; swimming pool water; domestic and industrial waters and wastes.

RANGE: 0.00–9.00 Bromine

METHOD: In buffered sample bromine reacts with diethyl-p-phenylene diamine (DPD) to produce a pink-red color in proportion to the concentration of bromine present.

SAMPLE HANDLING & PRESERVATION: Bromine in aqueous solutions is not stable, and the bromine content of samples or solutions, particularly weak solutions, will rapidly decrease. Exposure to sunlight or agitation will accelerate the reduction of bromine present in such solutions. For best results start analysis immediately after sampling. Samples to be analyzed for bromine cannot be preserved or stored.

INTERFERENCE: The only interfering substance likely to be encountered in water is oxidized manganese. The extent of this interference can be determined by treating a sample with sodium arsenite to destroy the bromine present so that the degree of interference can be estimated.

Iodine and chlorine can also interfere, but these are not normally present unless they have been added as sanitizers.

PROCEDURE A: BROMINE (NO CHLORINE)

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 9 Bromine-LR) from TESTING MENU.
5. Scroll to and select 9 Bromine-LR from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Pour out all but a sufficient amount of sample water to cover a tablet. Add one *Chlorine DPD #1 Instrument Grade Tablet (6903A). Crush tablet with crusher (0175), then add sample water until tube is filled to 10 mL line. Cap tube and shake until tablet is dissolved. Solution will turn pink if bromine is present. Wait 15 seconds. Mix.
9. Insert tube into chamber, close lid and select SCAN SAMPLE.
10. Press **OFF** button to turn colorimeter off or press the **EXIT** button to exit to a previous menu or make another menu selection.

PROCEDURE B:
BROMINE IN THE PRESENCE OF CHLORINE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `9 Bromine-LR`) from `TESTING MENU`.
5. Scroll to and select `9 Bromine-LR` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber close lid and select `SCAN BLANK`.
8. Rinse a second clean tube (0290) with sample water. Fill to the 10 mL line with sample. Add 5 drops of Glycine Solution (6811). Cap and mix.
9. Remove blank from colorimeter. Pour out all of the sample water. To this tube add just enough of Glycine treated sample (Step 8) to cover a tablet. Add one *Chlorine DPD#1 Instrument Grade Tablet (6903). Crush tablet with a tablet crusher (0175). Add all remaining Glycine-treated sample. Cap tube and shake until tablet dissolves. Solution will turn pink if bromine is present. Wait 15 seconds. Mix.
10. Insert tube into chamber, close lid and select `SCAN SAMPLE`.
11. Press **OFF** button to exit to previous menu or make another menu selection.

PROCEDURE C: FREE AVAILABLE, TOTAL AVAILABLE & COMBINED CHLORINE IN THE PRESENCE OF BROMINE

1. Perform the test for free and combined chlorine as previously described.
2. Perform the test for bromine in the presence of chlorine.
3. Calculations:

Residual Bromine (ppm) = Reading BR

Free Chlorine in the Presence of Bromine =
Free Chlorine – 0.45 (Reading BR)

Total Chlorine in the Presence of Bromine =
Total Chlorine – 0.45 (Reading BR)

Combined Chlorine in the Presence of Bromine =
Total Chlorine – Free Chlorine

☑ NOTE: Combined chlorine is not affected by the presence of bromine, so the calculation is the same as when only chlorine is present.

IODINE

Like chlorine and bromine, iodine is an effective germicidal agent employed in drinking water treatment, pool and spa water sanitization, food service sanitation, and other public health applications.

APPLICATION: Drinking, surface, and saline waters; swimming pool water; domestic and industrial wastes.

RANGE: 0.00–14.00 ppm Iodine

METHOD: In a buffered sample iodine reacts with diethyl-p-phenylene-diamine (DPD) to produce a pink-red color in proportion to the concentration of iodine present.

SAMPLE HANDLING & PRESERVATION: Iodine in aqueous solutions is not stable, and the iodine content of samples or solutions, particularly weak solutions, will rapidly decrease. Exposure to sunlight or agitation will accelerate the reduction of iodine present in such solutions. For best results start analysis immediately after sampling. Samples to be analyzed for iodine cannot be preserved or stored.

INTERFERENCE: The only interfering substance likely to be encountered in water is oxidized manganese. The extent of this interference can be determined by treating a sample with sodium arsenite to destroy the iodine present so that the degree of interference can be measured.

Chlorine and bromine can give a positive interference, but these are not normally present unless they have been added as sanitizers.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `50 Iodine`) from `TESTING MENU`.
5. Scroll to and select `50 Iodine` from menu.
6. Rinse a clean tube (0290) with sample water. Fill tube to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Pour off all but a sufficient amount of sample water to cover a tablet. Add one *DPD #1 Tablet Instrument Grade (6903A). Crush tablet with tablet crusher (0175). Add sample water until tube is filled to 10 mL line. Cap and shake until tablet dissolves. Solution will turn pink if iodine is present. Wait 15 seconds. Mix.
9. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

CHLORINE, FREE - UDV

DPD METHOD–UNIT DOSE VIALS • CODE 4311-H

QUANTITY	CONTENTS	CODE
1	*Free Chlorine Unit Dose Vials, 10 pouches	*4311-H

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Equipment needed but not supplied:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 6 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tip (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

All water for cities and communities must be sanitized; even waters that come from clean sources, protected watersheds, reservoirs, and deep wells, are commonly sanitized to assure safety. Chlorine is the most commonly used sanitizer for several reasons: it is effective against a wide range of microorganisms, the cost is low, and the methods of applying it have been well developed. If an adequate concentration of chlorine is present in the water for a few minutes, disease producing bacteria will be destroyed. A number of conditions affect the sanitizing action of chlorine. In municipal systems these can be controlled so that if chlorine is detectable, it can be assumed that bacteria have been killed. The factors that influence the rate of sanitization are temperature, pH, presence of other materials that react with chlorine, time, and the concentrations of the various chlorine combinations that are formed in the water with ammonia and other substances that react with chlorine.

The fact that chlorine can be easily detected and measured makes chlorine a favorite water sanitizer of those concerned with the public safety of water supplies. Chlorine concentrations in the range of 0.1 to 0.4 parts per million are usually maintained in municipal supplies.

Chlorine can be added in the form of chlorine gas, liquid sodium hypochlorite (bleach), granular calcium hypochlorite or as organic chlorine compounds. Chlorine is not present in natural water supplies; if it is present it is the result of chlorination of a water supply or of chlorinated compounds being discharged as waste from industrial operations. The presence of chlorine in concentrations above 0.5 parts per million should be considered evidence of pollution from chlorine treated effluents or from a process in which high concentrations of chlorine are used.

APPLICATION: Drinking, surface, and saline waters; swimming pool water; domestic and industrial wastes.

RANGE: 0.00–10.00 ppm Chlorine

METHOD: In the absence of iodide, free available chlorine reacts instantly with DPD to produce a red color. Subsequent addition of potassium iodide evokes a rapid color response from the combined forms of chlorine (chloramines).

SAMPLE HANDLING & PRESERVATION: Chlorine in aqueous solutions is not stable, and the chlorine content of samples or solutions, particularly weak solutions, will rapidly decrease. Exposure to sunlight or agitation will accelerate the reduction of chlorine present in such solutions. For best results, start analysis immediately after sampling. Samples to be analyzed for chlorine cannot be preserved or stored.

INTERFERENCE: The only interfering substance likely to be encountered in water is oxidized manganese. The extent of this interference can be determined by treating a sample with sodium arsenite to destroy the chlorine present so that the degree of interference can be measured.

Iodine and bromine can give a positive interference, but these are not normally present unless they have been added as sanitizers.

PROCEDURE

Use 10 mm square cell adapter.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `Testing Menu`.
4. Select `ALL TESTS` (or another sequence containing `16 Cl Free-UDV`) from `TESTING MENU`.
5. Scroll to and select `16 Cl Free-UDV` from menu.
6. Rinse a clean vial (0156) with sample water.
7. Use the syringe (1184) to add 3mL of sample to the vial.
8. Insert the vial into chamber, close the lid and select `SCAN BLANK`.
9. Remove the vial from the colorimeter.
10. Use the syringe (1184) to add 3mL of sample to a *Free Chlorine UDV vial (4311).
11. Shake vigorously until powder dissolves completely.
    - ☑ NOTE: If powder residue remains in the bottom of the vial after inverting or air bubbles form, invert once more and tap bottom of vial sharply once or twice to dislodge powder and bubbles. Mix.
12. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm free chlorine.
13. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack with indicator. When the indicator in the window turns from blue to pink, the bag should be replaced.

CHLORINE, TOTAL - UDV

DPD METHOD–UNIT DOSE VIALS • CODE 4312-H

QUANTITY	CONTENTS	CODE
1	*Total Chlorine Unit Dose Vials, 10 pouches	*4312-H

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Equipment needed but not supplied:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 6 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tip (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

All water for cities and communities must be sanitized; even waters that come from clean sources, protected watersheds, reservoirs, and deep wells, are commonly sanitized to assure safety. Chlorine is the most commonly used sanitizer for several reasons: it is effective against a wide range of microorganisms, the cost is low, and the methods of applying it have been well developed. If an adequate concentration of chlorine is present in the water for a few minutes, disease producing bacteria will be destroyed. A number of conditions affect the sanitizing action of chlorine. In municipal systems these can be controlled so that if chlorine is detectable, it can be assumed that bacteria have been killed. The factors that influence the rate of sanitization are temperature, pH, presence of other materials that react with chlorine, time, and the concentrations of the various chlorine combinations that are formed in the water with ammonia and other substances that react with chlorine.

The fact that chlorine can be easily detected and measured makes chlorine a favorite water sanitizer of those concerned with the public safety of water supplies. Chlorine concentrations in the range of 0.1 to 0.4 parts per million are usually maintained in municipal supplies.

Chlorine can be added in the form of chlorine gas, liquid sodium hypochlorite (bleach), granular calcium hypochlorite or as organic chlorine compounds. Chlorine is not present in natural water supplies; if it is present it is the result of chlorination of a water supply or of chlorinated compounds being discharged as waste from industrial operations. The presence of chlorine in concentrations above 0.5 parts per million should be considered evidence of pollution from chlorine treated effluents or from a process in which high concentrations of chlorine are used.

APPLICATION: Drinking, surface, and saline waters; swimming pool water; domestic and industrial wastes.

RANGE: 0.00–10.00 ppm Chlorine

METHOD: In the absence of iodide, free available chlorine reacts instantly with DPD to produce a red color. Subsequent addition of potassium iodide evokes a rapid color response from the combined forms of chlorine (chloramines).

SAMPLE HANDLING & PRESERVATION: Chlorine in aqueous solutions is not stable, and the chlorine content of samples or solutions, particularly weak solutions, will rapidly decrease. Exposure to sunlight or agitation will accelerate the reduction of chlorine present in such solutions. For best results, start analysis immediately after sampling. Samples to be analyzed for chlorine cannot be preserved or stored.

INTERFERENCE: The only interfering substance likely to be encountered in water is oxidized manganese. The extent of this interference can be determined by treating a sample with sodium arsenite to destroy the chlorine present so that the degree of interference can be measured.

Iodine and bromine can give a positive interference, but these are not normally present unless they have been added as sanitizers.

PROCEDURE

Use 10 mm square cell adapter.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select Testing Menu.
4. Select `ALL TESTS` (or another sequence containing `18 Cl Total-UDV`) from `TESTING MENU`.
5. Scroll to and select `18 Cl Total-UDV` from menu.
6. Rinse a clean vial (0156) with sample water.
7. Use the syringe (1184) to add 3mL of sample to the vial.
8. Insert the vial into chamber, close the lid and select `SCAN BLANK`.
9. Remove the vial from the colorimeter.
10. Use the syringe (1184) to add 3mL of sample to a *Total Chlorine UDV vial (4312).
11. Shake vigorously until powder dissolves completely.
    - ☑ NOTE: If powder residue remains in the bottom of the vial after inverting or air bubbles form, invert once more and tap bottom of vial sharply once or twice to dislodge powder and bubbles. Mix.
12. Wait 2 minutes.
13. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm total chlorine.
14. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack with indicator. When the indicator in the window turns from blue to pink, the bag should be replaced.

CHLORINE DIOXIDE

DPD METHOD • CODE 3644-SC

QUANTITY	CONTENTS	CODE
100	*Chlorine #1 Instrument Grade Tablets	*6903A-J
15 mL	Glycine Solution	6811-E
1	Tablet Crusher	0175

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Chlorine dioxide is used as a substitute for and an adjunct to chlorine in water treatment. It is better than chlorine in eliminating taste and odor in certain cases. Chlorine dioxide, unlike chlorine, does not produce carcinogenic chlorinated organic compounds when reacted with organic materials. A disadvantage is the higher cost of producing chlorine dioxide compared to chlorine.

APPLICATION: Drinking and pool waters; domestic and industrial wastewater; food sanitization.

RANGE: 0.00–8.00 Chlorine Dioxide

METHOD: Chlorine dioxide reacts with DPD to form a red color in proportion to the concentration.

SAMPLE HANDLING & PRESERVATION: Test as soon as possible to avoid loss of chlorine dioxide.

INTERFERENCE: Chlorine interference can be removed with the use of glycine. Very high levels of chloramines may interfere if the test result is not read immediately. Oxidized manganese interferes but can be removed with arsenite. Bromine and iodine interfere. Chromate interference can be removed with a thioacetamide blank correction.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 20 CHLOR DIOX) from TESTING MENU.
5. Scroll to and select 20 CHLOR DIOX from menu.
6. Rinse a clean tube (0290) with sample water. Fill to 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Pour out all but a sufficient amount of sample water to cover tablet. Add 5 drops of Glycine Solution (6811).
9. Add one *Chlorine DPD #1 Instrument Grade Tablet (6903A). Crush tablet with tablet crusher. Cap and shake until tablet dissolves. Fill to 10 mL line with sample water. Solution will turn pink if chlorine dioxide is present. Wait 15 seconds, but no longer than 30 seconds. Mix.
10. Insert tube into chamber, close lid and select SCAN SAMPLE.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

CHROMIUM

DIPHENYLCARBOHYDRAZIDE METHOD • CODE 3697-SC

QUANTITY	CONTENTS	CODE
50	*Chromium Spectrophotometric Grade Tablets	*3889A-H
1	Tablet Crusher	0175

***WARNING:** Reagents marked with a * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact LaMotte by email, phone or fax.

Chromium is one of a class of heavy metals found in the bottom mud of polluted bodies of water. It is considered to be a toxic chemical. Chromium will become concentrated in some shellfish, endangering the health of the human or animal that consumes them. Chromium may be present in water containing waste from industries such as metal plating. If more than 0.5 ppm chromium is present, it is evidence of contamination from untreated or incompletely treated industrial waste.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastewaters.

RANGE: 0.00–1.00 ppm Chromium

METHOD: Hexavalent chromium reacts with 1,5 diphenylcarbohydrazide under acidic conditions to form a red-purple color in proportion to the amount of chromium present.

SAMPLE HANDLING & PRESERVATION: Analysis for chromium should be made as quickly as possible. Storage in plastic or glass containers may result in low results.

INTERFERENCES: High concentrations of mercurous and mercuric ions may impart a blue color to the chromium determination. Iron and vanadium in concentrations above 1 ppm may result in a yellow color.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 23 Chromium-TT) from TESTING MENU.
5. Scroll to and select 23 Chromium-TT from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL line with sample.
7. Insert the tube into chamber, close lid and select SCAN BLANK.
8. Remove the tube from colorimeter.
9. Add one *Chromium Spectrophotometric Grade Tablet (3889A).
10. Use Tablet Crusher (0175) to crush tablet.
11. Cap tube.
12. Shake vigorously for 30 seconds.
13. Wait 3 minutes.
14. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result in ppm chromium.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples.

To convert results to ppm chromate (CrO_4^{-2}), multiply by 2.23. To convert result to ppm sodium chromate (Na_2CrO_4) multiply by 3.12.

CHROMIUM-HEXAVALENT

DIPHENYLCARBOHYDRAZIDE METHOD • CODE 3645-SC

QUANTITY	CONTENTS	CODE
10 g	*Chromium Reagent Powder	*V-6276-D
1	Spoon, 0.1 g, plastic	0699
50	Filter Paper	0465-H
1	Funnel, Plastic	0459

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Chromium may be present in water containing waste from industries such as metal plating. It is considered to be a toxic chemical and, if present in an amount of over 0.5 ppm, is evidence of contamination from untreated or incompletely treated industrial waste.

Chromium is one of a class of heavy metals found in the bottom mud of polluted bodies of water. Certain shellfish are capable of concentrating this element, endangering the health of its ultimate consumer, human or animal.

APPLICATION: Drinking, surface, & saline waters; domestic and industrial wastewaters.

RANGE: 0.00–1.00 Chromium

METHOD: Hexavalent chromium reacts with 1,5 diphenylcarbohydrazide under acidic conditions to form a red-purple color in proportion to the amount of chromium present.

SAMPLE HANDLING & PRESERVATION: Analysis for chromium should be made as quickly as possible after sample collection since storage in glass or plastic containers may result in low chromate values.

INTERFERENCES: High concentrations of mercurous and mercuric ions may impart a blue color to the chromium determination. Iron and vanadium in concentrations above 1 mg/L may result in a yellow color. However, the vanadium color becomes negligible 10 minutes after the addition of diphenylcarbohydrazide.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `22 Chromium`) from `TESTING MENU`.
5. Scroll to and select `22 Chromium` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Use the 0.1g spoon (0699) to add one measure of *Chromium Reagent Powder (V-6276). Cap and shake until powder dissolves. Wait 3 minutes for full color development.
9. During waiting period, fold a piece of filter paper (0465) in half then half again to form a cone. Push corners together to open end, and insert into funnel (0459).
10. At the end of 3 minute waiting period, filter sample into a clean tube. Mix. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: To convert result to ppm chromate (CrO_4^{2-}) multiply by 2.23. To convert result to ppm sodium chromate (Na_2CrO_4) multiply by 3.12.

Highly buffered waters may give poor results and require a more careful pH adjustment. Before adding *Chromium Reagent Powder, adjust pH of sample to pH 3–4.

CHROMIUM–HEXAVALENT, TRIVALENT & TOTAL

DIPHENYLCARBOHYDRAZIDE METHOD • CODE 3698-SC

QUANTITY	CONTENTS	CODE
60 mL	*Sulfuric Acid, 5N	*7681-H
10 g	*Chromium Reagent Powder	*V-6276-D
15 mL	*Sodium Azide, 5%	*7683-E
30 mL	Potassium Permanganate, 0.5%	7682-G
60 mL	Deionized Water	5115PT-H
1	Pipet, plain, glass, w/cap	0341
1	Pipet, 1.0 mL, plastic	0354
1	Spoon, 0.1 g, plastic	0699
1	Graduated Cylinder, 50 mL, glass	0418
1	Erlenmeyer Flask, 125 mL, glass	0431
1	Test tube holder	1113
1	Filter Paper	0465
1	Funnel, Plastic	0459

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

A toxic chemical, chromium is found in two forms in the water; trivalent chromium (Cr^{3+}) and hexavalent chromium (Cr^{6+}). Chromium enters the water from industrial waste. Hexavalent chromium is more toxic than trivalent chromium. Levels greater than 0.5 ppm indicate improperly treated industrial waste. It is important to maintain chromium levels at or below 0.5 ppm, because clams and other shellfish will store chromium in their systems, accumulating levels which may be dangerous to the consumer, whether human or animal.

APPLICATION: Drinking, surface, & saline water; domestic and industrial waste.

RANGE: 0.00–1.00 Chromium

METHOD: The trivalent chromium is converted to hexavalent chromium by permanganate under acidic conditions. Hexavalent chromium reacts with 1,5 diphenylcarbohydrazide under acidic conditions to form a red-purple color in proportion to the amount of chromium present.

SAMPLE HANDLING & PRESERVATION:	Analysis for chromium should be made as quickly as possible after sample collection since storage in glass or plastic containers may result in low chromate values.
INTERFERENCES:	High concentrations of mercurous and mercuric ions may interfere.

HEXAVALENT CHROMIUM PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 22 Chromium) from TESTING MENU.
5. Scroll to and select 22 Chromium from menu.
6. Rinse a clean tube (0290) with sample water. Fill to 10 mL line with sample water.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Use 0.1 g spoon (0699) to add one level measure of *Chromium Reagent Powder (V-6276). Cap and shake for one minute. Wait 3 minutes.
9. During the waiting period, fold a piece of filter paper in half, then in half again to form a cone. Push corners together to open end, and insert into funnel (0459).
10. At the end of 3 minute waiting period, filter sample into a clean tube (0290). Cap and mix. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

TOTAL CHROMIUM WITH ACID DIGESTION PROCEDURE

1. Fill graduated cylinder (0418) to 50 mL line with sample water. Transfer to Erlenmeyer flask (0431).
2. Use the 1 mL pipet (0354) to add 5 mL (five measures) of *Sulfuric Acid, 5N (7681). Swirl to mix.

   ☑ NOTE: Highly buffered waters may require pH adjustment. Adjust the pH of highly buffered samples to 7.0 ±0.5. Continue procedure.
3. Place flask on burner or hot plate. Bring solution to a gentle boil.
4. Fill pipet (0341) with Potassium Permanganate, 0.5% (7682). While gently swirling flask, add Potassium Permanganate, 0.5% (7682), 2 drops at a time to boiling solution, until solution turns a dark pink color which persists for 10 minutes. Continue boiling.
5. Add one drop of *Sodium Azide, 5% (7683) to boiling solution. Boil for approximately 30 seconds. If pink color does not fade, add another drop of *Sodium Azide, 5%. Continue adding *Sodium Azide, 5% one drop at a time until pink color disappears.
6. Remove flask from heat. Cool sample under running water. This is the digested sample.
7. Pour digested sample into clean graduated cylinder (0418). Dilute to the 50 mL line with Deionized Water (5115).
8. Press and hold **ON** button until colorimeter turns on.
9. Press **ENTER** to start.
10. Press **ENTER** to select `TESTING MENU`.
11. Select `ALL TESTS` or another sequence containing `22 Chromium`) from `TESTING MENU`.
12. Scroll to and select `22 Chromium` from menu.
13. Rinse a clean tube (0290) with sample water. Fill to 10 mL line with sample water.
14. Insert tube into chamber, close lid and select `SCAN BLANK`.
15. Remove tube from colorimeter. Use 0.1 g spoon (0699) to add one level measure of *Chromium Reagent Powder (V-6276). Cap and shake for one minute. Wait 3 minutes.
16. During the waiting period, fold a piece of filter paper in half, then in half again to form a cone. Push corners together to open end, and insert into funnel (0459).
17. Filter sample into a clean tube (0290). Cap and mix. Insert tube of filtered sample into chamber, close lid and select `SCAN SAMPLE`. Record result.
18. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

TRIVALENT CHROMIUM PROCEDURE

Subtract hexavalent chromium from total chromium. Record as ppm trivalent chromium.

Trivalent Chromium = Total Chromium – Hexavalent Chromium

COBALT

PAN METHOD • CODE 4851

QUANTITY	CONTENTS	CODE
60 mL	*Cobalt Buffer	*4852-H
60 mL	*Cobalt Indicator Reagent	*4853-H
30 mL	*Stabilizer Solution	*4854-G
2	Pipet, 1.0 mL, plastic	0354
1	Pipet, 0.5 mL, plastic	0353

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Cobalt rarely occurs in natural water. It is used in the manufacture of alloys to increase corrosion resistance and strength. It is found in wastewaters as a corrosion by-product.

APPLICATION: Industrial wastewater.

RANGE: 0.00–2.00 Cobalt

METHOD: PAN (1-[2-Pyridylazo]-2-Naphthol) forms a greenish complex with Cobalt (Co^{+2}) at a pH of 5.

SAMPLE HANDLING & PRESERVATION: Store samples in acid-washed plastic bottles. Adjust pH to less than 2 with nitric acid. Adjust sample pH to 5 before testing.

INTERFERENCES: Iron (+2) and high concentrations of heavy metals.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `All Tests` (or another sequence containing `24 Cobalt`) from `TESTNG MENU`.
5. Scroll to and select `24 Cobalt` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Use the 1.0 mL pipet (0354) to add 1 mL of *Cobalt Buffer (4852). Cap and mix.
10. Use the other 1.0 mL pipet (0354) to add 1 mL of *Cobalt Indicator Reagent (4853). Cap and mix.
11. Wait 3 minutes.
12. Use the 0.5 mL pipet (0353) to add 0.5 mL *Stabilizer Solution (4854). Cap and invert 15 times to thoroughly mix.
13. Wait 5 minutes. DO NOT MIX.
14. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm cobalt.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

COD–LOW RANGE

MERCURY FREE DIGESTION • CODE 0072-SC
MERCURY DIGESTION • CODE 0075-SC

QUANTITY	CONTENTS	CODE
25	*COD Low Range Mercury Free Tubes	*0072-SC
or 25	*COD Low Range Mercury Tubes	*0075-SC

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

COD Low Range Mercury Free Tubes are not USEPA approved.

COD Low Range Mercury Tubes are USEPA approved.

Equipment needed but not supplied:

QUANTITY	CONTENTS	CODE
1	COD Adapter	5-0087
1	COD Reactor, 12 vial, 110V	5-0102
or 1	COD Reactor, 12 vial, 230V	5-0102-EX2
1	Measuring Pipet, 1.0 mL	2-2110
1	Pipet Bulb	2-2164

Chemical Oxygen Demand (COD) is a measure of the amount of organic matter in water which is susceptible to oxidation by chemical oxidants. COD can be empirically related to the Biological Oxygen Demand (BOD) and organic carbon content of a specific source of water. This correlation must be determined experimentally for each source of water.

APPLICATION: Domestic and industrial wastes.

RANGE: 5–150 mg/L COD

METHOD: Dichromate in the presence of silver salts, at high temperature in a closed system, oxidizes most organic compounds to 95-100% of the theoretical amount. This process is called digestion. As dichromate oxidizes the organic compounds, the amount of yellow color is reduced. The remaining yellow color is measured colorimetrically at the 420 nm and is directly proportional to the COD of the sample.

SAMPLE HANDLING & PRESERVATION: Collect samples in glass and test as soon as possible. If samples must be stored, preservation is accomplished by the addition of concentrated H_2SO_4 to adjust the pH below 2. Samples with suspended solids should be homogenized in a

blender (100 mL for 30 seconds) and then stirred gently with a magnetic stirrer.

INTERFERENCES: Volatile organic compounds are not oxidized to the extent that they are in the vapor above the digestion solution. Therefore, they do not contribute to the COD reading. Chloride concentrations above 10% of COD interfere with the mercury free tubes. Chloride above 2000 ppm will interfere with the mercury tubes. Nitrite gives a positive interference of 1.1 ppm O_2 per ppm NO_2–N which is insignificant unless nitrite concentrations are very high. Other reduced inorganic compounds are stoichiometrically oxidized, causing a positive interference. Corrections can be made for these compounds based upon their stoichiometry and concentrations.

When scanning samples in 16 mm tubes, such as COD, the sample chamber lid can not be closed. Use the COD adapter to minimize stray light interference. To further reduce stray light interference, do not scan sample in direct sunlight.

PROCEDURE

Use COD adapter (see p. 22).

1. Homogenize sample if necessary.
2. Preheat COD heater block to 150±2°C.
3. Remove cap from COD tube vial. Hold vial at a 45° angle. Use a volumetric pipet, to carefully add 2.0 mL sample water allowing the sample to run down the side of the vial.
4. Cap and mix thoroughly.
5. Rinse the outside of the vial with distilled water. Wipe dry with a paper towel.
6. Repeat steps 3 through 5 using 2.0 mL distilled water. This is the reagent blank.
7. Place vials in preheated COD block heater and maintain temperature at 150±2°C for two hours.
8. At the end of the heating period turn the heater off. Wait 20 minutes for the vials to cool to 120°C or less.
9. Remove vials from block heater. Invert several times to mix.
10. Allow to cool to room temperature.
11. Press and hold **ON** button until colorimeter turns on.
12. Press **ENTER** to start.
13. Press **ENTER** to select `TESTING MENU`.
14. Select `ALL TESTS` (or another sequence containing `25 COD LR`) from `PROGRAMMED TESTS` menu.
15. Scroll to and select `25 COD LR` from menu.
16. Wipe the blank vial with a damp towel to remove fingerprints and smudges. Wipe with a dry towel.
17. Insert reagent blank tube into chamber. Select `SCAN BLANK`.
18. Remove tube from colorimeter.
19. Insert digested water sample tube into chamber. Select `SCAN SAMPLE`. Record result. For the most accurate results, take three readings on each sample and average the results.
20. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ **NOTES:** Reagents are light sensitive. Unused reagents should be stored in the shipping container, and in the refrigerator if possible, until needed.

A reagent blank should be run with each set of samples and with each lot of reagents.

The reacted blank will be stable if stored in the dark.

To eliminate error caused by contamination, wash all glassware with 20% sulfuric acid.

For greater accuracy, a minimum of three repetitions should be performed and the results averaged.

Some samples may be digested completely in less than two hours. The concentration may be measured at 15 minute intervals while the vials are still hot until the reading remains unchanged. The vials should be cooled to room temperature before the final measurement is taken.

COD–STANDARD RANGE

MERCURY FREE DIGESTION • CODE 0073-SC
MERCURY DIGESTION • CODE 0076-SC

QUANTITY	CONTENTS	CODE
25	*COD Standard Range Mercury Free Tubes	*0073-SC
or 25	*COD Standard Range Mercury Tubes	*0076-SC

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

COD Standard Range Mercury Free Tubes are not USEPA approved.

COD Standard Range Mercury Tubes are USEPA approved.

Equipment needed but not supplied:

QUANTITY	CONTENTS	CODE
1	COD Adapter	5-0087
1	COD Reactor, 12 vial, 110V	5-0102
or 1	COD Reactor, 12 vial, 230V	5-0102-EX2
1	Measuring Pipet, 1.0 mL	2-2110
1	Pipet Bulb	2-2164

Chemical Oxygen Demand (COD) is a measure of the amount of organic matter in water which is susceptible to oxidation by chemical oxidants. COD can be empirically related to the Biological Oxygen Demand (BOD) and organic carbon content of a specific source of water. This correlation must be determined experimentally for each source of water.

APPLICATION: Domestic and industrial wastes.

RANGE: 0–1500 mg/L COD

METHOD: Dichromate in the presence of silver salts, at high temperature in a closed system, oxidizes most organic compounds to 95-100% of the theoretical amount. This process is called digestion. As dichromate oxidizes the organic compounds, a green complex is formed. The concentration of the green complex is measured at 605 nm and is directly proportional to the COD of the sample.

SAMPLE HANDLING & PRESERVATION: Collect samples in glass and test as soon as possible. If samples must be stored, preservation is accomplished by the addition of concentrated H_2SO_4 to adjust the pH below 2. Samples with suspended solids should be homogenized in a blender (100 mL for 30 seconds) and then stirred gently with a magnetic stirrer.

INTERFERENCES: Volatile organic compounds are not oxidized to the extent that they are in the vapor above the digestion solution. Therefore, they do not contribute to the COD reading. Chloride concentrations above 10% of COD interfere with the mercury free tubes. Chloride above 2000 ppm will interfere with the mercury tubes. Nitrite gives a positive interference of 1.1 ppm O_2 per ppm NO_2–N which is insignificant unless nitrite concentrations are very high. Other reduced inorganic compounds are stoichiometrically oxidized, causing a positive interference. Corrections can be made for these compounds based upon their stoichiometry and concentrations.

When scanning samples in 16 mm tubes, such as COD, the sample chamber lid can not be closed. Use the COD adapter to minimize stray light interference. To further reduce stray light interference, do not scan sample in direct sunlight.

PROCEDURE

Use COD adapter (see p. 22).

1. Homogenize sample if necessary.
2. Preheat COD heater block to 150±2°C.
3. Remove cap from COD tube vial. Hold vial at a 45° angle. Use a volumetric pipet, to carefully add 2.0 mL sample water allowing the sample to run down the side of the vial.
4. Cap and mix thoroughly.
5. Rinse the outside of the vial with distilled water. Wipe dry with a paper towel.
6. Repeat steps 2 through 5 using 2.0 mL distilled water. This is the reagent blank.
7. Place vials in preheated COD block heater and maintain temperature at 150±2°C for two hours.
8. At the end of the heating period turn the heater off. Wait 20 minutes for the vials to cool to 120°C or less.
9. Remove vials from block heater. Invert several times to mix.
10. Allow to cool to room temperature.
11. Press and hold **ON** button until colorimeter turns on.
12. Press **ENTER** to start.
13. Press **ENTER** to select TESTING MENU.
14. Select ALL TESTS (or another sequence containing 26 COD SR) from PROGRAMMED TESTS menu.
15. Wipe the blank vial with a damp towel to remove fingerprints and smudges. Wipe with a dry towel.
16. Scroll to and select 26 COD SR from menu.
17. Insert reagent blank tube into chamber. Select SCAN BLANK.
18. Remove tube from colorimeter.
19. Insert digested water sample tube into chamber. Select SCAN SAMPLE. Record result. For the most accurate results, take three readings on each sample and average the results.
20. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: Reagents are light sensitive. Unused reagents should be stored in the shipping container, and in the refrigerator if possible, until needed.

A reagent blank should be run with each set of samples and with each lot of reagents.

The reacted blank will be stable if stored in the dark.

To eliminate error caused by contamination, wash all glassware with 20% sulfuric acid.

For greater accuracy, a minimum of three repetitions should be performed and the results averaged.

Some samples may be digested completely in less than two hours. The concentration may be measured at 15 minute intervals while the vials are still hot until the reading remains unchanged. The vials should be cooled to room temperature before the final measurement is taken.

COD–HIGH RANGE

MERCURY FREE DIGESTION • CODE 0074-SC
MERCURY DIGESTION • CODE 0077-SC

QUANTITY	CONTENTS	CODE
25	*COD High Range Mercury Free Tubes	*0074-SC
or 25	*COD High Range Mercury Tubes	*0077-SC

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

COD High Range Mercury Free Tubes and COD High Range Mercury Tubes are not USEPA approved.

Equipment needed but not supplied:

QUANTITY	CONTENTS	CODE
1	COD Adapter	5-0087
1	COD Reactor, 12 vial, 110V	5-0102
or 1	COD Reactor, 12 vial, 230V	5-0102-EX2
1	Measuring Pipet, 1.0 mL	2-2110
1	Pipet Bulb	2-2164

Chemical Oxygen Demand (COD) is a measure of the amount of organic matter in water which is susceptible to oxidation by chemical oxidants. COD can be empirically related to the Biological Oxygen Demand (BOD) and organic carbon content of a specific source of water. This correlation must be determined experimentally for each source of water.

APPLICATION: Domestic and industrial wastes.

RANGE: 0–15000 mg/L COD

METHOD: Dichromate in the presence of silver salts, at high temperature in a closed system, oxidizes most organic compounds to 95-100% of the theoretical amount. This process is called digestion. As dichromate oxidizes the organic compounds, a green complex is formed. The concentration of the green complex is measured at 605 nm and is directly proportional to the COD of the sample.

SAMPLE HANDLING & PRESERVATION: Collect samples in glass and test as soon as possible. If samples must be stored, preservation is accomplished by the addition of concentrated H_2SO_4 to adjust the pH below 2. Samples with suspended solids should be homogenized in a blender (100 mL for 30 seconds) and then stirred gently with a magnetic stirrer.

INTERFERENCES: Volatile organic compounds are not oxidized to the extent that they are in the vapor above the digestion solution. Therefore, they do not contribute to the COD reading. Contains mercury sulfate to prevent interference from chloride. Nitrite gives a positive interference of 1.1 ppm O_2 per ppm NO_2–N, which is insignificant unless nitrite concentrations are very high. Other reduced inorganic compounds are stoichiometrically oxidized, causing a positive interference. Corrections can be made for these compounds based upon their stoichiometry and concentrations.

When scanning samples in 16 mm tubes, such as COD, the sample chamber lid can not be closed. Use the COD adapter to minimize stray light interference. To further reduce stray light interference, do not scan sample in direct sunlight.

PROCEDURE

Use COD adapter (see p. 22).

1. Homogenize sample if necessary.
2. Preheat COD heater block to 150±2°C.
3. Remove cap from COD tube vial. Hold vial at a 45° angle. Use a graduated pipet, to carefully add 0.2 mL sample water allowing the sample to run down the side of the vial.
4. Cap and mix thoroughly.
5. Rinse the outside of the vial with distilled water. Wipe dry with a paper towel.
6. Repeat steps 3 through 5 using 0.2 mL distilled water. This is the reagent blank.
7. Place vials in preheated COD block heater and maintain temperature at 150±2°C for two hours.
8. At the end of the heating period turn the heater off. Wait 20 minutes for the vials to cool to 120°C or less.
9. Remove vials from block heater. Invert several times to mix.
10. Allow to cool to room temperature.
11. Press and hold **ON** button until colorimeter turns on.
12. Press **ENTER** to start.
13. Press **ENTER** to select `TESTING MENU`.
14. Select `ALL TESTS` (or another sequence containing `27 COD HR`) from `PROGRAMMED TESTS` menu.
15. Wipe the blank vial with a damp towel to remove fingerprints and smudges. Wipe with a dry towel.
16. Scroll to and select `27 COD HR` from menu.
17. Insert reagent blank tube into chamber. Select `SCAN BLANK`.
18. Remove tube from colorimeter.
19. Insert digested water sample tube into chamber. Select `SCAN SAMPLE`. Record result. For the most accurate results, take three readings on each sample and average the results.
20. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: Reagents are light sensitive. Unused reagents should be stored in the shipping container, and in the refrigerator if possible, until needed.

A reagent blank should be run with each set of samples and with each lot of reagents.

The reacted blank will be stable if stored in the dark.

To eliminate error caused by contamination, wash all glassware with 20% sulfuric acid.

For greater accuracy, a minimum of three repetitions should be performed and the results averaged.

COLOR

PLATINUM COBALT METHOD • NO REAGENTS REQUIRED

Color in water may be attributed to humus, peat, plankton, vegetation, and natural metallic ions, such as iron and manganese, or industrial waste. Color is removed to make water suitable for domestic and industrial use. Color may have to be removed from industrial waste before it is discharged to a waterway.

APPLICATION:	Potable water and water with color due to natural materials.
RANGE:	0–1000 color units
METHOD:	Color is determined by a meter that has been calibrated with colored standards of known platinum cobalt concentration. True color, the color of water in which the turbidity has been removed, is measured.
SAMPLE HANDLING & PRESERVATION:	Collect all samples in clean glassware. Determine color as soon as possible to avoid biological or chemical changes that could occur in the sample during storage.
INTERFERENCES:	Turbidity will interfere. Filter before testing.

PROCEDURE

1. Press and hold **ON** burton until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `28 Color`) from `TESTING MENU`.
5. Scroll to and select `28 Color` from menu.
6. Rinse a tube (0290) with color-free water (distilled or deionized water). Fill to 10 mL line with color-free water.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Empty tube.
9. Rinse tube with sample water. Fill to 10 mL line with water sample.
10. Insert tube with sample water, close lid and select `SCAN SAMPLE`. Record result in color units.
11. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

COPPER–LOW RANGE

BICINCHONINIC ACID METHOD • CODE 3640-SC

QUANTITY	CONTENTS	CODE
50	*Copper Tablets	*T-3808-H

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

The copper content of drinking water generally falls below 0.03 parts per million, but copper levels as high as 1.0 part per million will give water a bitter taste. Waters testing as high as 1.0 part per million copper have probably been treated with a copper compound, like those used in the control of algae, or have become contaminated from untreated industrial wastes. The addition of copper sulfate to lakes causes an increase in the copper content of the sediments. Acid waters and those high in free carbon dioxide may cause the corrosion or "eating away" of copper, brass and bronze pipes and fittings. This corrosion results in the addition of copper into the water supply.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastes.

RANGE: 0.00–3.50 ppm Copper

METHOD: Copper ions form a purple complex with bicinchoninic acid around pH 6-7, in proportion to the concentration of copper in the sample.

SAMPLE HANDLING & PRESERVATION: Copper has a tendency to be adsorbed to the surface of the sample container. Samples should be analyzed as soon as possible after collection. If storage is necessary, 0.5 mL of 20% HCl per 100 mL of sample will prevent "plating out." However, a correction must be made to bring the reaction into the optimum pH range.

INTERFERENCES: High concentrations of oxidizing agents, calcium, and magnesium interfere. Silver can also interfere.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 29 Copper BCA-LR) from TESTING MENU.
5. Scroll to and select 29 Copper BCA-LR from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter and add one *Copper Tablet (T-3808). Cap and shake vigorously until tablet dissolves. Solution will turn purple if copper is present. Wait 2 minutes.
9. At end of 2 minute waiting period, mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

COPPER

CUPRIZONE METHOD • CODE 4023

QUANTITY	CONTENTS	CODE
15 mL	Copper A	P-6367-E
15 mL	*Copper B	*P-6368-E

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

The copper content of drinking water generally falls below 0.03 parts per million, but copper levels as high as 1.0 part per million will give water a bitter taste. Waters testing as high as 1.0 part per million copper have probably been treated with a copper compound, like those used in the control of algae, or have become contaminated from untreated industrial wastes. The addition of copper sulfate to lakes causes an increase in the copper content of the sediments. Acid waters and those high in free carbon dioxide may cause the corrosion or "eating away" of copper, brass and bronze pipes and fittings. This corrosion results in the addition of copper to the water supply.

APPLICATION: Drinking, surface, and domestic waters. Pools and spas.

RANGE: 0.00–2.00 ppm Copper

METHOD: Copper ions form a blue complex with cuprizone, in a 1 to 2 ratio, at a pH of about 8, in proportion to the concentration of copper in the sample.

SAMPLE HANDLING & PRESERVATION: Copper has a tendency to be adsorbed to the surface of the sample container. Samples should be analyzed as soon as possible after collection. If storage is necessary, 0.5 mL of 20% hydrochloric acid per 100 mL of sample will prevent "plating out". However, a correction must be made to bring the reaction into the optimum pH range.

INTERFERENCES: Hg^{+1} at 1 ppm. Cr^{+3}, Co^{+2}, and silicate at 10 ppm. As^{+3}, Bi^{+3}, Ca^{+2}, Ce^{+3}, Ce^{+4}, Hg^{+2}, Fe^{+2}, Mn^{+2}, Ni^{+2} and ascorbate at 100 ppm.

Many other metal cations and inorganic anions at 1000 ppm. EDTA at all concentrations.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `31 Cu-Cuprizone`) from `TESTING MENU`.
5. Scroll to and select `31 Cu-Cuprizone` from menu.
6. Rinse a tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter and add 5 drops of Copper A (6367). Cap and mix.
9. Add 5 drops of *Copper B (6368). Cap and mix.
10. Wait 5 minutes. Mix.
11. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
12. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

The reaction may stain the tubes. Scrub tubes thoroughly after each use.

COPPER

DIETHYLDITHIOCARBAMATE METHOD • CODE 3646-SC

QUANTITY	CONTENTS	CODE
15 mL	*Copper 1	*6446-E

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

The copper content of drinking water generally falls below 0.03 parts per million, but copper levels as high as 1.0 part per million will give water a bitter taste. Waters testing as high as 1.0 part per million copper have probably been treated with a copper compound, like those used in the control of algae, or have become contaminated from untreated industrial wastes. The addition of copper sulfate to lakes causes an increase in the copper content of the sediments. Acid waters and those high in free carbon dioxide may cause the corrosion or "eating away" of copper, brass and bronze pies and fittings. This corrosion results in the addition of copper into the water supply.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastes.

RANGE: 0.00–6.00 ppm Copper

METHOD: Copper ions form a yellow colored chelate with diethyldithiocarbamate around pH 9-10 in proportion to the concentration of copper in the sample.

SAMPLE HANDLING & PRESERVATION: Copper has a tendency to be adsorbed to the surface of the sample container. Samples should be analyzed as soon as possible after collection. If storage is necessary, 0.5 mL of 20% hydrochloric acid per 100 mL of sample will prevent "plating out." However, a correction must be made to bring the reaction into the optimum pH range.

INTERFERENCES: Bismuth, cobalt, mercurous, nickel and silver ions and chlorine (6 ppm or greater) interfere and must be absent.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence contining `32 Copper DDC`) from `TESTING MENU`.
5. Scroll to and select `32 Copper DDC` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter and add 5 drops of *Copper 1 (6446). Cap and mix. Solution will turn yellow if copper is present.
9. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: The reaction may stain the tubes. Scrub the tubes thoroughly after each use.

COPPER–UDV

BICINCHONINIC ACID METHOD–UNIT DOSE VIALS
CODE 4314-H

QUANTITY	CONTENTS	CODE
1	Copper Unit Dose Vials, 10 pouches	4314-H

Equipment needed but not suppled:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 3 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tip (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

The copper content of drinking water generally falls below 0.03 parts per million, but copper levels as high as 1.0 part per million will give water a bitter taste. Waters testing as high as 1.0 part per million copper have probably been treated with a copper compound, like those used in the control of algae, or have become contaminated from untreated industrial wastes. The addition of copper sulfate to lakes causes an increase in the copper content of the sediments. Acid waters and those high in free carbon dioxide may cause the corrosion or "eating away" of copper, brass and bronze pipes and fittings. This corrosion results in the addition of copper to the water supply.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastes.

RANGE: 0.0–4.0 ppm Copper

METHOD: Cupric ions form a purple complex with bicinchoninic acid around pH 6–7, in proportion to the concentration of copper in the sample.

SAMPLE HANDLING & PRESERVATION: Copper has a tendency to be adsorbed to the surface of the sample container. Samples should be analyzed as soon as possible after collection. If storage is necessary, 0.5 mL of 20% hydrochloric acid per 100 mL of sample will prevent "plating out". However, a correction must be made to bring the reaction into the optimum pH range.

INTERFERENCES: High concentrations of oxidizing agents, calcium, and magnesium interfere. Silver can also interfere.

PROCEDURE

Use 10 mm square cell adapter.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
3. Select ALL TESTS (or another sequence containing 33 Copper-UDV) from TESTING MENU.
4. Scroll to and select 33 Copper-UDV from menu.
5. Rinse a clean vial (0156) with sample water.
6. Use the syringe (1184) to add 3 mL of sample to the vial.
7. Insert the vial into chamber, close lid and select SCAN BLANK.
8. Remove vial from the colorimeter.
9. Use the syringe (1184) to add 3 mL of sample to a Copper UDV vial (4314).
10. Wait 2 minutes.
11. Invert vial 3 times to mix.
    - ☑ NOTE: If powder residue remains in the bottom of the vial after inverting, or if air bubbles form, invert once more and tap bottom of vial sharply once or twice to dislodge powder or bubbles. Mix.
12. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
13. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack with indicator. When the indicator in the window turns from blue to pink, the bag should be replaced.

CYANIDE

PYRIDINE-BARBITURIC ACID METHOD • CODE 3660-SC

QUANTITY	CONTENTS	CODE
60 mL	Cyanide Buffer	2850PS-H
5 g	*Cyanide Cl Reagent	*2794DS-C
5 g	*Cyanide Indicator Reagent	*2793DS-C
15 mL	*Hydrochloric Acid 1N	*6130-E
15 mL	*Sodium Hydroxide 1N	*4004-E
2	Spoons, 0.1 g, plastic	0699
1	Pipet, plastic, 1.0 mL	0354
1	pH Short Range Test Paper, pH 9–14	2955
1	Stirring Rod, Plastic	0519

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

The presence of cyanide in water has a significant effect on the biological activity of the system. Cyanides may exist in water in a variety of forms which vary in toxicity. Cyanide is a by-product of industrial waste from petroleum refining and plating.

APPLICATION: Low level concentrations in drinking and surface waters; domestic and industrial waters. This method determines only those cyanides amenable to chlorination.

RANGE: 0.00–0.50 Cyanide

METHOD: Cyanides react with a chlorine donor to form cyanogen chloride, which subsequently reacts with Pyridine and Barbituric Acid to form a red-blue compound in proportion to the amount of cyanide originally present. The concentration of the red-blue compound is determined spectrophotometrically.

SAMPLE HANDLING & PRESERVATION: Cyanide solutions tend to be unstable and should be analyzed as soon as possible. Samples can be stabilized by adjusting the pH to greater than 12 with NaOH. However, the pH will have to be readjusted to pH 10.5 before performing the test.

INTERFERENCES: Oxidizing agents and aldehydes can react with cyanide, while reducing agents, such as sulfite, react with the chlorine donor; both can cause negative interferences. Thiocyanate and chloride both react as cyanide in this test and will give a positive interference. Color and turbidity can also interfere.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 35 Cyanide) from TESTING MENU.
5. Scroll to and select 35 Cyanide from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Dip the end of plastic rod (0519) into water sample and touch it to a small piece (1/4 inch) of pH test paper (2955) to wet paper. Read pH immediately from color chart.

   a) If pH is below 10, raise the pH by adding *Sodium Hydroxide, 1N (4004) one drop at a time with stirring. Check pH after each drop with a new piece of pH test paper. Continue adjustment until pH is between 10.5 and 11.0.

   b) If pH is above 11.5, lower pH by adding *Hydrochloric Acid (6130) one drop at a time with stirring. Check pH after each drop with a new piece of pH test paper. Continue adjustment until pH is between 10.5 and 11.0.
8. Insert tube into chamber, close lid and select SCAN BLANK.
9. Remove tube from colorimeter. Use the 1.0 mL pipet (0354) to add 1.0 mL of Cyanide Buffer (2850PS) to tube. Cap and mix.
10. Use one 0.1 g spoon (0699) to add one level measure of *Cyanide Cl Reagent (2794DS). Cap and invert 10 times to mix. Wait 30 seconds.
11. During the 30 second waiting period, carefully fill a second 0.1 g spoon (0699) with one level measure of *Cyanide Indicator Reagent (2793DS).
12. At the end of the 30 second waiting period, immediately add the level measure of *Cyanide Indicator Reagent (2793DS). Cap and shake vigorously for 20 seconds. Wait 20 minutes for maximum color development.
13. At the end of the twenty minute waiting period, mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
14. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

CYANURIC ACID

MELAMINE METHOD–TURBIDITY • CODE 366I-SC

QUANTITY	CONTENTS	CODE
2 x 250 mL	*Acid Test Solution	*4856-K
1	Syringe, 5 mL	0807

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Cyanuric acid is added to swimming pool water as a stabilizing agent for free chlorine residuals. It minimizes the loss of chlorine from the action of ultraviolet rays in sunlight. Cyanuric acid levels in pools should be maintained between 25 and 75 ppm and various public health associations recommend that the concentration should never exceed 100-150 ppm.

APPLICATION: Swimming pool waters.

RANGE: 5–200 Cyanuric Acid

METHOD: A buffered solution of melamine forms a precipitate with cyanuric acid in proportion to the amount of cyanuric acid present. The amount of particles in suspension is measured turbidimetrically.

SAMPLE HANDLING & PRESERVATION: Cyanuric acid samples should be analyzed as soon as possible after collection. Deterioration of the sample can be minimized by keeping samples in the dark or refrigerated until analysis can be performed.

INTERFERENCES: No known interference from compounds normally found in pool water. Temperature of the sample should be maintained between 70°F and 80°F for best results. Check for stray light interference (see p. 17).

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `36 Cyanuric`) from `TESTING MENU`.
5. Scroll to and select `36 Cyanuric` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter and pour out water. Use a graduated cylinder or similar to measure 5 mL of sample water and pour into colorimeter tube.
9. Use the 5 mL syringe (0807) to add 5 mL of *Cyanuric Acid Test Solution (4856). Cap and mix thoroughly. A precipitate will form if cyanuric acid is present. Wait 1 minute.

   ☑ NOTE: This reagent bottle has a special fitting which enables the syringe to be inserted into the top of the bottle. With syringe in place, invert bottle and withdraw syringe plunger until 5 mL of reagent is contained in the syringe barrel. Remove syringe from reagent bottle and depress plunger to dispense into the tube.
10. At end of 1 minute waiting period, mix thoroughly, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For the most accurate results, the sample and reagents should be at 25 ±4°C.

CYANURIC ACID–UDV

MELAMINE METHOD-TURBIDITY–UNIT DOSE VIALS CODE 4313-H

QUANTITY	CONTENTS	CODE
1	Cyanuric Acid Unit Dose Vials, 10 pouches	4313-H

Equipment needed but not suppled:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 3 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tip (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

Cyanuric acid is added to swimming pool water as a stabilizing agent for free chlorine residuals. It minimizes the loss of chlorine from the action of ultraviolet rays in sunlight. Cyanuric acid levels should be maintained between 25 and 75 ppm and various public health associations recommend that the concentration should never exceed 100–150 ppm.

APPLICATION: Swimming pool water.

RANGE: 5–150 ppm Cyanuric Acid

METHOD: A buffered solution of melamine forms a precipitate with cyanuric acid in proportion to the amount of cyanuric acid present. The amount of particles in suspension is measured turbidimetrically.

SAMPLE HANDLING & PRESERVATION: Cyanuric acid samples should be analyzed as soon as possible after collection. Deterioration of the sample can be minimized by keeping samples in the dark or refrigerated until analysis can be performed.

INTERFERENCES: No known interference from compounds normally found in pool water. Temperature of the sample should be maintained between 70°F and 80°F for best results. Check for stray light interference (see p. 17).

PROCEDURE

Use 10 mm square cell adapter.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to selct `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `37 Cyanuric-UDV`) from `TESTING MENU`.
5. Scroll to and select `37 Cyanuric-UDV` from menu.
6. Rinse a clean vial (0156) with sample water.
7. Use the syringe (1184) to add 3 mL of sample to the vial.
8. Insert the vial into chamber, close lid and select `SCAN BLANK`.
9. Remove vial from colorimeter.
10. Use the syringe (1184) to add 3 mL of sample to a Cyanuric Acid UDV vial (4313).
11. Invert the vial 3 times to mix.
12. Wait 2 minutes.
13. Invert vial 3 more times to mix.

    ☑ NOTE: Firmly tap side of vial 5-10 times to remove all air bubbles.
14. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack.

DISSOLVED OXYGEN

WINKLER COLORIMETRIC METHOD • CODE 3688-SC

QUANTITY	CONTENTS	CODE
30 mL	*Manganese Sulfate Solution	*4167-G
30 mL	*Alkaline Potassium Iodide Azide	*7166-G
30 mL	*Sulfuric Acid 1:1	*6141WT-G
1	Sample Tube, screw cap	29180
1	Cap	28570

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Dissolved oxygen is vital to the survival of aquatic organisms. Naturally present, dissolved oxygen enters the water when plants photosynthesize. Wind and wave action also cause oxygen from the air to dissolve into water. Dissolved oxygen is consumed by aquatic animals and by the oxidation, or chemical breakdown, of dead and decaying plants and animals. The concentration of dissolved oxygen in natural waters can range from 0 to 14 ppm and is effected by temperature and salinity.

APPLICATION: This method is applicable for the determination of dissolved oxygen in drinking water, all surface waters and wastewater.

RANGE: 0.0–11.0 Dissolved Oxygen

METHOD: This method uses the azide modification of the Winkler Method with a colorimetric determination of the yellow iodine produced from the reaction with the dissolved oxygen.

INTERFERENCES: The presence of other oxidizing agents may cause positive interferences. Reducing may cause negative interferences. Nitrite interferences are eliminated with the azide modification.

COLLECTION & TREATMENT OF THE WATER SAMPLE

Steps 1 through 4 below describe proper sampling technique in shallow water. For sample collection at depths beyond arm's reach, special water sampling apparatus is required (e.g. the LaMotte Water Sampling Chamber, Code 1060; Model JT-1 Water Samplers, Code 1077; Water Sampling Outfit, Code 3103; or Water Sampling Bottle, Code 3-0026).

1. To avoid contamination, thoroughly rinse the screw cap Sample Tube (29180) with sample water.
2. Tightly cap Sample Tube and submerge to the desired depth. Remove cap and allow the Sample Tube to fill.
3. Tap the sides of the submerged tube to dislodge any air bubbles clinging to the inside. Replace the cap while the Sample Tube is still submerged.
4. Retrieve Sample Tube and examine it carefully to make sure that no air bubbles are trapped inside. Once a satisfactory sample has been collected, proceed immediately with Steps 5 and 6 to "fix" the sample.

   ☑ NOTE: Be careful not to introduce air into the sample while adding the reagents in steps 5 and 6. Simply drop the reagents into the sample. Cap carefully, and mix gently.
5. Add 2 drops of *Manganese Sulfate Solution (4167) and 2 drops of *Alkaline Potassium Iodide Azide (7166). Cap and mix by inverting several times. A precipitate will form. Allow the precipitate to settle below the shoulder of the tube before proceeding.
6. Add 8 drops of *Sulfuric Acid, 1:1 (6141WT). Cap and gently mix until the reagent and the precipitate have dissolved. A clear-yellow to brown-orange color will develop, depending on the oxygen content of the sample.

   ☑ NOTE: It is very important that all "brown flakes" are dissolved completely. If the water has a high DO level this could take several minutes. If flakes are not completely dissolved after 5 minutes, add 2 drops of *Sulfuric Acid 1:1 (6141WT) and continue mixing.

☑ NOTE: Following the completion of step 6, contact between the water sample and the atmosphere will not affect the test result. Once the sample has been "fixed" in this manner, it is not necessary to perform the actual test procedure immediately. Thus, several samples can be collected and "fixed" in the field, and then carried back to a testing station or laboratory where the test procedure is to be performed.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 39 DO) from TESTING MENU.
5. Scroll to and select 39 DO from menu.
6. Rinse a clean tube (0290) with untreated sample water. Fill to the 10 mL line with sample. This tube is the BLANK.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Fill a second tube (0290) to the 10 line with the treated "Fixed" sample. This tube is the SAMPLE.
9. Remove BLANK from colorimeter, insert SAMPLE tube into chamber, close lid and select SCAN SAMPLE. Record result.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

FLUORIDE

SPADNS METHOD • CODE 3647-01-SC

QUANTITY	CONTENTS	CODE
4 x 30 mL	*Acid Zirconyl SPADNS Reagent	*3875-G
60 mL	*Sodium Solution	*4128-H
1	Pipet, 0.5 mL, plastic	0353
1	Pipet, 1.0 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Fluoride may occur naturally in some ground waters or it may be added to public drinking water supplies to maintain a 1.0 mg/L concentration to prevent dental cavities. At higher concentrations, fluoride may produce an objectionable discoloration of tooth enamel called fluorosis, though levels up to 8 mg/L have not been found to be physiologically harmful.

☑ NOTE: This procedure uses the EPA approved Reagent System for fluoride found in method 4500-F-D, 18th Edition of *Standard Methods*, pp. 1-27.

APPLICATION: Drinking and surface waters; domestic and industrial waters.

RANGE: 0.00–2.00 Fluoride

METHOD: Colorimetric test based upon the reaction between fluoride and zirconium dye lake. The fluoride reacts with the dye lake, dissociating a portion of it into a colorless complex ion and dye. As the fluoride concentration increases, the color produced becomes progressively lighter.

SAMPLE HANDLING & PRESERVATION: Samples may be stored and refrigerated in plastic containers.

INTERFERENCES: The following substances produce a positive interference at the concentration given:

Chloride (Cl^-)	7000 mg/L
Phosphate (PO_4^{-3})	16 mg/L
$(NaPO_3)_6$	1 mg/L

The following substances produce a negative interference at the concentration given:

Alkalinity ($CaCO_3$)	5000 mg/L
Aluminum (Al^{3+})	0.1 mg/L
Iron (Fe^{3+})	10 mg/L
Sulfate (SO_4^{-2})	200 mg/L

Color and turbidity must be removed or compensated for in the procedure. Temperature should be maintained within 5°C of room temperature.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `41 Fluoride`) from `TESTING MENU`.
5. Scroll to and select `41 Fluoride` from menu.
6. This test requires a reagent blank. Rinse a clean tube (0290) with clear, colorless, fluoride free water. Fill to the 10 mL line with clear, colorless, fluoride free water.
7. Use the 0.5 mL pipet (0353) to add 0.5 mL of *Sodium Arsenite Solution (4128). Cap and mix.
8. Use the 1.0 mL pipet (0354) to add 2 measures of *Acid-Zirconyl SPADNS Reagent (3875). Cap and mix thoroughly. (This is the reagent blank.)
9. Insert tube into chamber, close lid and select `SCAN BLANK`.
10. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample water. Repeat steps 7 and 8.
11. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

HYDRAZINE

p-DIMETHYLAMINOBENZALDEHYDE METHOD
CODE 3656-SC

QUANTITY	CONTENTS	CODE
2x60 mL	*Hydrazine Reagent A	*4841-H
10 g	*Hydrazina Reagent B Powder	*4842-D
1	Pipet, 1.0 mL, plastic	0354
1	Spoon, 0.15 g, plastic	0727

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Hydrazine, N_2H_4, is added to the water in high pressure boilers to reduce corrosion by acting as an oxygen scavenger.

APPLICATION: Water and boiler water, industrial waste water.

RANGE: 0.00–1.00 Hydrazine

METHOD: p-Dimethylaminobenzaldehyde reacts with hydrazine under acidic conditions to form a yellow color in proportion to the amount of hydrazine present.

SAMPLE HANDLING & PRESERVATION: Samples should be analyzed as soon as possible after collection due to the ease with which hydrazine becomes oxidized. Acidification of the sample may increase the time between collection and analysis.

INTERFERENCES: The substances normally present in water do not interfere with the test, with the exception of strong oxidizing agents.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `45 Hydrazine`) from `TESTING MENU`.
4. Scroll to and select `45 Hydrazine` from menu.
5. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
6. Insert tube into chamber, close lid and select `SCAN BLANK`.
7. Remove tube from colorimeter. Use the 1 mL pipet (0354) to add 4 mL of *Hydrazine Reagent A (4841). Cap and mix.
8. Use the 0.15 g spoon (0727) to add one measure of *Hydrazine Reagent B Powder (4842). Cap and shake vigorously for 10 seconds. Wait 2 minutes for maximum color development. An undissolved portion of Hydrazine Reagent B may remain in bottom of tube without adversely affecting results.
9. At the end of the 2 minute waiting period, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

HYDROGEN PEROXIDE–LOW RANGE

DPD METHOD • CODE 3662-SC

QUANTITY	CONTENTS	CODE
30 mL	*Hydrogen Peroxide Reagent #1	*6452-G
100	*Hydrogen Peroxide LR Tablets	*6454A-J
1	Tablet Crusher	0175

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Hydrogen peroxide, H_2O_2, is a colorless compound that is widely used as a bleaching or decolorizing agent in the manufacture of many commercial products. As an oxidizing compound it is also used in the treatment of sewage to reduce odors and corrosion due to hydrogen sulfide. It may also be used as a sanitizing agent for water treatment. Hydrogen peroxide is relatively unstable, and for this reason it dissipates quickly and leaves no residuals.

APPLICATION: Drinking and surface waters; domestic and industrial waste water.

RANGE: 0.00–1.50 ppm Hydrogen Peroxide

METHOD: Hydrogen peroxide reacts with an excess of potassium iodide through the action of a catalyst and buffer to release an equivalent amount of iodine. The iodine in turn reacts with diethyl-p-phenylenediamine (DPD) to produce a pink-red color in proportion to the iodine released.

SAMPLE HANDLING & PRESERVATION: Hydrogen peroxide is not stable in aqueous solutions. Exposure to sunlight and agitation will accelerate the reduction of hydrogen peroxide in dilute solutions. For best results start analysis immediately after sampling.

INTERFERENCES: The likelihood of other oxidizing compounds interfering with this method is eliminated by the presence of hydrogen peroxide. Manganese may interfere and should be removed before analysis

PROCEDURE

Use universal sample holder

1. Press and hold **ON** button until spectrophotometer turns on.
2. Scroll to and select `PROGRAMMED TESTS`.
3. Scroll to and select `ALL TESTS` (or another sequence containing `46 H Peroxide-LR`) from `TESTING MENU`.
4. Scroll to and select `46 H Peroxide-LR` from menu.
5. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
6. Insert tube into chamber, close lid and select `SCAN BLANK`. (See Note)
7. Remove tube from Spectro and add 4 drops of *Hydrogen Peroxide Reagent #1 (6452). Cap and mix.
8. Add one *Hydrogen Peroxide LR Tablet (6454A). Crush tablet with tablet crusher (0175). Cap and mix for 30 seconds. Solution will turn pink if hydrogen peroxide is present Wait 5 minutes for full color development.
9. At end of 5 minute waiting period, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
10. Press **OFF** button to turn spectrophotometer off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

For the most accurate results, the sample and reagents should be at 25 ±4°C.

HYDROGEN PEROXIDE–HIGH RANGE

DPD Method • CODE 4045

QUANTITY	CONTENTS	CODE
30 mL	*Hydrogen Peroxide Reagent #1	*6452-G
100	*Hydrogen Peroxide LR Tablets	*6454A-J
1	Tablet Crusher	0175
1	Pipet, glass	0342

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Large quantities of hydrogen peroxide are added to a swimming pool to "shock" it. Shocking breaks down waste products and re-establishes a positive level of sanitizer. While many types of shock can be used with chlorine or bromine pools, only hydrogen peroxide can be used to shock biguanide pools.

Hydrogen peroxide, H_2O_2, is a colorless compound that is widely used as a bleaching or decolorizing agent in the manufacture of many commercial products. As an oxidizing compound it is also used in the treatment of sewage to reduce odors and corrosion due to hydrogen sulfide. It may also be used as a sanitizing agent for water treatment. Hydrogen peroxide is relatively unstable, and for this reason it dissipates quickly and leaves no residuals.

APPLICATION:	Drinking, industrial, domestic and swimming pool waters
RANGE:	0–60 ppm Hydrogen Peroxide
METHOD:	Hydrogen peroxide reacts with an excess of potassium iodide through the action of a catalyst and buffer to release an equivalent amount of iodine. The iodine in turn reacts with diethyl-p-phenylenediamine (DPD) to produce a pink-red color in proportion to the iodine released.
SAMPLE HANDLING & PRESERVATION:	Hydrogen peroxide is not stable in aqueous solutions. Exposure to sunlight and agitation will accelerate the reduction of hydrogen peroxide in dilute solutions. For best results start analysis immediately after sampling.
INTERFERENCES:	The likelihood of other oxidizing compounds interfering with this method is eliminated by the presence of hydrogen peroxide. Manganese may interfere and should be removed before analysis

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `Testing Menu`.
4. Select `ALL TESTS` (or another sequence containing `47 H Per-HR`) from `TESTING MENU`.
5. Scroll to and select `47 H Per-HR` from menu.
6. Use the pipet (0342) to add 5 drops of the sample water to a tube (0290).
7. Dilute to the 10 mL line with distilled or hydrogen peroxide-free water.
8. Insert the tube into chamber, close lid and select `SCAN BLANK`.
9. Remove the tube from colorimeter and add 4 drops of *Hydrogen Peroxide Reagent #1 (6452). Cap and mix.
10. Add one *Hydrogen Peroxide LR Tablet (6454A). Crush tablet with Tablet Crusher (0175). Cap and mix for 30 seconds. Solution will turn pink if hydrogen peroxide is present. Wait 5 minutes for full color development.
11. At the end of 5 minute waiting period, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
12. Press **OFF** button to turn the meter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

For the most accurate results, the sample and reagents should be at 25 ±4°C.

HYDROGEN PEROXIDE–SHOCK

DPD Method • CODE 4045

QUANTITY	CONTENTS	CODE
30 mL	*Hydrogen Peroxide Reagent #1	*6452-G
100	*Hydrogen Peroxide LR Tablets	*6454A-J
1	Tablet Crusher	0175
1	Pipet, glass	0342

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Large quantities of hydrogen peroxide shock are added to a swimming pool to "shock" it. Shocking breaks down waste products and re-establishes a positive level of sanitizer. While many types of shock can be used with chlorine or bromine pools, only hydrogen peroxide shock can be used to shock biguanide pools.

APPLICATION: Swimming pools

RANGE: 0–225 ppm Hydrogen Peroxide Shock

METHOD: Hydrogen peroxide reacts with an excess of potassium iodide through the action of a catalyst and buffer to release an equivalent amount of iodine. The iodine in turn reacts with diethyl-p-phenylenediamine (DPD) to produce a pink-red color in proportion to the iodine released.

SAMPLE HANDLING & PRESERVATION: Hydrogen peroxide is not stable in aqueous solutions. Exposure to sunlight and agitation will accelerate the reduction of hydrogen peroxide in dilute solutions. For best results start analysis immediately after sampling.

INTERFERENCES: The likelihood of other oxidizing compounds interfering with this method is eliminated by the presence of hydrogen peroxide. Manganese may interfere and should be removed before analysis

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `Testing Menu`.
4. Select `ALL TESTS` (or another sequence containing `48 H Per Shock`) from `TESTING MENU`.
5. Scroll to and select `48 H Per Shock` from menu.
6. Use the pipet (0342) to add 5 drops of the sample water to a tube (0290).
7. Dilute to the 10 mL line with distilled or hydrogen peroxide-free water.
8. Insert the tube into chamber, close lid and select `SCAN BLANK`.
9. Remove the tube from colorimeter and add 4 drops of *Hydrogen Peroxide Reagent #1 (6452). Cap and mix.
10. Add one *Hydrogen Peroxide LR Tablet (6454A). Crush tablet with Tablet Crusher (0175). Cap and mix for 30 seconds. Solution will turn pink if hydrogen peroxide is present. Wait 5 minutes for full color development.
11. At the end of 5 minute waiting period, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
12. Press **OFF** button to turn the meter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

For the most accurate results, the sample and reagents should be at 25 +ÛÛ±4°C.

IRON

1,10-PHENANTHROLINE METHOD • CODE 3668-SC

QUANTITY	CONTENTS	CODE
15 mL	*Acid Phenanthroline Indicator	*2776-E
5 g	*Iron Reducing Reagent	*2777-C
1	Spoon, 0.1 g, plastic	0699

***WARNING:** Reagents marked with a * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact LaMotte by email, phone or fax.

Most natural waters contain some iron. Its presence may vary from small traces to very large amounts in water which is contaminated by acid mine wastes. For domestic use, the concentration should not exceed 0.2 ppm and for some industrial applications not even a trace of iron can be tolerated. There are many means available for removing or reducing the iron content. Water softening resins are effective for removing small amounts of iron and special ion exchange materials are selective for iron removal. High concentrations of iron can be removed by such chemical processes as oxidation and lime or lime-soda softening. Because of the many means of removing or reducing the amount of iron in water, the particular method employed will depend largely on the form of iron which is present and the end use of the treated water.

APPLICATION: Drinking, surface and saline waters; domestic and industrial wastes.

RANGE: 0.00–5.00 Iron

METHOD: Ferric iron is reduced to ferrous iron and subsequently forms a colored complex with phenanthroline for a quantitative measure of total iron.

SAMPLE HANDLING & PRESERVATION The sample container should be cleaned with acid and rinsed with deionized water. Addition of acid to adjust the sample to pH 2–3 will prevent deposition of iron on the container walls. Samples should be analyzed as soon as possible after collection since ferrous iron undergoes oxidation to ferric iron.

INTERFERENCES: Strong oxidizing agents, cyanide, nitrite, and phosphates, chromium, zinc in concentrations exceeding 10 times that of iron; cobalt and copper in excess of 5 mg/L, and nickel in excess of 2 mg/L. Bismuth, cadmium, mercury, , and silver precipitate phenanthroline.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `53 Iron Phen`) from `TESTING MENU`.
5. Scroll to and select `53 Iron Phen` from menu.
6. Rinse a clean tube (0290)with sample water. Fill to the 10 mL mark with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter. Remove the cap and add 6 drops of *Acid Phenanthroline Indicator (2776). Cap and invert the tube 4 times to mix reagents. Wait five minutes for maximum color development.
9. After five minutes, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result as ppm Ferrous Iron.
10. Remove the tube from colorimeter. Use the 0.1g spoon (0699) to add one measure of *Iron Reducing Reagent (2777). Cap and invert 15-20 times times to mix. Wait 5 minutes for maximum color delelopment.
11. After 5 minutes, mix, insert tube into chamber, close lid and select `SCAN SAMPLE.` Record result as ppm Total Iron.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.
13. Total Iron (ppm) - Ferrous Iron (ppm) = Ferric Iron (ppm)

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

IRON

BIPYRIDYL METHOD • CODE 3648-SC

QUANTITY	CONTENTS	CODE
30 mL	*Iron Reagent #1	*4450-G
5 g	*Iron Reagent #2 Powder	*V-4451-C
1	Pipet, 0.5 mL, plastic	0353
1	Spoon, 0.1 g, plastic	0699

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Most natural waters contain some iron. Its presence may vary from small traces to very large amounts in water which is contaminated by acid mine wastes. For domestic use, the concentration should not exceed 0.2 ppm and for some industrial applications not even a trace of iron can be tolerated. There are many means available for removing or reducing the iron content. Water softening resins are effective for removing small amounts of iron and special ion exchange materials are selective for iron removal. High concentrations of iron can be removed by such chemical processes as oxidation and lime or lime-soda softening. Because of the many means of removing or reducing the amount of iron in water, the particular method employed will depend largely on the form of iron which is present and the end use of the treated water.

APPLICATION: Drinking, surface and saline waters; domestic and industrial wastes.

RANGE: 0.00–6.00 Iron

METHOD: Ferric iron is reduced to ferrous iron and subsequently forms a colored complex with bipyridyl for a quantitative measure of total iron.

SAMPLE HANDLING & PRESERVATION: The sample container should be cleaned with acid and rinsed with deionized water. Addition of acid to adjust the sample to pH 2–3 will prevent deposition of iron on the container walls. Samples should be analyzed as soon as possible.

INTERFERENCES: Strong oxidizing agents interfere, as well as copper and cobalt in excess of 5.0 mg/L.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 51 Iron Bipyr) from TESTING MENU.
5. Scroll to and select 51 Iron Bipyr from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Use the 0.5 mL pipet (0353) to add one measure of *Iron Reagent #1 (V-4450). Cap and mix.
9. Use the 0.1 g spoon (0699) to add 0.1 g of *Iron Reagent #2 Powder (V-4451). Cap and shake vigorously for 30 seconds. Wait three minutes for maximum color development.
10. At the end of 3 minute waiting period, do not mix. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

IRON–UDV

BIPYRIDYL METHOD–UNIT DOSE VIALS • CODE 4315-H

QUANTITY	CONTENTS	CODE
1	*Iron Unit Dose Vials, 10 pouches	*4315-H

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Equipment needed but not supplied:

STANDARD ACCESSORY PACKAGE • CODE 1961

1	Package of 3 Vials (empty)	0156
1	Syringe, 6 mL, plastic	1184
1	Foil Storage Bag	9467

Or:

ADVANCED ACCESSORY PACKAGE • CODE 1962

1	Pipettor	30528
1	Pipet Tip (0-5 mL)	30695
1	Cuvette Rack	31695
1	Package of 3 Vials (empty)	0156
1	Foil Storage Bag	9467

Most natural waters contain some iron. Its presence may vary from small traces to very large amounts in water which is contaminated by acid mine wastes. For domestic use, the concentration should not exceed 0.2 ppm and for some industrial applications not even a trace of iron can be tolerated. There are many means available for removing or reducing iron content. Water softening resins are effective for removing small amounts of iron and special ion exchange materials are selective for iron removal. High concentrations of iron can be removed by such chemical processes as oxidation and lime or lime-soda softening. Because of the many means of removing or reducing the amount of iron in water, the particular method employed will depend largely on the form of iron which is present and the end use of the treated water.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial wastes.

RANGE: 0.00–10.00 ppm

METHOD: Ferric iron is reduced to ferrous iron and subsequently forms a colored complex with bipyridyl for a quantitative measure of total iron.

SAMPLE HANDLING & PRESERVATION: The sample container should be cleaned with acid and rinsed with deionized water. Addition of acid to adjust the sample th pH 2-3 will prevent deposition of iron on the container walls. Samples should be analyzed as soon as possible.

INTERFERENCES: Strong oxidizing agents interfere, as well as copper and cobalt in excess of 5.0 ppm.

PROCEDURE

Use 10 mm square cell adapter.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `52 Iron-UDV`) from `TESTING MENU`.
5. Scroll to and select `52 Iron-UDV` from menu.
6. Rinse a clean vial (0156) with sample water.
7. Use the syringe (1184) to add 3 mL of sample to the vial.
8. Insert the vial into the chamber, close the lid and select `SCAN BLANK`.
9. Remove the vial from the colorimeter.
10. Use the syringe (1184) to add 3 mL of sample to an *Iron UDV vial (4315).
11. Wait 2 minutes.
12. Invert vial 3 times to mix.
    ☑ NOTE: If powder residue remains in the bottom of the vial after inverting, or air bubbles form, invert vial once more and tap bottom of vial sharply once or twice to dislodge powder or bubbles. Mix.
13. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
14. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

UDVs from opened pouches should be used promptly. Store unused vials from opened pouches in the Foil Storage Bag (9467) to extend the shelf life of the reagent. Generally, UDVs stored in the bag should be used within 10 days if the humidity is less than 50% and within 5 days if humidity is greater than 50%. The Foil Storage Bag contains a desiccant pack with indicator. When the indicator in the window turns from blue to pink, the bag should be replaced.

LEAD

PAR METHOD • CODE 4031

QUANTITY	CONTENTS	CODE
250 mL	Ammonium Chloride Buffer	4032-K
15 mL	*Sodium Cyanide, 10%	*6565-E
30 mL	PAR Indicator	4033-G
30 mL	Stabilizing Reagent	4022-G
15 mL	DDC Reagent	4034-E
1	Syringe, 5 mL, plastic	0807
2	Pipet, 0.5 mL, plastic	0353

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The average concentration of lead is 0.003 ppm in streams and less than 0.1 ppm in groundwater. Lead in a water supply may come from mine and smelter discharges or from industrial waste. Lead is used in the production of batteries, solder, pigments, insecticides, ammunition and alloys. Tetraethyl Lead has been used for years as an anti-knock reagent in gasoline. Lead may also enter water supplies when corrosive water dissolves pipes, plumbing fixtures and materials containing lead. Lead accumulates in the body and is toxic by ingestion.

APPLICATION: Drinking and surface waters; domestic and industrial wastewater.

RANGE: 0.00–5.00 Lead

METHOD: Lead and calcium ions form a red complex with PAR (4- [2'-pyridylazo] resorcinol), at a pH of about 10. When sodium diethyldithiocarbamate is added, the lead/PAR complex is destroyed leaving the calcium/PAR complex. The difference between the two measurements is due to the lead concentration.

SAMPLE HANDLING & PRESERVATION: Analyze sample as soon as possible. If sample must be stored, acidify with nitric acid to a pH of below 2.

INTERFERENCES: Calcium greater than 100 ppm (250 ppm $CaCO_3$) will interfere. Low concentrations of cerium, iron, manganese, magnesium, sulfur, tin, and EDTA will also interfere.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 54 Lead) from TESTING MENU.
5. Scroll to and select 54 Lead from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select SCAN BLANK.
8. Remove the tube from colorimeter. Use the Syringe (0807) to remove 5mL of sample from tube. Discard remaining sample.
9. Add the 5 mL of sample in the syringe to the tube. Add 5 mL Ammonium Chloride Buffer (4032) to fill the tube to the 10 mL line. Swirl to mix.
10. Add 3 drops *Sodium Cyanide, 10% (6565). Swirl to mix.
11. Use the 0.5 mL pipet (0353) to add 0.5 mL PAR Indicator (4033). Swirl to mix.
12. Use the 0.5 mL pipet (0353) to add 0.5 mL Stabilizing Reagent (4022). Cap and mix.
13. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result in ppm as Reading A.
14. Remove tube from colorimeter. Add 3 drops DDC Reagent (4034). Cap and mix.
15. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result in ppm as Reading B.
16. Calculate result:

Lead (ppm) = Reading A – Reading B

17. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

MANGANESE–LOW RANGE

PAN METHOD • CODE 3658-01-SC

QUANTITY	CONTENTS	CODE
4x30 mL	*Hardness Buffer Reagent	*4255-G
30 mL	*Manganese Indicator Reagent	*3956-G
15 mL	*Sodium Cyanide, 10%	*6565-E
1	Pipet, 0.5 mL, plastic	0369
1	Pipet, 1.0 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Manganese is present in ground water in the divalent state due to the lack of oxygen. In surface waters manganese may be in various oxidation states as soluble complexes or as suspended compounds. Manganese is rarely present in excess of 1 mg/L. It may cause an objectionable taste or cause staining problems in laundry, but manganese levels normally encountered in water seldom produce any health hazard.

Manganese is removed from water by various means including chemical precipitation, pH adjustment, aeration, superchlorination and the use of ion exchange resins.

APPLICATION: Drinking and surface waters; domestic and industrial wastewaters.

RANGE: 0.00–0.70 ppm Manganese

METHOD: PAN (1-[2-Pyridylazo]-2-Naphthol) forms a red complex with Manganese (Mn^{2+}) at a pH of 10 to 11.

SAMPLE HANDLING & PRESERVATION: Manganese may oxidize readily in neutral water and precipitate from solution. It may adhere to or be absorbed by container walls, especially glass. Acidified samples can be stored in plastic.

INTERFERENCES: None. Test is quite specific.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 55 Manganese L) from TESTING MENU.
5. Scroll to and select 55 Manganese L from menu.
6. Rinse a tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Use the 1.0 mL pipet (0354) to add 2.0 mL (two measures) of *Hardness Buffer Reagent (4255). Swirl to mix.
9. Add 2 drops of *Sodium Cyanide, 10% (6565). Cap and mix.
10. Use the 0.5 mL pipet (0369) to add 0.5 mL of *Manganese Indicator Reagent (3956). Cap and mix.
11. Immediately insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

MANGANESE–HIGH RANGE

PERIODATE METHOD • CODE 3669-SC

QUANTITY	CONTENTS	CODE
10 g	Manganese Buffer Reagent	6310-D
15 g	*Manganese Reagent	*6311-E
1	Spoon, 0.1 g, plastic	0699
1	Spoon, 0.15 g, plastic	0727

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Manganese is present in ground water in the divalent state due to the lack of oxygen. In surface waters, manganese may be in various oxidation states as soluble complexes or as suspended compounds. Manganese is rarely present in excess of 1 mg/L. It may impart an objectionable taste or cause staining problems in laundry, but manganese levels normally encountered in water seldom produce any health hazards. Manganese is removed from water by various means, including chemical precipitation, pH adjustment, aeration, superchlorination and the use of ion exchange resins.

APPLICATION: Drinking and surface waters, domestic and industrial wastewaters.

RANGE: 0.0–15.0 Manganese

METHOD: Periodate oxidizes soluble manganous compounds into permanganate.

SAMPLE HANDLING & PRESERVATION: Manganese may oxidize readily in a neutral water and precipitate from solution. It may adhere to or be absorbed by container walls, especially glass. Acidified samples can be stored in plastic.

INTERFERENCES: Reducing substances capable of reacting with periodate or permanganate must be removed or destroyed before the periodate oxidation is attempted.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 56 Manganese H) from TESTING MENU.
5. Scroll to and select 56 Manganese H from menu.
6. Rinse a tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Use the 0.1 g spoon (0699) to add two measures of Manganese Buffer Reagent (6310). Cap and mix until powder dissolves.
9. Use the 0.15 g spoon (0727) to add one measure of *Manganese Periodate Reagent (6311). Cap and shake for one minute. An undissolved portion of the reagent may remain in the bottom of the tube without adversely affecting the test results. Wait two minutes for maximum color development. Solution will turn pink if manganese is present.
10. At the end of the two minute waiting period, mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

MERCURY

TMK METHOD • CODE 4861

QUANTITY	CONTENTS	CODE
50	*TMK Tablets	*4862-H
2 x 250 mL	*Propyl Alcohol	*4863-K
250 mL	*Acetate Buffer	*4864-K
1	Tablet Crusher	0175
1	Test Tube, 10 , glass, w/cap	0778
1	Pipet, 1.0 mL, plastic	0354
1	0.5 mL, plastic	0353

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Mercury occurs in small amounts in soil, streams and groundwater. It is used in the production of amalgams, mirror coatings and measuring devices such as thermometers, barometers and manometers. Pharmaceuticals and paints contain mercury. It is also used in fungicides and pesticides and as a mold retardant on paper. Some forms of mercury are very toxic and can accumulate in the aquatic food chain.

APPLICATION: Drinking and surface waters; domestic and industrial wastewater.

RANGE: 0.00–1.50 Mercury

METHOD: Mercuric ions (Hg^{+2}) form a colored complex with 4, 4'-bis (dimethylamino) thiobenzophenone (Thio-Michler's ketone, TMK) at pH 3.

SAMPLE HANDLING & PRESERVATION: Analyze sample as soon as possible. If sample must be stored, treat with HNO_3 to reduce th pH to less than 2 and store in a glass container.

INTERFERENCES: Palladium and other noble metals (gold, platinum, rhodium, iridium, ruthenium), iodide and reducing agents such as hydroxylamine hydrochloride, ascorbic acid, sulfite and thiosulfate. Interference due to silver is eliminated if chloride is present.

PREPARATION OF *TMK INDICATOR

☑ NOTE: Prepare *TMK Indicator daily. Keep out of direct sunlight.

1. Fill test tube (0778) to the 10 mL line with *Propyl Alcohol (4863).
2. Add one *TMK Tablet (4862).
3. Use tablet crusher (0175) to completely crush tablet.
4. Cap and mix. Shake vigorously for 30 seconds.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 57 Mercury) from TESTING MENU.
5. Scroll to and select 57 Mercury from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select SCAN BLANK.
8. Remove the tube from colorimeter.
9. Use the 1.0 mL pipet (0354) to add 3 mL of *Acetate Buffer (4864). Cap and mix.
10. Use the 0.5 mL pipet (0353) to add 0.5 mL of prepared *TMK Indicator. Cap and mix.
11. Wait one minute.
12. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result as ppm Mercury.
13. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure using distilled or deionized water. This test result is the reagent blank. Subtract the reagent blank results from all subsequent test results of unknown samples. It is recommended that a reagent blank be determined each time *TMK Indicator is prepared.

MOLYBDENUM–HIGH RANGE

THIOGLYCOLATE METHOD • CODE 3699-02-SC

QUANTITY	CONTENTS	CODE
2 x 30 mL	*Mo Buffer	*3997-G
2 x 30 mL	*Molybdenum Oxidizing Reagent	*6485-G
2.5g	*Molybdenum Indicator Powder	*6486-S
1	Spoon, 0.05g, plastic	0696
2	Pipets, 1.0 mL, plastic w/cap	0372

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Molybdenum occurs naturally in the earth's crust as molybdenite and wolfenite, and is an important element in many biochemical reactions, including nitrogen fixation. In industrial processes, such as the operation of boilers and cooling towers, molybdenum, in the form of sodium molybdate, is used as a corrosion inhibitor.

APPLICATIONS: Boiler and cooling water.

RANGE: 0.0–50.0 ppm Molybdenum

METHOD: Calcium thioglycolate reacts with molybdenum to give a yellow color with an intensity proportional to the amount of molybdenum present.

SAMPLE HANDLING & PRESERVATION: Molybdenum samples may be stored in either plastic or glass containers.

INTERFERENCES: Nickel levels less than 50 ppm do not interfere; aluminum levels less than 10 ppm do not interfere; chromate at higher concentrations interferes due to the intense yellow color. Ferrous iron levels below 50 ppm do not interfere, but low levels of ferric iron will cause a large blank. Highly buffered samples may exceed the capacity of the system possibly producing inaccurate results. Samples with high levels of nitrite will eventually develop a pale orange color. Scan the sample immediately to avoid this inteference.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 61 Moly-HR) from TESTING MENU.
5. Scroll to and select 61 Moly-HR from menu.
6. Fill clean tube (0290) to 10 mL line with sample water.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Use a 1.0 mL pipet (0372) to add 1.0 mL of *Mo Buffer (3997). Cap and mix.
9. Use a second 1.0 mL pipet (0372) to add 1.0 mL of *Molybdenum Oxidizing Reagent (6485). Cap and mix.
10. Use 0.05 g spoon (0696) to add one measure of Molybdenum Indicator Powder (6486). Cap and mix until powder dissolves. Solution will turn yellow if molybdenum is present.
11. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

NICKEL

DIMETHYLGLYOXIME METHOD • CODE 3663-SC

QUANTITY	CONTENTS	CODE
60 mL	*Hydrochloric Acid, 2.5N	*6251PS-H
30 g	*Ammonium Persulfate Reagent	*6566-G
30 mL	*Silver Nitrate Solution, 0.0141N	*6346WT-G
250 mL	Sodium Citrate, 10%	6253-K
60 mL	*Dimethylglyoxime, 1%	*6254-H
60 mL	*Ammonium Hydroxide, Conc.	*6537-H
3	Pipets, 1.0 mL, plastic	0354
1	Spoon, 0.1 g, plastic	0699
1	Test tube, 5-10-12.9-15-20-25, glass, w/cap	0608
1	Graduated Cylinder, 10 mL, glass	0416

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Nickel is not usually found in natural waters except as a result of contamination from industrial wastewaters as a corrosion product of stainless steel and nickel alloys. Nickel may also enter surface waters from plating bath process water.

APPLICATION: Drinking and surface waters; domestic and industrial wastewater.

RANGE: 0.00–8.00 ppm Nickel

METHOD: Nickel under basic conditions forms a colored complex with dimethylglyoxime in proportion to the concentration of nickel.

SAMPLE HANDLING & PRESERVATION: Samples may be collected in either plastic or glass containers and preserved by adding 5 mL of concentrated nitric acid per liter.

INTERFERENCES: Organic matter interferes. Cobalt, iron, copper, manganese and chromium do not interfere if each of the concentrations is below 15 ppm.

PROCEDURE

1. Use the 10 mL graduated cylinder (0416) to measure 10 mL of sample water. Pour into glass test tube (0608).
2. Use the 1 mL pipet (0354) to add 1 mL of *Hydrochloric Acid, 2.5N (6251).
3. Use the 0.1 g spoon (0699) to add 2 measures of *Ammonium Persulfate Reagent (6566). Add two drops of *Silver Nitrate Solution, 0.0141N (6346WT). Mix until the powder has dissolved. The solution will be slightly cloudy at this point.
4. Use 10 mL graduated cylinder (0416) to add 5 mL of Sodium Citrate, 10% (6253).
5. Use a second 1 mL pipet (0354) to add 1 mL of *Ammonium Hydroxide, Conc. (6537). Mix, then dilute to 25 mL with deionized water.
6. Use a third 1 mL pipet (0354) to add 1 mL of *Dimethylglyoxime, 1% (6254). Mix. Wait 20 minutes for color development.
7. At end of 20 minute waiting period fill a clean tube (0290) to the 10 mL line with the developed test sample.
8. Fill a second clean tube (0290) to 10 mL line with deionized water or untreated sample water. This is the blank.
9. Press and hold **ON** button until colorimeter turns on.
10. Press **ENTER** to start.
11. Press **ENTER** to select `TESTING MENU`.
12. Select `ALL TESTS` (or another sequence containing `63 Nickel`) from `TESTING MENU`.
13. Scroll to and select `63 Nickel` from menu.
14. Insert the blank into chamber, close lid and select `SCAN BLANK`.
15. Insert test sample into chamber, close lid and select `SCAN SAMPLE`. Record result.
16. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

NITRATE

ZINC REDUCTION • CODE 3689-SC

QUANTITY	CONTENTS	CODE
50	*Nitrate Spectrophotometric Grade Tablets	*3881A-H
1	Tablet Crusher	0175

***WARNING:** Reagents marked with a * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact LaMotte by email, phone or fax.

Nitrogen is essential for plant growth, but excessive amounts in water supplies can result in nutrient pollution. Nitrates, in conjunction with phosphate, stimulate the growth of algae creating water quality problems. Nitrogen compounds may enter water as nitrates or be converted to nitrates from agricultural fertilizers, sewage, industrial and packing house wastes, drainage from livestock feeding areas and manure. Nitrates in large amounts in drinking water can cause "blue baby syndrome" (methemoglobenemia) in infants in less than 6 months of age and other health problems. US Public Health Service Drinking Water Standards state that 44 ppm nitrate should not be exceeded. To the sanitary and industrial engineer, concentrations of less than 4 ppm are acceptable.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial waters.

RANGE: 0.0–60.0 ppm Nitrate

METHOD: Zinc is used to reduce nitrate to nitrite. The nitrite that was originally present, plus the reduced nitrate, reacts with chromotropic acid to form a red color in proportion to the amount of nitrite in the sample.

SAMPLE HANDLING & PRESERVATION: Analysis should be made as soon as possible. If analysis cannot be made within 24 hours, the sample should be refrigerated at 4°C. When samples must be stored for more than 24 hours, add 2 mL of concentrated sulfuric acid per liter of sample. For best results, the analysis for nitrate should be determined at temperatures between 20°C and 25°C.

INTERFERENCES: Nitrite interferes at all concentrations. Strong oxidizing and reducing substances interfere. Low results might be obtained for samples that contain high concentrations of copper and iron.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `66 Nitrate-TT`) from `TESTING MENU`.
5. Scroll to and select `66 Nitrate-TT` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL line with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add one *Nitrate Spectrophotometric Grade Tablet (3881A).
10. Use Tablet Crusher (0175) to crush tablet.
11. Cap tube.
12. Invert tube 60 times per minute for 2 minutes (one inversion equals 180°).
13. Wait 5 minutes. Do not mix.
14. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm nitrate.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples.

To convert nitrate (NO_3) results to nitrate-nitrogen (NO_3-N), divide by 4.4.

NITRATE-NITROGEN–LOW RANGE

CADMIUM REDUCTION METHOD • CODE 3649-SC

QUANTITY	CONTENTS	CODE
2 x 60 mL	*Mixed Acid Reagent	*V-6278-H
5 g	*Nitrate Reducing Reagent	*V-6279-C
1	Spoon, 0.1 g, plastic	0699
1	Dispenser Cap	0692

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Nitrogen is essential for plant growth, but the presence of excessive amounts in water supplies presents a major pollution problem. Nitrogen compounds may enter water as nitrates or be converted to nitrates from agricultural fertilizers, sewage, industrial and packing house wastes, drainage from livestock feeding areas, farm manures and legumes. Nitrates in large amounts can cause "blue babies" (methemoglobinemia) in infants less than six months of age. Nitrate concentration is an important factor to be considered in livestock products, where, in addition to causing methemoglobinemia, it is responsible for many other problems. Nitrates in conjunction with phosphate stimulate the growth of algae with all of the related difficulties associated with excessive algae growth.

U.S. Public Health Service Drinking Water Standards state that 10 ppm nitrate nitrogen should not be exceeded. To the sanitary and industrial engineer, concentrations of less than 1 ppm are acceptable.

APPLICATION: This method determines nitrate levels in drinking, surface, saline waters, domestic and industrial waters.

RANGE: 0.00–3.00 ppm Nitrate Nitrogen

METHOD: Powdered cadmium is used to reduce nitrate to nitrite. The nitrite that is originally present plus reduced nitrate is determined by diazotization of sulfanilamide and nitrite followed by coupling with N-(1 naphthyl)-ethylenediamine dihydrochloride to form a highly colored azo dye which is measured colorimetrically.

SAMPLE HANDLING & PRESERVATION: Analysis should be made as soon as possible. If analysis cannot be made within 24 hours, the sample should be preserved by refrigeration at 4°C. When samples must be stored for more than 24 hours, they can be preserved by adding 2 mL of concentrated sulfuric acid per liter of sample. For best results, the analysis for nitrate should be determined at temperatures between 20°C and 25°C.

INTERFERENCES: Nitrite interferes at all levels. Strong oxidizing and reducing substances interfere. Low results might be obtained for samples that contain high concentrations of iron and copper.

PROCEDURE

☑ NOTE: Place Dispenser Cap (0692) on *Mixed Acid Reagent (V-6278). Save this cap for refill reagents.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 64 Nitrate-N LR) from TESTING MENU.
5. Scroll to and select 64 Nitrate-N LR from menu.
6. Rinse a clean tube (0290) with sample water. Fill to 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter and pour off 5 mL into graduated cylinder or similar. Discard the remaining sample.
9. Pour the 5mL sample from a graduated cylinder or similar into the tube. Use the graduated cylinder or similar to measure 5 mL of *Mixed Acid Reagent (V-6278) and add to tube. Cap and mix. Wait 2 minutes before proceeding to Step 10.
10. Use the 0.1 g spoon (0699) to add two measures of *Nitrate Reducing Reagent (V-6279). Cap.
11. Hold tube by index finger and thumb and mix by inverting approximately 50-60 times a minute for four minutes. Wait 10 minutes for maximum color development.

    ☑ NOTE: At end of waiting period an undissolved portion of Nitrate Reducing Reagent may remain in bottom of the tube without affecting results.
12. At the end of the 10 minute waiting period, mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
13. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

To convert Nitrate Nitrogen (NO_3–N) results to ppm Nitrate (NO_3), multiply by 4.4.

NITRITE

DIAZOTIZATION • CODE 3694-SC

QUANTITY	CONTENTS	CODE
50	*Nitrite Spectrophotometric Grade Tablets	*3886A-H
1	Tablet Crusher	0175

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Nitrite represents an intermediate stage of the nitrogen cycle, usually resulting from the bacterial decomposition of compounds containing organic nitrogen. Under aerobic conditions bacteria oxidize ammonia to nitrites; and under anaerobic conditions, bacteria reduce nitrates to nitrites. Nitrites are often used as food preservatives. The nitrite concentration of drinking water rarely exceeds 0.1 ppm.

APPLICATION: Drinking, surface, and saline waters; domestic and industrial waters.

RANGE: 0.00–1.25 ppm Nitrite

METHOD: The compound formed by diazotization of sulfanilamide and nitrite is coupled with N-(1-naphthyl)-ethylenediamine to produce a reddish purple color in proportion to the nitrite concentration.

SAMPLE HANDLING & PRESERVATION: Samples should be analyzed as soon as possible. They may be stored for 24 to 48 hours at 4°C.

INTERFERENCES: There are few known interfering substances at concentrations at less than 1000 times the nitrite-nitrogen concentration; however, the presence of strong oxidizing agents or reductants may readily affect nitrite concentrations.

PROCEDURE

1. Press and hold ON button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `69 Nitrite-TT`) from `TESTING MENU`.
5. Scroll to and select `69 Nitrite-TT` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL line with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add one *Nitrite Spectrophotometric Grade Tablet (3886).
10. Use Tablet Crusher (0175) to crush tablet.
11. Cap tube.
12. Shake vigorously for 20 seconds.
13. Wait 2 minutes.
14. Immediately, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm nitrite.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples.

To convert nitrite (NO_2) results to nitrite-nitrogen (NO_2–N), divide results by 3.3.

NITRITE-NITROGEN–LOW RANGE

DIAZOTIZATION METHOD • CODE 3650-SC

QUANTITY	CONTENTS	CODE
2 x 60 mL	*Mixed Acid Reagent	*V-6278-H
5 g	*Color Developing Reagent	*V-6281-C
1	Spoon, 0.1 g, plastic	0699
1	Dispenser Cap	0692

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Nitrite represents an intermediate state in the nitrogen cycle, usually resulting from the bacterial decomposition of compounds containing organic nitrogen. Under aerobic conditions bacteria oxidize ammonia to nitrites; and under anaerobic conditions, bacteria reduce nitrates to nitrites. Nitrites are often used as preservatives when added to certain foods.

The nitrite concentration of drinking water rarely exceeds 0.1 ppm (mg/L).

APPLICATION: This method is applicable for the determination of nitrite in drinking, surface and saline waters; domestic and industrial wastes.

RANGE: 0.00–0.80 ppm Nitrite-Nitrogen

METHOD: The compound formed by diazotization of sulfanilamide and nitrite is coupled with N–(1–naphthyl)–ethylenediamine to produce a reddish-purple color, which is read colorimetrically.

SAMPLE HANDLING & PRESERVATION: Samples should be analyzed as soon as possible. They may be stored for 24 to 48 hours at 4°C.

INTERFERENCES: There are few known interfering substances at concentration less than 1000 times the nitrite-nitrogen concentration; however, the presence of strong oxidants or reductants may readily affect nitrite concentrations. High alkalinity (above 600 mg/L) will give low results due to a shift in pH.

PROCEDURE

☑ NOTE: Place Dispenser Cap (0692) on *Mixed Acid Reagent (V-6278). Save this cap for refill reagents.

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `67 Nitrite-N LR`) from `TESTING MENU`.
5. Scroll to and select `67 Nitrite-N LR` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter and pour off 5 mL into a graduated cylinder or similar. Discard the remaining sample.
9. Pour the 5 mL sample from the graduated cylinder into the colorimeter tube. Use graduated cylinder or similar to measure 5 mL of *Mixed Acid Reagent (V-6278) and add to tube. Cap and mix.
10. Use the 0.1 g spoon (0699) to add two measures of *Color Developing Reagent (V-6281). Cap and mix by gently inverting for 1 minute. Wait 5 minutes for maximum color development.
11. At the end of the 5 minute waiting period, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: To convert nitrite-nitrogen (NO_2–N) results to ppm nitrite (NO_2), multiply results by 3.3.

NITROGEN, TOTAL

CHROMOTROPIC ACID WITH PERSULFATE DIGESTION METHOD • CODE 4026

QUANTITY	CONTENTS	CODE
25	Nitrogen Hydroxide Reagent Tubes	4040-G
5 g	*Digestion Reagent Powder	4036-C
60 mL	Deionized Water	*5115PS-H
5 g	*Total Nitrogen Reagent A Powder	*4041-C
30	*Total Nitrogen Reagent B Tablets	*4042
25	*Total Nitrogen Acid Reagent Tubes	*4043-G
2	Spoon, 0.15 g, plastic	0727
4	Pipets, 1.0 mL, plastic	0354
2	Funnels, plastic	0459

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Note: for greater accuracy, use laboratory grade pipets.

Equipment needed but not supplied:

1	COD Adapter	5-0087
1	COD Reactor, 12 tubes, 110V	5-0102
or 1	COD Reactor, 12 tubes, 230V	5-0102-EX

Optional Equipment:

4	Pipet, Measuring, 1.0 mL	2-2110
1	Pipet Bulb	2-2164
1	Wipes	2-2069
1	Test Tube Rack	23371
1	Timer	9371-W13
1	Test Tube Holder	2-2190

Nitrogen is essential for plant growth, but the presence of excessive amounts in water supplies presents a major pollution problem. Nitrogen compounds may enter water as nitrates or be converted to nitrates from agricultural fertilizers, sewage, industrial and packing house wastes, drainage from livestock feeding areas, farm manures and legumes. Nitrates in large amounts can cause "blue babies" (methemoglobinemia) in infants less than six months of age. Nitrate concentration is an important factor to be considered in livestock products, where, in addition to causing methemoglobinemia, it is responsible for many other problems. Nitrates in conjunction with phosphate stimulate the growth of algae with all of the related difficulties associated with excessive algae growth.

U.S. Public Health Service Drinking Water Standards state that 10 ppm nitrate nitrogen should not be exceeded. To the sanitary and industrial engineer, concentrations of less than 1 ppm are acceptable.

APPLICATION:	Drinking, surface, saline, domestic and industrial waters.
RANGE:	0–25 mg/L Total Nitrogen
METHOD:	All forms of nitrogen are converted to nitrate by an alkaline persulfate digestion. Interference from halogen oxides is eliminated by the addition of sodium metabisulfite. Nitrate reacts in acid with chromotropic acid to form a yellow color in proportion to the amount of nitrate in the treated sample.
SAMPLE HANDLING & PRESERVATION:	If the sample can not be analyzed immediately, the sample should be preserved by adjusting the pH to 2 or less with concentrated sulfuric acid and refrigerated at 4°C. Warm the sample to room temperature and neutralize with 5.0 N sodium hydroxide before analyzing.
INTERFERENCES:	Bromide (>60 ppm) and chloride (>1000 ppm) will have a positive interference.

PROCEDURE

Use COD adapter.

1. Preheat COD reactor to 100 ±2°C. Follow safety precautions.
2. Remove caps from two *Total Nitrogen Hydroxide Reagent Tubes (4040).
3. Use a 0.15 g spoon (0727) and a funnel (0459) to add one level measure of *Digestion Reagent Powder (4036) to each tube. Tap funnel to dispense powder completely.
4. Use a 1.0 mL pipet (0354) to add 2.0 mL of Deionized Water (5115PS), or organic-free water, to one tube. This is the blank.
5. Use another 1.0 mL pipet (0354) to add 2.0 mL of sample to the other tube. This is the sample.
6. Cap both tubes and shake vigorously for 30 seconds.
7. Place the tubes in the COD reactor for 30 minutes.
8. After exactly 30 minutes, turn the reactor off. Carefully remove the tubes from the reactor and allow them to cool to room temperature.
9. At the end of the cooling period, press and hold **ON** button until colorimeter turns on.
10. Press **ENTER** to start.
11. Press **ENTER** to select `TESTING MENU`.
12. Select `ALL TESTS` (or another sequence containing `62 Nitrogen T`) from `TESTING MENU`.
13. Scroll to and select `62 Nitrogen T` from the menu.
14. Carefully remove caps from the digested tubes.
15. Use a 0.15 g spoon (0727) and a funnel (0459) to add one level measure of *Total Nitrogen Reagent A Powder (4041). Tap funnel to dispense powder completely. Cap the tubes and shake for 15 seconds.
16. Wait 3 minutes.
17. Remove the caps from the tubes. Add one *Total Nitrogen Reagent B Tablet (4042) to each tube. Cap the tubes and shake for 45 seconds or until the tablet disintegrates.
18. Wait 2 minutes.
19. Remove the caps from the reacted tubes. Carefully remove the caps from two *Total Nitrogen Acid Reagent Tubes (4043).
    CAUTION: Tubes contain a strong acid.
20. Use another 1.0 mL pipet (0354) to add 2 mL of digested, treated blank to one Total Nitrogen Acid Reagent Tube. This is the blank.
21. Use another 1.0 mL pipet (0354) to add 2 mL of digested, treated sample to the other Total Nitrogen Acid Reagent Tube. This is the sample.
22. Cap the tubes and invert 10 times to mix.
    CAUTION: The tubes will be hot.

☑ Note: Invert slowly and completely for accurate results. Hold tubes with caps up. Invert the tube and wait for the air bubble to flow to the bottom of the tube. Turn the tube upright and wait for the air bubble to return to the top of the tube. This is one inversion.

23. Wait 5 minutes.
24. Wipe the tubes with a damp towel to remove fingerprints and smudges. Wipe with a dry towel.
25. Insert the blank tube into the chamber. Select SCAN BLANK. Remove the blank tube from the colorimeter.
26. Insert the sample tube into the chamber. Select SCAN SAMPLE. Record the result as Total Nitrogen in mg/L N.
27. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For greater accuracy, use laboratory grade pipets.

To order reagent refills, Order Code R-4026.

OXYGEN SCAVENGERS

DEHA (Diethylhydroxylamine), Carbohydrazide, Erythorbic Acid, Hydroquinone, Mehtylethylketoxime

IRON REDUCTION METHOD • CODE 4857

QUANTITY	CONTENTS	CODE
15 mL	*DEHA Reagent #1	*4791-E
15 mL	*DEHA Reagent #2	*4792-E
15 mL	*DEHA Reagent #3	*4793-E

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Oxygen can lead to corrosion in many parts of a boiler. Oxygen scavengers are added to the water to eliminate oxygen and thus decrease the chance of corrosion. Diethylhydroxylamine (DEHA) is a volatile oxygen scavenger used in boilers. It can also passivate steel and has a low toxicity.

APPLICATION: Boilers

RANGE: 0.000–0.700 ppm DEHA (Diethylhydroxylamine)
0.000–0.900 ppm Carbohydrazide
0.00–3.00 ppm Erythorbic Acid
0.00–2.00 ppm Hydroquinone
0.00–3.00 ppm Methylethylketoxime

METHOD: Ferric iron is reduced to ferrous iron by oxygen scavengers in proportion to the concentration in the sample. The resulting ferrous iron reacts with an indicator to produce a purple color.

SAMPLE HANDLING & PRESERVATION: Analyze samples immediately. Rinse sample containers and glassware with 1:1 hydrochloric acid to avoid iron contamination.

INTERFERENCES: Other oxygen scavengers, such as DEHA, carbohydrazide, erythorbic acid, hydroquinone and methylethylketoxime will interfere. Stray light and substances which complex iron or reduce ferric iron will also interfere.

DEHA PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `38 DEHA` from `TESTING MENU`.
5. Scroll to and select `38 DEHA` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add 3 drops of *DEHA Reagent #1 (4791). Swirl to mix.
10. Add 3 drops of *DEHA Reagent #2 (4792). Swirl to mix.
11. Add 3 drops of *DEHA Reagent #3 (4793). Invert 3 times to mix.
12. Insert the tube into chamber. Close lid.
13. Wait 15 minutes. Do not open the lid during the waiting time. The reaction is photosensitive.
14. Remove tube from chamber. Invert 2 times to mix.
15. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Read within 30 seconds. Record result in ppm DEHA.
16. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

CARBOHYDRAZIDE PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `14 c-hydrazide` from `TESTING MENU`.
5. Scroll to and select `14 c-hydrazide` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add 3 drops of *DEHA Reagent #1 (4791). Swirl to mix.
10. Add 3 drops of *DEHA Reagent #2 (4792). Swirl to mix.
11. Add 3 drops of *DEHA Reagent #3 (4793). Invert 3 times to mix.
12. Insert the tube into chamber. Close lid.
13. Wait 15 minutes. Do not open the lid during the waiting time. The reaction is photosensitive.
14. Remove tube from chamber. Invert 2 times to mix.
15. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Read within 30 seconds. Record result in ppm carbohydrazide.
16. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

ERYTHORBIC ACID PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `40 E-thorbic A` from `TESTING MENU`.
5. Scroll to and select `40 E-thorbic A` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add 3 drops of *DEHA Reagent #1 (4791). Swirl to mix.
10. Add 3 drops of *DEHA Reagent #2 (4792). Swirl to mix.
11. Add 3 drops of *DEHA Reagent #3 (4793). Invert 3 times to mix.
12. Insert the tube into chamber. Close lid.
13. Wait 15 minutes. Do not open the lid during the waiting time. The reaction is photosensitive.
14. Remove tube from chamber. Invert 2 times to mix.
15. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Read within 30 seconds. Record result in ppm erythorbic acid.
16. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

HYDROQUINONE PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `49 H-quinone` from `TESTING MENU`.
5. Scroll to and select `49 H-quinone` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add 3 drops of *DEHA Reagent #1 (4791). Swirl to mix.
10. Add 3 drops of *DEHA Reagent #2 (4792). Swirl to mix.
11. Add 3 drops of *DEHA Reagent #3 (4793). Invert 3 times to mix.
12. Insert the tube into chamber. Close lid.
13. Wait 15 minutes. Do not open the lid during the waiting time. The reaction is photosensitive.
14. Remove tube from chamber. Invert 2 times to mix.
15. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Read within 30 seconds. Record result in ppm hydroquinone.
16. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

METHYLETHYLKETOXIME PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `58 m-e-ketoxim` from `TESTING MENU`.
5. Scroll to and select `58 m-e-ketoxim` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Add 3 drops of *DEHA Reagent #1 (4791). Swirl to mix.
10. Add 3 drops of *DEHA Reagent #2 (4792). Swirl to mix.
11. Add 3 drops of *DEHA Reagent #3 (4793). Invert 3 times to mix.
12. Insert the tube into chamber. Close lid.
13. Wait 15 minutes. Do not open the lid during the waiting time. The reaction is photosensitive.
14. Remove tube from chamber. Invert 2 times to mix.
15. Immediately insert tube into chamber, close lid and select `SCAN SAMPLE`. Read within 30 seconds. Record result in ppm methylethylketoxime.
16. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

OZONE

DPD METHOD • CODE 4881

QUANTITY	CONTENTS	CODE
30 mL	DPD #1A Free Chlorine Reagent	P-6740-G
30 mL	*DPD #1B Free Chlorine Reagent	*P-6741-G
30 mL	*DPD #3 Total Chlorine Reagent	*P-6743-G

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact LaMotte by email, phone or fax.

Note: The primary use for this kit is in applications that use only ozone and no other oxidizing disinfectants.

Ozone is sometimes used in place of, or in conjunction with, chlorine or other halogens for disinfection of pool, spa or drinking waters. Recently, large aquatic facilities have begun using ozone as a disinfectant in many artificial habitats.

APPLICATION: Bottled water, aquatic waters, and non-chlorinated waters.

RANGE: 0.00 – 3.00 mg/L Ozone

METHOD: In the presence of iodide, ozone reacts instantly with the buffered diethyl-p-phenylenediamine indicator (DPD) to produce a red color in proportion to the amount of ozone present.

SAMPLE HANDLING & PRESERVATION: Ozone in aqueous solutions, particularly weak solutions, is not stable. Exposure to sunlight or agitation will accelerate the reduction of ozone. Fill sample containers to the top and cap tightly. Analyze samples as soon as possible after collection.

INTERFERENCES: Interferences are other oxidizers, such as, chlorine, bromine, iodine, and oxidized manganese. The DPD reagent system used in this kit has a significant interference from chlorine. The primary use for this kit is in applications that use only ozone and not other oxidizing disinfectants in conjunction with ozone.

PROCEDURE

1. Press and hold ON button until colorimeter turns on..
2. Press ENTER to start.
3. Press ENTER to select Testing Menu.
4. Select ALL TESTS (or another sequence containing 73. Ozone DPD) from TESTING MENU.
5. Scroll to and select 73. Ozone DPD from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL with sample.
7. Insert the tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter.
9. Add 5 drops *DPD #3 Total Chlorine Reagent (6743). Swirl to mix.
10. Add 5 drops DPD #1 A Free Chlorine Reagent (6740) and 5 drops *DPD #1B Free Chlorine Reagent (6741).
11. Cap and invert to mix. Make reading within 30 seconds.
12. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result as mg/L ozone.
13. Press OFF button to turn the meter off or press EXIT button to exit to a previous menu or make another menu selection

☑ NOTES: For best possible results, a blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

OZONE

INDIGO METHOD • CODE 365I-SC

QUANTITY	CONTENTS	CODE
15 mL	Chlorine Inhibitor	3990-E
250 mL	*Ozone Buffer	*3991-K
30 mL	Indigo Blue Stock Solution	3989-G
1	Sampling Apparatus	0681
1	Pipet, transfer, 1.0 mL	2-2170
1	Pipet, transfer, 5 mL	0329
1	Pump, 10 mL	30527
1	Bottle, HR Reagent, amber glass	0680-J
1	Graduated Cylinder, 50 mL, glass	0418

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Ozone is sometimes used in place of, or in conjunction with, chlorine or other halogens for disinfection of pool, spa, or drinking waters. Recently, large aquatic facilities have begun using ozone as a disinfectant in many artificial habitats.

APPLICATION: Drinking, pool and aquatic waters.

RANGE: 0.00–0.40 ppm Ozone, Low Range
0.00–2.50 ppm Ozone, High Range

METHOD: Ozone rapidly and stoichiometrically decolorizes Indigo Trisulfonate under acidic conditions.

SAMPLE HANDLING & PRESERVATION: Ozone is extremely unstable in aqueous solutions. Test must be performed immediately and the sample must not be agitated.

INTERFERENCES: Manganese at any level interferes.

PROCEDURE–LOW RANGE

A. PREPARATION OF HR REAGENT

☑ NOTE: The quantity of Indigo Blue Stock solution (3989) supplied will prepare one batch of HR Reagent for the High Range Ozone procedure or five batches of HR Reagent for the Low Range Ozone procedure.

1. Use the 50 mL graduated cylinder to carefully add 45 mL of *Ozone Buffer (3991) to amber glass bottle marked HR Reagent (0680).
2. Use the 5 mL transfer pipet (0329) and pump (30527) to add 5 mL of Indigo Blue Stock Solution (3989) to the amber glass bottle. Cap and mix.

B. DETERMINATION OF OZONE

3. Use the 1.0 mL transfer pipet (2-2170) and pump (30527) to add 1.0 mL of HR Reagent to each of 2 clean tubes (0290).
4. If chlorine is present add 3 drops Chlorine Inhibitor (3990) to each tube. Cap tubes.
5. Take one of the prepared tubes (0290) and sampling apparatus (0681) to sampling site.
6. Lower end of tubing of sampling apparatus to desired depth. Slowly withdraw and depress plunger several times to purge syringe and tubing. Slowly withdraw plunger to fill purged syringe.
7. Remove plastic tubing from syringe. Remove cap from the prepared tube. Place tip of syringe against inside of the prepared tube. Slowly depress plunger and fill to the 10 mL line and cap. This is the Sample Tube.

   ☑ **NOTE: DO NOT SHAKE OR INVERT THE SAMPLE.**
8. Fill the second prepared tube (0290) to the 10 mL line with ozone free water. This is the Reagent Blank.
9. Press and hold **ON** button until colorimeter turns on.
10. Press **ENTER** to start.
11. Press **ENTER** to select `TESTING MENU`.
12. Select `ALL TESTS` (or another sequence containing `71 Ozone-LR`) from `TESTING MENU`.
13. Scroll to and select `71 Ozone-LR` from menu.
14. Insert the **Reagent Blank** tube into chamber, close lid and select `SCAN BLANK`.
15. Insert reacted **Sample Tube** into chamber, close lid and select `SCAN SAMPLE`. Record result.
16. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: HR Reagent must be made fresh **each week**. If reagent is refrigerated, it may be kept up to 3 weeks.

PROCEDURE–HIGH RANGE

A. PREPARATION OF HR REAGENT

☑ NOTE: The quantity of Indigo Blue Stock solution (3989) supplied will prepare one batch of HR Reagent for the High Range Ozone procedure or five batches of HR Reagent for the Low Range Ozone procedure.

1. Use the 50 mL graduated cylinder to carefully add 25 mL of *Ozone Buffer (3991) to amber glass bottle marked HR Reagent (0680).
2. Use the 50 mL graduated cylinder to carefully add 25 mL of Indigo Blue Stock Solution (3989) to the amber glass bottle. Cap and mix.

B. DETERMINATION OF OZONE

3. Use the 1.0 mL transfer pipet (2-2170) and pump (30527) to add 1.0 mL of HR Reagent to each of 2 clean tubes (0290).
4. If chlorine is present add 3 drops Chlorine Inhibitor (3990) to each tube. Cap tubes.
5. Take one of the prepared tubes (0290) and sampling apparatus (0681) to sampling site.
6. Lower end of tubing of sampling apparatus to desired depth. Slowly withdraw and depress plunger several times to purge syringe and tubing. Slowly withdraw plunger to fill purged syringe.
7. Remove plastic tubing from syringe. Remove cap from the prepared tube. Place tip of syringe against inside of the prepared tube. Slowly depress plunger and fill to the 10 mL line and cap. This is the Sample Tube.

   ☑ **NOTE: DO NOT SHAKE OR INVERT THE SAMPLE.**
8. Fill the second prepared tube (0290) to the 10 mL line with ozone free water. This is the Reagent Blank.
9. Press and hold **ON** button until colorimeter turns on.
10. Press **ENTER** to start.
11. Press **ENTER** to select `TESTING MENU`.
12. Select `ALL TESTS` (or another sequence containing `72 Ozone-HR`) from `TESTING MENU`.
13. Scroll to and select `72 Ozone-HR` from menu.
14. Insert the **Reagent Blank** tube into chamber, close lid and select `SCAN BLANK`.
15. Insert reacted **Sample Tube** into chamber, close lid and select `SCAN SAMPLE`. Record result.
16. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: HR Reagent must be made fresh **each week**. If reagent is refrigerated, it may be kept up to 3 weeks.

pH

COLORIMETRIC METHOD • CODE 3700-SC

QUANTITY	CONTENTS	CODE
60 mL	Chlorphenol Red Indicator	V-2209-H
60 mL	Phenol Red Indicator	V-2304-H
60 mL	Thymol Blue Indicator	V-2213-H
3	Pipets, 0.5 mL, plastic w/caps	0369

The term pH (always written with a lower case p and an upper case H) is correctly defined as the negative logarithm of the hydrogen ion concentration. More simply, the term pH can be considered to be an index of the amount of hydrogen ion present in a substance, or is a measure of the acidity of the substance. This index is important as it can be used to quickly identify the acid, neutral or alkaline (basic) nature of materials. Acidic substances have a pH less than 7.0, neutral substances have a pH equal to 7.0 and alkaline substances have a pH greater than 7.0.

Most natural waters have pH values from pH 5.0 to pH 8.5. Acidic, freshly fallen rain water may have a pH value of pH 5.5 to pH 6.0. When it reacts with soils and minerals containing weakly alkaline materials, the hydroxyl ion concentration will increase and the hydrogen ion concentration will decrease. Then the water may become slightly alkaline with a pH of 8.0 to 8.5. Natural sea water has a pH value of 8.1, and changes from this value indicate that water from an inland source is entering the body of sea water.

Waters more acidic than pH 5.0 and more alkaline than pH 8.5 to 9.0 should be viewed with suspicion. Mine drainage and acidic industrial wastes are the principal factors in increasing the acidity of water, and alkaline industrial wastes are the cause of high pH values.

Because pH measurements can be made so simply, and because they can tell so much about the past and future reactions of water, they are routinely made in water quality studies. Sudden changes in pH values serve as warning signals that water quality may be adversely affected through the introduction of contaminants.

APPLICATION: Drinking, surface, and saline waters, swimming pool water; domestic and industrial wastes.

METHOD: The various pH indicators exhibit a specific color change over a narrow pH range. The color changes are measured colorimetrically.

SAMPLE HANDLING & PRESERVATION: Sample should be analyzed immediately after collection.

INTERFERENCES: Sample color and turbidity interfere with the colorimetric pH measurement. Color interference may be removed bystandardizing the instrument with the original water sample. Two drops of 0.1N sodium thiosulfate per 100 mL of sample will eliminate chlorine interference.

INDICATOR, RANGE, & TEST NAME:

pH Indicator	pH	Smart2 Test Name
Chlorphenol Red	5.0-6.8	74 pH CPR
Phenol Red	6.6-8.4	75 pH PR
Thymol Blue	8.0-9.6	76 pH TB

PROCEDURE

1. Use *Indicator, Range, & Test Name* chart to select the indicator, corresponding to anticipated pH range and to determine corresponding test name to select from colorimeter menu.
2. Press and hold **ON** button until colorimeter turns on.
3. Press **ENTER** to start.
4. Press **ENTER** to select `TESTING MENU`.
5. Select `ALL TESTS` (or another sequence containing the appropriate pH test name) from `TESTING MENU`.
6. Scroll to and select the appropriate pH test name from menu.
7. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
8. Insert tube into chamber, close lid and select `SCAN BLANK`.
9. Remove tube from colorimeter. Use the 0.5 mL pipet (0369) to add exactly 0.5 mL of the pH indicator for the chosen range. Cap and mix.
10. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

PHENOL

AMINOANTIPYRINE METHOD • CODE 3652-SC

QUANTITY	CONTENTS	CODE
5 g	Aminoantipyrine Reagent	7825-C
30 mL	*Ammonium Hydroxide Solution	*7826-G
2 x 60 mL	*Potassium Ferricyanide Solution	*7827-H
1	Spoon, 0.1 g, plastic	0699
1	Pipet, plain, plastic	0352
1	Pipet, 1.0 mL, plastic	0354

***WARNING:** Reagents marked with a * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact LaMotte by email, phone or fax.

Phenols may occur in domestic and industrial waste waters and in drinking water supplies. Chlorination of waters containing phenols may produce odiferous and objectionable tasting chlorophenols. Natural waters sedom contain more than 1 mg/L phenol.

APPLICATION: Drinking and surface waters; domestic and industrial waste water.

RANGE: 0.00–6.00 ppm Phenol

METHOD: 4-Aminoantipyrine is oxidized in the presence of all ortho- and meta- substituted phenols to form a colored complex in proportion to the amount of phenol present.

SAMPLE HANDLING & PRESERVATION: Phenols are subject to biological and chemical oxidation. Samples should be analyzed within 4 hours after collection. If sample cannot be analyzed within 4 hours, it can be preserved by acidification with phosphoric acid to pH 4.0.

INTERFERENCES: Oxidizing and reducing chemicals, alkaline pH values, and phenol decomposing bacteria may interfere with the test.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `77 Phenol`) from `TESTING MENU`.
5. Scroll to and select `77 Phenol` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Use the 0.1 g spoon (0699) to add one measure of Aminoantipyrine Reagent (7825-C). Cap and mix.
9. Use the plain pipet (0352) to add 4 drops of *Ammonium Hydroxide Solution (7826). Cap and mix.
10. Use the 1 mL pipet (0354) to add 2 mL of *Potassium Ferricyanide Solution (7827). Cap and mix. Solution will almost immediately develop a reddish hue if phenols are present.
11. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

PHOSPHATE–LOW RANGE

ASCORBIC ACID REDUCTION METHOD • CODE 3653-SC

QUANTITY	CONTENTS	CODE
60 mL	*Phosphate Acid Reagent	*V-6282-H
5 g	*Phosphate Reducing Reagent	*V-6283-C
1	Pipet, 1 mL, plastic	0354
1	Spoon, 0.1 g, plastic	0699

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Phosphorus is an important nutrient for aquatic plants. The amount found in water is generally not more than 0.1 ppm unless the water has become polluted from waste water sources or excessive drainage from agricultural areas. When phosphorus is present in excess of the concentrations required for normal aquatic plant growth, a process called eutrophication takes place. This creates a favorable environment for the increase in algae and weeds. When algae cells die, oxygen is used in the decomposition and fish kills often result. Rapid decomposition of dense algae scums with associated organisms give rise to foul odors and hydrogen sulfide gas.

APPLICATION: Drinking, surface and saline waters; domestic and industrial wastes (Method based on reactions that are specific for orthophosphate).

RANGE: 0.00–3.00 ppm Orthophosphate

METHOD: Ammonium molybdate and antimony potassium tartrate react in a filtered acid medium with dilute solution of PO_4^{-3} to form an antimony-phosphomolybdate complex. This complex is reduced to an intense blue colored complex by ascorbic acid. The color is proportional to the amount of phosphate present. (Only orthophosphate forms a blue color in this test.) Polyphosphates (and some organic phosphorus compounds) may be converted to the orthophosphate form by sulfuric acid digestion. Organic phosphorus compounds may be converted to the orthophosphate form by persulfate digestion.

SAMPLE HANDLING & PRESERVATION: If benthic deposits are present in the area being sampled, great care should be taken not to include these deposits. If the analysis cannot be performed the same day of collection, the sample should be preserved by the addition of 2 mL of concentrated sulfuric acid or 40 mg mercuric chloride per liter and refrigerated at 4°C.

INTERFERENCES:

a. No interference from copper, iron, or silicate at concentrations many times the concentration of sea water. However, high iron concentrations can cause precipitation and subsequent loss of phosphorus.

b. Salt error for samples ranging from 5% to 20% salt content was found to be less than 1%.

c. Mercuric chloride, $HgCl_2$, when used as the preservative, interferes when the chloride levels are low (less than 50 mg/L). This interference is overcome by spiking samples with a minimum of 50 mg/L of sodium chloride.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 78 Phosphate L) from TESTING MENU.
5. Scroll to and select 78 Phosphate L from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Use 1.0 mL pipet (0354) to add 1.0 mL of *Phosphate Acid Reagent (V-6282). Cap and mix.
9. Use the 0.1 g spoon (0699) to add one measure of *Phosphate Reducing Reagent (V-6283). Cap and shake until powder dissolves. Wait 5 minutes for full color development. Solution will turn blue if phosphates are present.
10. At end of 5 minute waiting period, mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

PHOSPHATE–HIGH RANGE

VANADOMOLYBDOPHOSPHORIC ACID METHOD
CODE 3655-SC

QUANTITY	CONTENTS	CODE
2 x 30 mL	*VM Phosphate Reagent	*4410-G
1	Pipet, 1.0 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Phosphate treatments in boiler and cooling water and other industrial water systems are run at levels up to 100 ppm orthophosphate. These high levels permit the use of a simpler, high range test.

APPLICATION: Boiler, cooling, and industrial water.

RANGE: 0.0–70.0 ppm Phosphate

METHOD: Orthophosphate reacts in acid conditions with ammonium vanadomolybdate to form vanadomolybdophosphoric acid. This yellow color is proportional to the concentration of orthophosphate and is read colorimetrically.

SAMPLE HANDLING & PRESERVATION: If the analysis cannot be performed the same day of collection, the sample should be preserved by the addition of 2 mL of concentrated sulfuric acid or 40 mg mercuric chloride per liter and refrigerated at 4°C.

INTERFERENCES: Silica interferes only if the sample is heated. Arsenate, fluoride, thorium, bismuth, sulfide, thiosulfate, and thiocyanate cause negative interference.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `79 Phosphate H`) from `TESTING MENU`.
5. Scroll to and select `79 Phosphate H` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Use the 1.0 mL pipet (0354) to add 2.0 mL of *VM Phosphate Reagent (4410). Cap and mix. Wait 5 minutes for full color development.
9. After 5 minute waiting period, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

PHOSPHORUS, TOTAL–LOW RANGE

ASCORBIC ACID REDUCTION WITH PERSULFATE DIGESTION METHOD • CODE 4024

QUANTITY	CONTENTS	CODE
25	*Total Phosphorus Acid Reagent Tubes	*4035-G
5 g	*Digestion Reagent Powder	*4036-C
2 X 30 mL	*Total Phosphorus LR Hydroxide Reagent	*4038-G
2 X 30 mL	*Phosphate Acid Reagent	*V-6282-G
5 g	Phosphate Reducing Reagent	V-6283-C
1	Spoon, 0.15 g, plastic	0727
3	Pipets, 1.0 mL, plastic	0354
1	Spoon, 0.1 g, plastic	0699
2	Funnels, plastic	0459

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

☑ NOTE: For greater accuracy, use laboratory grade pipets.

Equipment needed but not supplied:

1	COD Adapter	5-0087
1	COD Reactor, 12 tubes, 110V	5-0102
or 1	COD Reactor, 12 tubes, 230V	5-0102-EX

Optional Equipment:

1	Volumetric pipet, 5.0 mL	2-2174
2	Volumetric pipets, 1.0 mL	2-2170
1	Pipet Bulb	2-2164
1	Wipes	2-2069
1	Test Tube Rack	23371
1	Timer	9371-W13
1	Test Tube Holder	2-2190

Phosphorus in natural waters and wastewaters occurs almost exclusively in the form of orthophosphates, condensed phosphates (pyro-, meta- and other polyphosphates) and organically bound phosphates. Phosphates may be added in small amounts to water supplies during treatment. Larger amounts are introduced to water used for cleaning or laundering as components of commercial cleaning preparations. Phosphates are used to treat boiler water and are components of agricultural and residential fertilizers. Phosphorus is an important nutrient for aquatic plants. The amount found in natural water is generally not more than 0.1 mg/L unless the water has become polluted from wastewater sources or excessive drainage from agricultural areas.

APPLICATION: Drinking, surface and saline waters; domestic and industrial waste water.

RANGE: 0.00 –3.50 mg/L Total Phosphorus as Phosphate

METHOD: Pretreatment of the sample with heat and acid provides conditions for the hydrolysis of condensed inorganic phosphates. Heat, acid and persulfate convert the organic phosphates to orthophosphate during the digestion. Ammonium molybdate and antimony potassium tartrate react in a filtered acid medium with dilute solutions of phosphate to form an antimony-phosphomolybdate complex. This complex is reduced to an intense blue colored complex by ascorbic acid. The color is proportional to the amount of phosphate present.

SAMPLE HANDLING & PRESERVATION: Rinse sample bottle with 1:1 hydrochloric acid followed by deionized water. Do not use phosphate detergents. If the sample can not be analyzed immediately, the sample should be preserved by adjusting the pH to 2 or less with concentrated sulfuric acid and refrigerated at 4°C. Warm the sample to room temperature and neutralize with 5.0 N sodium hydroxide before analyzing.

INTERFERENCES: Large amounts of turbidity may interfere. Aluminum (200 ppm), Arsenate (any level), Chromium (100 ppm), Copper (10 ppm), Iron (100 ppm), Nickel (300 ppm), Silica (50 ppm), Silicate (10 ppm), Sulfide (90 ppm) and Zinc (80 ppm) will interfere.

PROCEDURE

Use COD adapter.

1. Preheat COD reactor to 150 ±2°C. Follow safety precautions.
2. Remove cap from a *Total Phosphorus Acid Reagent Tube (4035). Use a 1.0 mL pipet (0354) to add 5.0 mL of sample.
3. Use the 0.15 g spoon (0727) and a funnel (0459) to add one level measure of *Digestion Reagent Powder (4036). Tap funnel to dispense powder completely. Cap tube tightly and shake until powder completely dissolves.
4. Place the tube in the COD reactor for 30 minutes.
5. At the end of the heating period, turn the reactor off. Carefully remove the tube from the reactor and allow it to cool to room temperature.
6. At the end of the cooling period, press and hold **ON** button until colorimeter turns on.
7. Press **ENTER** to start.
8. Press **ENTER** to select `TESTING MENU`.
9. Select `ALL TESTS` (or another sequence containing `82 Phos T LR`) from `TESTING MENU`.
10. Scroll to and select `82 Phos T LR` from the menu.
11. Carefully remove the cap from the digested tube. Use another 1 mL pipet (0354) to add 1.0 mL of *Total Phosphorus LR Hydroxide Reagent (4038) to the tube. Cap and invert to mix.
12. Wipe the tube with a damp towel to remove fingerprints and smudges. Wipe with a dry towel.
13. Insert the tube into the chamber. Select `SCAN BLANK`. Remove the tube from the colorimeter.
14. Use another 1 mL pipet (0354) to add *1.0 mL of Phosphate Acid Reagent (V-6282). Cap and invert tube to mix.
15. Use the 0.1g spoon (0699) and a funnel (0459) to add one level spoon of Phosphate Reducing Reagent (V-6283). Tap funnel to dispense powder completely. Cap tube and shake until powder dissolves.
16. Wait 5 minutes.
17. Wipe the vials with a damp towel to remove fingerprints and smudges. Wipe with a dry towel.
18. Insert the tube into the chamber. Select `SCAN SAMPLE`. Record the result as Total Phosphorus in mg/L PO_4.
19. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For greater accuracy, use laboratory grade pipets.
To order reagent refills, Order Code R-4024.

PHOSPHORUS, TOTAL–HIGH RANGE

MOLYBDOVANADATE METHOD WITH ACID PERSULFATE DIGESTION • CODE 4025

QUANTITY	CONTENTS	CODE
25	*Total Phosphorus Acid Reagent Tubes	*4035-G
60 mL	Deionized Water	5115PS-H
5 g	*Digestion Reagent Powder	*4036- C
2 X 30 mL	*Total Phosphorus HR Hydroxide Reagent	*4037-G
30 mL	*Total Phosphorus HR Indicator Reagent	*4039-G
1	Spoon, 0.15 g	0727
3	Pipets 1.0 mL, plastic	0354
1	Pipet, 0.5 mL	0353
1	Funnel, plastic	0459

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

☑ NOTE: For greater accuracy, use laboratory grade pipets.

Equipment needed but not supplied:

1	COD Adapter	5-0087
1	COD Reactor, 8 vial, 110V	5-0069
Or 1	COD Reactor, 8 vial, 220V	5-0070
Or 1	COD Reactor, 25 vial, 115V/230V	5-0094

Optional Equipment:

1	Volumetric pipet, 2.0 mL	2-2168
2	Volumetric pipet, 5.0 mL	2-2174
1	Volumetric pipet, 0.5 mL	30503
1	Pipet Bulb	2-2164
1	Wipes	2-2069
1	Test Tube Rack	23371
1	Timer	9371-W13
1	Test Tube Holder	2-2190

Phosphorus in natural waters and wastewaters occurs almost exclusively in the form of orthophosphates, condensed phosphates (pyro-, meta- and other polyphosphates) and organically bound phosphates. Phosphates may be added in small amounts to water supplies during treatment. Larger amounts are introduced to water used for cleaning or laundering, as components of commercial cleaning preparations. Phosphates are used to treat boiler water and are components of agricultural and residential fertilizers. Phosphorus is an important nutrient for aquatic plants. The amount found in natural water is generally not more than 0.1 mg/L unless the water has become polluted from wastewater sources or excessive drainage from agricultural areas.

APPLICATION: Boiler, cooling, and industrial water.

RANGE: 0.0–100.0 mg/L Total Phosphorus as phosphate

METHOD: Pretreatment of the sample with heat and acid provides conditions for the hydrolysis of condensed inorganic phosphates. Heat, acid and persulfate convert the organic phosphates to orthophosphate during digestion. Orthophosphate reacts in acidic conditions with ammonium vanadomolybdate to form vanadomolybdophosphoric acid. The resulting yellow color is proportional to the concentration of orthophosphate.

SAMPLE HANDLING & PRESERVATION: Rinse sample bottle with 1:1 hydrochloric acid followed by deionized water. Do not use phosphate detergents. If the sample can not be analyzed immediately, the sample should be preserved by adjusting the pH to 2 or less with concentrated sulfuric acid and refrigerated at 4°C. Warm the sample to room temperature and neutralize with 5.0 N sodium hydroxide before analyzing.

INTERFERENCES: Large amounts of turbidity may interfere. Silica and arsenate interfere only if the sample is heated. Arsenite, fluoride, thorium, bismuth, molybdate, thiosulfate, and thiocyanate cause negative interference. Ferrous iron concentrations above 100 ppm will interfere.

PROCEDURE

Use COD adapter.

1. Preheat COD reactor to 150 ±2°C. Follow safety precautions.
2. Remove cap from a *Total Phosphorus Acid Reagent Tube (4035). Use a 1.0 mL pipet (0354) to add 5.0 mL of Deionized Water (5115PS). This is the blank.
3. Remove cap from a *Total Phosphorus Acid Reagent Tube (4035). Use the 1.0 mL pipet (0354) to add 5.0 mL of sample water. This is the sample.
4. Use the 0.15 g spoon (0727) and a funnel (0459) to add one level measure of *Digestion Reagent Powder (4036) to each tube. Tap funnel to dispense powder completely. Cap tube tightly and shake until powder dissolves completely.
5. Place the tubes in the COD reactor for 30 minutes.
6. At the end of the heating period, turn the reactor off. Carefully remove the tubes from the reactor block and allow them to cool to room temperature.
7. Carefully remove the caps from the digested tubes. Use another 1 mL pipet (0354) to add 2.0 mL of *Total Phosphorus HR Hydroxide Reagent (4037) to each tube. Cap and invert to mix.
8. Use the 0.5 mL pipet (0353) to add 0.5 mL *Total Phosphorus HR Indicator Reagent (4039) to each tube. Cap and invert to mix. Wait 7 minutes.
9. During the waiting period, press and hold **ON** button until colorimeter turns on.
10. Press **ENTER** to start.
11. Press **ENTER** to select `TESTING MENU`.
12. Select `ALL TESTS` (or another sequence containing `83 Phos T HR`) from `TESTING MENU`.
13. Scroll to and select `83 Phos T HR` from the menu.
14. Wipe the tubes with a damp towel to remove fingerprints and smudges. Wipe with a dry towel.
15. Insert the blank tube into the chamber. Select `SCAN BLANK`. Remove the blank tube from the colorimeter.
16. Insert the sample tube into the chamber. Select `SCAN SAMPLE`. Record the result as Total Phosphorus in mg/L PO_4.
17. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For greater accuracy, use laboratory grade pipets.
To order reagent refills, order code R-4025.

POTASSIUM

TETRAPHENYLBORON METHOD • CODE 3639-SC

QUANTITY	CONTENTS	CODE
30 mL	*Sodium Hydroxide, 1.0N	*4004WT-G
5 g	*Tetraphenylboron Powder	*6364-C
1	Spoon, 0.05 g, plastic	0696

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Potassium, as the seventh most common element on the Earth, may be found in minor quantities in most water supplies. It seldom exceeds 10 ppm in drinking water and usually is less than 2 ppm. In some brine or runoff in agricultural areas the potassium concentration may reach 100 ppm.

APPLICATION: Drinking, surface, and saline water.

RANGE: 0.0–10.0 ppm Potassium

METHOD: Potassium reacts with sodium tetraphenylborate to form a colloidal white precipitate in quantities proportional to the potassium concentration.

SAMPLE HANDLING & PRESERVATION: Store samples in polyethylene bottles, not in soft glass where leaching of potassium from the glass may occur. Samples may be acidified to pH 2 with nitric acid, but should be neutralized before analyzing.

INTERFERENCE: Calcium and magnesium interfere at very high concentrations. Check for stray light interference (see p. 17).

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `81 Potassium`) from `TESTING MENU`.
5. Scroll to and select `81 Potassium` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Add 4 drops of *Sodium Hydroxide, 1.0N (4004WT). Cap and mix.
9. Use the 0.05 g spoon (0696) to add one measure of *Tetraphenylboron Powder (6364). Cap and shake vigorously until all of the powder has dissolved. Wait 5 minutes.
10. At end of 5 minute waiting period, mix tube again to suspend any settled precipitate. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

For the most accurate results, the sample and reagents should be at 25±4°C.

SILICA–LOW RANGE

HETEROPOLY BLUE METHOD • CODE 3664-SC

QUANTITY	CONTENTS	CODE
30 mL	*Silica Reagent #1	*V-4466-G
30 mL	*Silica Reagent #2	*V-4467-G
30 mL	*Silica Reagent #3	*V-4468-G
10 g	*Silica Reagent #4	*V-6284-D
1	Spoon, 0.1 g, plastic	0699

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Silicon dioxide, SiO_2, commonly known as silica, occurs in all natural water. Silica may be present as suspended, insoluble particles in a colloidal or polymeric state. It may also be present in a reactive form as silicic acid or silicate ions. Silica is a major nutrient for diatoms. A silica cycle occurs in many bodies of water containing organisms, such as diatoms, that use silica in their skeletal structure. The silica removed from the water may be slowly returned to solution by the decomposition of the dead organisms. The major source of silica in natural water is from the decomposition of silicate minerals in the drainage basin from which the waters flow.

The presence of silica is particularly objectionable in water used for boiler feed water purposes, as it may cause the formation of a hard, dense scale which has unusually high resistance to heat transfer. Serious loss of turbine efficiency results from insoluble silica turbine blade deposits caused by vaporization of silica from boiler water.

APPLICATION: Drinking, surface and saline waters; domestic and industrial wastes.

RANGE: 0.0–4.0 ppm Silica

METHOD: Reactive silica forms a complex with ammonium molybdate in an acidic solution to produce a yellow-green color in proportion to the amount of silica present. Phosphate also reacts with molybdate but the addition of oxalic acid eliminates the molybdophosphoric acid complex. This silica molybdate complex is then reduced by ascorbic acid to produce an intense blue color.

SAMPLE HANDLING & PRESERVATION: Silica samples may be preserved by refrigeration at 4°C in plastic containers up to one week without any change in silica concentration.

INTERFERENCES: Sulfides and large amounts of iron interfere. Color and turbidity may be removed by standardizing the instrument with the original water sample. Since silica is a component of glass waste and a common contaminant, it is suggested to run a reagent blank using silica-free water. The blank value is subtracted from the sample concentrations.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 85 Silica Lo) from TESTING MENU.
5. Scroll to and select 85 Silica Lo from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK. (See Note)
8. Remove tube from colorimeter. Add 6 drops *Silica Reagent #1 (V-4466). Cap and invert to mix.
9. Add 12 drops of *Silica Reagent #2 (V-4467). Cap and mix. Wait 5 minutes.
10. Add 8 drops of *Silica Reagent #3 (V-4468). Cap and mix. Wait 2 minutes.
11. Use the 0.1 g spoon (0699) to add one measure of *Silica Reagent #4 (V-6284). Cap and mix gently until powder has dissolved. Wait 5 minutes for full color development.
12. At end of 5 minute waiting period, mix, insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
13. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

SILICA–HIGH RANGE

SILICOMOLYBDATE METHOD • CODE 3687-SC

QUANTITY	CONTENTS	CODE
30 mL	*Silica Reagent #1	*V-4466-G
30 mL	*Silica Reagent #2	*V-4467-G
15 mL	*Silica Reagent #3	*V-4468-G

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Silicon dioxide, SiO_2, commonly known as silica, occurs in all natural water. Silica may be present as suspended, insoluble particles in a colloidal or polymeric state. It may also be present in a reactive form as silicic acid or silicate ions. Silica is a major nutrient for diatoms. A silica cycle occurs in many bodies of water containing organisms, such as diatoms, that use silica in their skeletal structure. The silica removed from the water may be slowly returned to solution by the decomposition of the dead organisms. The major source of silica in natural water is from the decomposition of silicate minerals in the drainage basin from which the waters flow.

The presence of silica is particularly objectionable in water used for boiler feed water purposes, as it may cause the formation of a hard, dense scale which has unusually high resistance to heat transfer. Serious loss of turbine efficiency results from insoluble silica turbine blade deposits caused by vaporization of silica from boiler water.

APPLICATION: Boilers and cooling towers; domestic and industrial wastes.

RANGE: 0–75 ppm Silica

METHOD: Silica forms a complex with ammonium molybdate in an acidic solution to produce a yellow color in proportion to the amount of silica present. Phosphate also reacts with molybdate but the addition of oxalic acid eliminates the molybdophosphoric acid complex.

SAMPLE HANDLING & PRESERVATION: Silica samples may be preserved by refrigeration at 4°C in plastic containers up to one week without any change in silica concentration.

INTERFERENCES: Sulfides and large amounts of iron interfere. Color and turbidity may be removed by standardizing the instrument with the original water sample.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `86 Silica Hi`) from `TESTING MENU`.
5. Scroll to and select `86 Silica Hi` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Add 6 drops *Silica Reagent #1 (V-4466). Cap and invert to mix.
9. Add 12 drops of *Silica Reagent #2 (V-4467). Cap and mix. Wait 5 minutes.
10. At end of 5 minute waiting period, add 8 drops of *Silica Reagent #3 (V-4468). Cap and mix.
11. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: To extend the range to 100 ppm, perform a 2:1 dilution of water sample, with silica-free water. Perform test and multiply result by 2.

SULFATE–HIGH RANGE

BARIUM CHLORIDE METHOD • CODE 3665-SC

QUANTITY	CONTENTS	CODE
10 g	*Sulfate Reagent	*V-6277-D
1	Spoon, 0.1 g, plastic	0699

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

The most common mineral forms of sulfur are iron sulfide, lead sulfide, zinc sulfide and as calcium sulfate and magnesium sulfate. In most fresh waters the sulfate ion is the second or third most abundant anion, being exceeded only by bicarbonate and, in some cases, silicate. Sulfur, in the form of sulfate, is considered an important nutrient element. Mineral springs are rich in sulfate and feed appreciable quantities of this compound to the watershed. Acid mine water drainage is a form of pollution which may contribute extremely large amounts of sulfate content to natural waters. Other sources of sulfate include waste material from pulp mills, steel mills, food processing operations and municipal wastes. Many bacteria obtain sulfur from sulfate for the synthesis of amino acids. In lakes and streams low in oxygen, this process of sulfate reduction causes the production of hydrogen sulfide, with its characteristic offensive odor. Calcium sulfate and magnesium sulfate contribute significantly to the hardness of water. Under natural conditions, the quantities ordinarily to be expected in lakes are between 3 and 30 parts per million.

APPLICATION: Drinking and surface waters, domestic and industrial wastes.

RANGE: 0–100 ppm Sulfate

METHOD: Sulfate ion is precipitated in an acid medium with barium chloride to form a barium sulfate suspension in proportion to the amount of sulfate present.

SAMPLE HANDLING & PRESERVATION: Sulfate samples may be preserved by refrigeration at 4°C up to 7 days in glass or plastic containers without any change in concentration.

INTERFERENCE: Suspended matter and color interference may be removed by a filtration step. Silica in excess of 500 mg/L will interfere. Check for stray light interference (see page 17).

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `89 Sulfate-HR`) from `TESTING MENU`.
5. Scroll to and select `89 Sulfate-HR` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Use the 0.1 g spoon (0699) to add one measure of *Sulfate Reagent (V-6277). Cap and shake until powder dissolves. A white precipitate will develop if sulfates are present. Wait 5 minutes.
9. Mix tube again. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: If the sulfate concentration of the test sample is greater than 100 ppm, it is recommended that a dilution be made with deionized water and the results multiplied by the dilution factor.

A white film is deposited on the inside of test tubes as a result of the sulfate test. Thoroughly clean and rinse test tubes after each test.

For the most accurate results, samples and reactions should be at 25±4°C.

SULFIDE–LOW RANGE

METHYLENE BLUE METHOD • CODE 3654-01-SC

QUANTITY	CONTENTS	CODE
2 x 30	*Sulfide Reagent A	*V-4458-G
15 mL	*Sulfide Reagent B	*V-4459-E
2 x 60 mL	Sulfide Reagent C	4460-H
2	Pipets, 1.0 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Sulfide occurs in many well water supplies and sometimes is formed in lakes or surface waters. In distribution systems, it may be formed as a result of bacterial action on organic matter under anaerobic conditions. It may also be found in waters receiving sewage or industrial wastes. Lake muds rich in sulfates produce hydrogen sulfide during periods of very low oxygen levels that result from stagnation. Concentrations of a few hundredths of a part per million (or milligram per liter) cause a noticeable odor. At low concentrations, this odor is described as "musty"; at high concentration, as "rotten eggs." Removal of sulfide odor is accomplished by aeration or chlorination. Hydrogen sulfide, a toxic substance, acts as a respiratory depressant in both humans and fish.

APPLICATION: Drinking, surface and saline waters; domestic and industrial wastes.

RANGE: 0.00–1.50 ppm Sulfide

METHOD: Under suitable conditions the sulfide ion reacts with p-aminodimethylaniline and ferric chloride to produce methylene blue in proportion to the sulfide concentration. Ammonium phosphate is added to remove the color due to the ferric iron.

SAMPLE HANDLING & PRESERVATION: Samples must be taken with a minimum of aeration since sulfide is volatilized by aeration and any oxygen which is taken up will destroy sulfides by chemical action. Samples that are used for total sulfide concentrations may be preserved by adding 2M zinc acetate solution at a dosage of 2 mL per liter of sample. This precipitates sulfide as inert zinc sulfide. Determination of dissolved sulfides in samples not preserved with zinc acetate must be started within 3 minutes of sampling.

INTERFERENCES: Strong reducing agents such as sulfite, thiosulfate, and hydrosulfite prevent the formation of the color or diminish its intensity. High concentrations of sulfide will inhibit the reaction, but dilution of the sample prior to analysis eliminates this problem.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 90 Sulfide-LR) from TESTING MENU.
5. Scroll to and select 90 Sulfide-LR from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK.
8. Remove tube from colorimeter. Use the 1.0 mL pipet (0354) to add 1.0 mL of *Sulfide Reagent A (V-4458). Cap and mix.
9. Add 6 drops of Sulfide Reagent B (V-4459). Cap and mix. Wait 1 minute. Solution will turn blue if sulfides are present.
10. Use the 1.0 mL pipet (0354) to add 2.0 mL of Sulfide Reagent C (4460). Cap and mix. Color development is immediate and stable.
11. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
12. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

SURFACTANTS

ION PAIR EXTRACTION–BROMPHENOL BLUE INDICATOR CODE 4876

QUANTITY	CONTENTS	CODE
50 g	pH Adjustment Powder	4509- H
10 g	Sodium Chloride Reagent	4877-D
2 X 60 mL	*DS Indicator Reagent	*4508-H
1	Spoon, 0.5 g, plastic	0698
1	Spoon, 0.1 g, plastic	0699
1	Pipet, 1.0 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Aqueous waste from households and industrial laundering operations is the main source of surfactants in waters. Surfactants are found in low concentrations in natural water except in areas of an outfall or other point source.

APPLICATION: Surface water, wastewater.

RANGE: 0.5–8.0 as Linear Alkyl Sulfonates (LAS)

METHOD: The presence of LAS in the water sample causes the transfer of bromphenol blue dye from the organic reagent layer to the aqueous layer. The amount of color in the aqueous layer is proportional to the concentration of the LAS in the sample. LAS are Methylene Blue Active Substances (MBAS). This calibration is based on sodium lauryl sulfate (dodecyl sodium sulfate). Some linear alkyl sulfonates may have a slightly different response. Prepare standards of a known concentration and follow the test procedure below to determine a conversion factor.

SAMPLE HANDLING & PRESERVATION: Analyze samples as soon as possible. May be stored at 4°C for 24 hours. Warm to room temperature before testing.

INTERFERENCES: Cationic surfactants and nonionic surfactants.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `94 Surfactants`) from `TESTING MENU`.
5. Scroll to and select `94 Surfactants` from menu.
6. Rinse a tube (0290) with sample water. Fill to 10 mL line with sample.
7. Insert the tube into chamber, close lid and select `SCAN BLANK`.
8. Remove the tube from colorimeter.
9. Use the 0.5 g spoon (0698) to add 0.5 g pH Adjustment Powder (4509). Cap and mix until powder dissolves.
10. Use the 0.1 g spoon (0699) to add two measures of Sodium Chloride Reagent (4877). Cap and mix until powder disintegrates.
11. Use the 1.0 mL pipet (0354) to add 2.0 mL of *DS Indicator (4508).
12. Cap and shake for 1 minute.
13. Wait 5 minutes. DO NOT MIX.
14. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result in ppm LAS.
15. Press **OFF** button to turn the colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents is obtained.

TANNIN

TUNGSTO-MOLYBDOPHOSPHORIC ACID METHOD
CODE 3666-SC

QUANTITY	CONTENTS	CODE
30 mL	*Tannin Reagent #1	*7833-G
2 x 60 mL	*Tannin Reagent #2	*7834-H
1	Pipet, plain, plastic	0352
1	Pipet, 1.0 mL, plastic	0354

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Tannin and lignin are examples of hydroxylated aromatic compounds found in discharge wastewater from paper mills, in some boiler water treatment, in natural brackish water, and in wastewater from leather tanning plants. The taste and odor of these compounds is generally offensive so that their control is important in many areas.

APPLICATION: Industrial wastewater, boiler water, and natural water.

RANGE: 0.0–10.0 ppm Tannic Acid

METHOD: The hydroxylated aromatic compounds will reduce a mixture of tungstophosphoric and molybdophosphoric acids to form a blue color in proportion to the concentration of aromatic hydroxyl groups.

SAMPLE HANDLING & PRESERVATION: Sample should be analyzed as soon as possible after collection.

INTERFERENCES: Other reducing compounds such as ferrous iron and sulfites. Results may be expressed as tannin like compounds, or aromatic hydroxy compounds.

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `96 Tannin`) from `TESTING MENU`.
5. Scroll to and select `96 Tannin` from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Remove tube from colorimeter. Use the plain pipet (0352) to add 4 drops of *Tannin Reagent #1 (7833). Cap and mix.
9. Use the 1.0 mL pipet (0354) to add 2.0 mL of *Tannin Reagent #2 (7834). Cap and mix. Wait 30 minutes for full color development.
10. At end of 30 minute waiting period, mix, insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result.
11. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTES: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

For the most accurate results, the sample and reagents should be at 20 ±2°C.

TURBIDITY

ABSORPTION METHOD • NO REAGENTS REQUIRED

Turbidity is a measure of water clarity and is independent of color. Turbidity is caused by undissolved and suspended solids. Mud, silt, algae, and microorganisms can all cause turbidity. Turbidity is a gross measurement of water quality.

APPLICATION:	Surface and industrial water for non-compliance monitoring. (For compliance monitoring at low turbidity levels, use a commercial nephelometer.)
RANGE:	0–400
METHOD:	Absorptimetric
SAMPLE HANDLING & PRESERVATION:	Measure sample as soon as possible after collection.
INTERFERENCES:	Check for stray light interference (see page 17).

PROCEDURE

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select `TESTING MENU`.
4. Select `ALL TESTS` (or another sequence containing `98 Turbidity`) from `TESTING MENU`.
5. Scroll to and select `98 Turbidity` from menu.
6. Rinse a clean tube (0290) with deionized water (turbidity free). Fill to the 10 mL line with deionized water.
7. Insert tube into chamber, close lid and select `SCAN BLANK`.
8. Rinse a second clean tube (0290) with sample water. Fill to the 10 mL line with sample. Cap tube. Wipe off excess water and fingerprints. Shake to resuspend particulate matter. Remove all bubbles before measurement.
9. Insert tube into chamber, close lid and select `SCAN SAMPLE`. Record result. Turbidity measurements should be taken as soon as possible after sample has been collected.
10. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For the most accurate results, the sample should be at 25±4°C.

PREPARING FORMAZIN SOLUTIONS

The turbidity calibration was prepared by using standard formazin solutions as a reference. These solutions can be prepared by carefully following the procedure below.†

1. Dissolve 1.000 g of Hydrazine Sulfate in deionized water and dilute to mark in 100 mL volumetric flask.
2. Dissolve 10.00 g of Hexamethylenetetramine in deionized water and dilute to mark in 100 mL volumetric flask.
3. Mix 5 mL of each solution in a 100 mL volumetric flask and allow to set undisturbed for 24 hours.
4. At the end of the waiting period, dilute to mark with deionized water and mix.
5. The turbidity of the stock solution is 400 FTU. The stock solution is stable for one month. Dilutions from the stock should be prepared fresh daily.

†Alternatively, a prepared concentrated formazin standard of 4000 NTU may be ordered in a 60 mL size by Code 6195-H.

ZINC–LOW RANGE

ZINCON METHOD • CODE 3667-SC

QUANTITY	CONTENTS	CODE
30 mL	*Zinc Indicator Solution	*6314-G
120 mL	*Methyl Alcohol	*6319-J
10 g	Sodium Ascorbate Powder	6316-D
25 g	*Zinc Buffer Powder	*6315-G
15 mL	*Sodium Cyanide, 10%	*6565-E
30 mL	*Formaldehyde Solution, 37%	*5128-G
1	"Dilute Zinc Indicator Solution" Bottle, w/1 pipet assembly	0128-MT
1	Graduated Cylinder, 10 mL, glass	0416
1	Spoon, 0.5 g, plastic	0698
2	Pipets, plain, plastic	0352
1	Spoon, 0.1 g, plastic	0699

*WARNING: Reagents marked with an * are considered to be potential health hazards. To view or print a Material Safety Data Sheet (MSDS) for these reagents see MSDS CD or www.lamotte.com. To obtain a printed copy, contact Lamotte by e-mail, phone or fax.

Zinc enters the domestic water supply from the deterioration of galvanized iron and brass pipes, and from industrial wastes. Zinc is an essential element for body growth and development and is an important plant nutrient. Concentrations of zinc above 5.0 mg/L in drinking water can cause a bitter astringent taste. In the U.S., zinc concentrations may vary between 0.06 to 7.0 mg/L, with an average value of 1.33 mg/L.

APPLICATION: Drinking and surface waters, domestic and industrial waste water.

RANGE: 0.00–3.00 ppm Zinc

METHOD: Zinc forms a blue colored complex with Zincon in a solution buffered at pH 9.0. Other heavy metals are complexed by cyanide and the zinc cyanide complex is released by the addition of formaldehyde before the other metal cyanide complexes are destroyed. Sodium ascorbate is added to reduce the interference of manganese.

SAMPLE HANDLING & PRESERVATION: Sample should be analyzed within 6 hours after collection. The addition of hydrochloric acid will help preserve the metal ion content, however the acid should be neutralized before analysis.

INTERFERENCES: The following ions interfere in concentrations greater than those listed.

Ion	mg/L	Ion	mg/L
Cd(II)	1	Cr(III)	10
Al (III)	5	Ni(II)	20
Mn (II)	5	Co (II)	30
Fe (III)	7	CrO4(II)	50
Fe (II)	9		

PROCEDURE

A. PREPARATION OF DILUTE ZINC INDICATOR SOLUTION

1. Use a pipet (0352) to measure exactly 5.0 mL of *Zinc Indicator Solution (6314) into 10 mL graduated cylinder (0416). The bottom of the curved surface (the meniscus) of liquid should be at 5.0 mL mark. Pour this into the bottle labeled "Dilute Zinc Indicator Solution".
2. Use unrinsed graduated cylinder to add 10.0 mL and then 7.8 mL (total of 17.8 mL) of *Methyl Alcohol (6319) to bottle labeled "Dilute Zinc Indicator Solution". Cap and mix ingredients in this bottle. Do not leave this bottle uncapped.

B. DETERMINATION OF ZINC

1. Press and hold **ON** button until colorimeter turns on.
2. Press **ENTER** to start.
3. Press **ENTER** to select TESTING MENU.
4. Select ALL TESTS (or another sequence containing 99 Zinc-LR) from TESTING MENU.
5. Scroll to and select 99 Zinc-LR from menu.
6. Rinse a clean tube (0290) with sample water. Fill to the 10 mL line with sample.
7. Insert tube into chamber, close lid and select SCAN BLANK. (See Note)
8. Remove tube from colorimeter. Use 0.1 g spoon (0699) to add one measure of Sodium Ascorbate Powder (6316). Use 0.5 g spoon (0698) to add one measure of *Zinc Buffer Powder (6315). Cap and shake vigorously for 1 minute. Some undissolved buffer may remain in the bottom of the tube.
9. Add 3 drops of *Sodium Cyanide, 10% (6565). Cap and mix.
10. Use the 1 mL pipet assembly to add 1 mL of "Dilute Zinc Indicator Solution". Cap and mix.
11. Use a second plain pipet (0352) to add 4 drops of *Formaldehyde Solution, 37% (5128). Cap and mix by inverting 15 times.
12. Insert tube into chamber, close lid and select SCAN SAMPLE. Record result.
13. Press **OFF** button to turn colorimeter off or press **EXIT** button to exit to a previous menu or make another menu selection.

☑ NOTE: For best possible results, a reagent blank should be determined to account for any contribution to the test result by the reagent system. To determine the reagent blank, follow the above test procedure to scan a distilled or deionized water blank. Then follow the above procedure to perform the test on a distilled or deionized water sample. This test result is the reagent blank. Subtract the reagent blank from all subsequent test results of unknown samples. It is necessary to determine the reagent blank only when a new lot number of reagents are obtained.

SMART 2

Colorimeter

APPENDIX

LaMotte

APPENDIX

Ammonia in water occurs in two forms: toxic unionized ammonia (NH_3) and the relatively non-toxic ionized form, ammonium ion (NH_4^+). This test method measures both forms as ammonia-nitrogen (NH_3^+–N) to give the total ammonia-nitrogen concentration in water. The actual proportion of each compound depends on temperature, salinity, and pH. A greater concentration of unionized ammonia is present when the pH value and salinity increase.

1. Consult the table below to find the percentage that corresponds to the temperature, pH, and salinity of the sample.
2. To express the test result as ppm Unionized Ammonia Nitrogen (NH_3–N), multiply the total ammonia-nitrogen test result by the percentage from the table.
3. To express the test result as ppm Ammonia Nitrogen (NH_3^+–N), subtract the unionized ammonia-nitrogen determined in step 2 from the total ammonia-nitrogen.

	10°C		15°C		20°C		25°C	
pH	FW[1]	SW[2]	FW	SW	FW	SW	FW	SW
7.0	0.19	—	0.27	—	0.40	—	0.55	—
7.1	0.23	—	0.34	—	0.50	—	0.70	—
7.2	0.29	—	0.43	—	0.63	—	0.88	—
7.3	0.37	—	0.54	—	0.79	—	1.10	—
7.4	0.47	—	0.68	—	0.99	—	1.38	—
7.5	0.59	0.459	0.85	0.665	1.24	0.963	1.73	1.39
7.6	0.74	0.577	1.07	0.836	1.56	1.21	2.17	1.75
7.7	0.92	0.726	1.35	1.05	1.96	1.52	2.72	2.19
7.8	1.16	0.912	1.69	1.32	2.45	1.90	3.39	2.74
7.9	1.46	1.15	2.12	1.66	3.06	2.39	4.24	3.43
8.0	1.83	1.44	2.65	2.07	3.83	2.98	5.28	4.28
8.1	2.29	1.80	3.32	2.60	4.77	3.73	6.55	5.32
8.2	2.86	2.26	4.14	3.25	5.94	4.65	8.11	6.61
8.3	3.58	2.83	5.16	4.06	7.36	5.78	10.00	8.18
8.4	4.46	3.54	6.41	5.05	9.09	7.17	12.27	10.10
8.5	5.55	4.41	7.98	6.28	11.18	8.87	14.97	12.40

[1] Freshwater data from Trussel (1972).

[2] Seawater values from Bower and Bidwell (1978).
Salinity for Seawater values = 34% at an ionic strength of 0.701m.

FOR EXAMPLE:

If a fresh water sample at 20°C has a pH of 8.5 and the test result is 1.0 ppm as Total Ammonia-Nitrogen:

1. The percentage from the table is 11.18% (or 0.1118).
2. 1 ppm Total Ammonia-Nitrogen x 0.1118 = 0.1118 ppm Unionized Ammonia-Nitrogen.
3.

Total Ammonia-Nitrogen		1.0000 ppm
Unionized Ammonia-Nitrogen	−	0.1118 ppm
Ionized Ammonia-Nitrogen	=	0.8882 ppm